cytiva

Glutathione **Sepharose** High Performance

Affinity media

Instructions for Use

Glutathione Sepharose™ High Performance is an affinity chromatography medium designed for easy, one-step purification of glutathione S-transferase (GST) tagged proteins produced using the pGEX series of expression vectors (1), other glutathione S-transferases and glutathione binding proteins.

GST-tagged proteins can be purified directly from pre-treated bacterial lysates using Glutathione Sepharose High Performance. The tagged proteins are eluted under mild, non-denaturing conditions that preserve protein antigenicity and function.

The medium Glutathione Sepharose High Performance is an excellent choice for high performance purifications. Glutathione Sepharose High Performance is also available in convenient prepacked GSTrap™ HP 1-mL and 5-mL columns.

Table 1, on page 3 lists the characteristics of Glutathione Sepharose High Performance.

Detailed information about the production of GST-tagged proteins can be found in the Handbook 18115758 (1).

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1 Description

Medium properties

The glutathione ligand is coupled via a 10-carbon linker to highly crosslinked 6% agarose. The coupling is optimized to give high binding capacity for GST-tagged proteins and other glutathione binding proteins.

The total binding capacity is approximately 10 mg GST-tagged protein/mL medium. The dynamic binding capacity will vary depending on several factors such as target protein, flow rate etc.

If removal of the GST moiety (a naturally occurring protein with M_r 26 000) is required, the tagged protein can be digested with the appropriate sitespecific protease while bound to Glutathione Sepharose High Performance or, alternatively, after elution. Cleavage of GST-tagged protein bound to the column/bulk medium eliminates the extra step of separating the released protein from GST, since the GST-tag remains bound. The cleaved target protein is eluted using binding buffer.

Table 1. Characteristics of Glutathione Sepharose High Performance.

Ligand	glutathione and 10-carbon linker arm
Ligand concentration	1.5 to 3.5 mg glutathione/mL medium (based on Gly)
Dynamic binding capacity[1]	≈ 10 mg GST-tagged protein/mL medium, M_r 63 000
Mean particle size	34 µm
Bead structure	highly cross-linked 6% agarose
Maximum back pressure	0.3 MPa (3 bar, 43 psi)
Maximum flow rate[2]	600 cm/h

Recommended flow rates[3]	
Sample loading:	30 to 150 cm/h (1 to 5 mL/min using XK 16/20 column)
Washing and elution:	150 to 300 cm/h (5 to 10 mL/min using XK 16/20 column)
Chemical stability	All commonly used aqueous buffers, e.g. 1 M acetate, pH 4.0 and 6 M guanidine hydrochloride for 1 hour at room temperature
pH stability	pH 3 to 12
Storage temperature	+4°C to 30°C
Storage buffer	20% ethanol

[1] Dynamic binding capacity conditions ($QB_{60\%}$)

Sample:	1 mg/mL pure GST-tagged protein in binding buffer
Column volume:	0.4 mL
Flow rate:	0.2 mL/min (60 cm/h)
Binding buffer:	10 mM sodium phosphate, 140 mM NaCl, 2.7 mM KCl, pH 7.4
Elution buffer:	50 mM Tris-HCl, 10 mM reduced glutathione, pH 8.0

[2] H_2O at room temperature

[3] Binding of GST to glutathione is flow rate dependent and lower flow rates often increase the binding capacity. This is important during sample loading and elution.

2 Operation

Buffer preparation

Water and chemicals used for buffer preparation should be of high purity. We recommend filtering the buffers by passing them through a 0.45 µm filter before use.

Binding buffer: PBS, pH 7.3 (140 mM NaCl, 2.7 mM KCl, 10 mM Na2 HPO4, 1.8 mM KH 2 PO4, pH 7.3)

Elution Buffer: 50 mM Tris-HCl, 10 mM reduced glutathione, pH 8.0

Note: *1 to 10 mM DTT can be included in the binding and elution buffer.*

Sample preparation

The sample should be centrifuged and/or filtered through a 0.45 µm filter immediately before it is applied to the medium.

If the sample is too viscous, dilute it with binding buffer to prevent clogging the column.

3 Purification procedure

Column packing

Glutathione Sepharose High Perfomance is supplied preswollen in 20% ethanol. Prepare a slurry by decanting the 20% ethanol solution and replacing it with distilled water in a ratio of 75% settled medium to 25% distilled water.

Table 2. Recommended lab-scale columns for Glutathione Sepharose High Performance

Empty Column[1]	Packing flow rate (mL/min)		Max. recommended flow rate for chromatography (mL/min)
	first step	second step	
Tricorn™ 5/20	0.5	1	0.5
Tricorn 5/50	0.5	1	0.5
Tricorn 10/20	2	4	2
Tricorn 10/50	2	4	2
Tricorn 10/100	2	4	2
XK 16/20	5	10	5

Empty Column[1]	Packing flow rate (mL/min)		Max. recommended flow rate for chromatography (mL/min)
	first step	second step	
XK 26/20	13	27	13

[1] For inner diameter and maximum bed volumes and bed heights, see *Chapter 9 Ordering information, on page 26*.

Packing a column:

Step	Action
1	Assemble the column (and packing reservoir if necessary).
2	Remove air from the end-piece and adapter by flushing with water. Make sure no air has been trapped under the column bed support. Close the column outlet leaving the bed support covered with water.
3	Resuspend the medium and pour the slurry into the column in a single continuous motion. Pouring the slurry down a glass rod held against the column wall will minimize the introduction of air bubbles.
4	If using a packing reservoir, immediately fill the remainder of the column and reservoir with water. Mount the adapter or lid of the packing reservoir and connect the column to a pump. Avoid trapping air bubbles under the adapter or in the inlet tubing.

Step	Action
5	Open the bottom outlet of the column and set the pump to run at the desired flow rate. Ideally, Sepharose High Performance media are packed in XK or Tricorn columns in a two-step procedure: Do not exceed 1.0 bar (0.1 MPa) in the first step and 3.5 bar (0.35 MPa) in the second step. If the packing equipment does not include a pressure gauge, use a packing flow rate of 5 mL/min (XK 16/20 column) or 2 mL/min (Tricorn 10/100 column) in the first step, and 9 mL/min (XK 16/20 column) or 3.6 mL/min (Tricorn 10/100 column) in the second step. See Table 3 for packing flow rates for other columns. If the recommended pressure or flow rate cannot be obtained, use the maximum flow rate your pump can deliver. This should also give a wellpacked bed. ***Note:*** *For subsequent chromatography procedures, do not exceed 75% of the packing flow rate. See Table 3 for flow rates for chromatography.*
6	Maintain packing flow rate for at least 3 bed volumes after a constant bed height is reached. Mark the bed height on the column.
7	Stop the pump and close the column outlet.
8	If using a packing reservoir, disconnect the reservoir and fit the adapter to the column.

Step	Action
9	With the adapter inlet disconnected, push the adapter down into the column until it reaches the mark. Allow the packing solution to flush the adapter inlet. Lock the adapter in position.
10	Connect the column to a pump or a chromatography system and start equilibration. Re-adjust the adapter if necessary.

Column purification

Step	Action
1	Equilibrate the column with approx. 5 column volumes of binding buffer.
2	Apply the centrifuged and/or filtered sample.
3	Wash the column with 5 to 10 column volumes of binding buffer or until no material appears in the flow-through. Save the flow-through for measuring the binding efficiency to the medium, i.e. by SDS-PAGE.
4	Elute the bound protein with 5 to 10 column volumes of elution buffer.

Note: • *One of the most important parameters affecting the binding of GST-tagged proteins or other glutathione binding proteins to Glutathione Sepharose High Performance is the flow rate. Due to the relatively slow binding kinetics between GST*

and glutathione, it is important to keep the flow rate low during sample application for maximum binding capacity. Protein characteristics, pH and temperature are other factors that may affect the binding capacity

- *Volumes and times used for elution may vary among tagged proteins. Additional elutions with higher concentrations of glutathione may be required. Flow-through, wash and eluted material from the column should be monitored for GST-tagged proteins using SDS-PAGE in combination with Western Blot if necessary.*
- *The GST Detection Module can be used to optimize conditions for elution or to trace steps in the purification of a GST-tagged protein. The Module is designed to identify GST-tagged proteins using either a biochemical or an immunological assay (1).*
- *The concentration of GST-tagged protein can be estimated by measuring the absorbance at 280 nm. The GST-tag can be approximated using the conversion; A280 ≈ 1 corresponds to ≈ 0.5 mg/mL.*
- *The concentration of GST-tagged proteins may also be determined by standard chromogenic methods (e.g. Lowry, BCA, and Bradford assays). If Lowry or BCA assays are to be used, the sample must first be buffer exchanged using a HiTrap Desalting column or dialysed against PBS to remove glutathione, which can interfere with the protein measurement. The Bradford method can be used in the presence of glutathione.*

- *The reuse of Glutathione Sepharose High Performance depends on the nature of the sample and should only be performed with identical samples to prevent cross-contamination.*

4 Cleaning Glutathione Sepharose High Performance

If the medium appears to be losing binding capacity, it may be due to an accumulation of precipitate, denatured or nonspecifically bound proteins.

Removal of precipitated or denatured substances:

- Wash with 2 column volumes of 6 M guanidine hydrochloride, immediately followed by 5 column volumes of PBS, pH 7.3.

Removal of hydrophobically bound substances:

- Wash with 3 to 4 column volumns of 70% ethanol or 2 column volumes of 1% Triton™ X-100, immediately followed by 5 column volumes of PBS, pH 7.3.

5 Storage

Store the packed column at +4°C to 30°C in 20% ethanol.

6 Cleavage of GST-tagged proteins

In most cases, the fusion partner of interest retains functional activity and the functional test can be performed using intact fusion with GST. If removal of the GST-tag is necessary, tagged proteins containing a PreScission™ Protease recognition site, a thrombin recognition site or a factor Xa recognition site may be cleaved either while bound to Glutathione Sepharose High Performance or in solution after elution.

Cleavage after elution is suggested if optimization of cleavage conditions is necessary. Samples can easily be removed at various time points and analyzed by SDS-PAGE to estimate the yield, purity and extent of digestion. The amount of protease used, the temperature and the length of incubation required for complete digestion may vary depending on the fusion protein.

Optimal conditions for each tagged protein should be determined in pilot experiments, e.g. incubation time may be reduced by adding agreater amount of enzyme.

PreScission Protease

PreScission Protease, M_r 46 000.

PreScission cleavage buffer: 50 mM Tris-HCl, 150 mM NaCl, 1 mM EDTA, 1 mM dithiothreitol (DTT), pH 7.5.

PreScission Protease cleavage of GST-tagged protein bound to the column/bulk medium

Assumption: 8 mg GST-tagged proteins bound/mL medium

Step	Action
1	Follow steps 1 to 3 under *Column purification, on page 8*.
2	Wash the tagged protein bound Glutathione Sepharose High Performance with 10 bed volumes of PreScission cleavage buffer.
3	Prepare the PreScission Protease mix: For each mL of Glutathione Sepharose High Performance bed volume, prepare a mixture of 80 µL (160 units) of PreScission Protease and 920 µL of PreScission cleavage buffer at +4°C. (8 mg tagged protein bound/mL medium).
4	Load the PreScission Protease mixture onto the column. Seal the column. If batch format is used, add PreScission Protease mixture to the Glutathione Sepharose High Performance pellet. Gently shake or rotate the suspension.
5	Incubate at +4°C for 4 hours.

Step	Action
6	Following incubation, wash the column with approx. 3 bed volumes of PreScission cleavage buffer. Collect the eluate in different tubes to avoid dilution of the tagged protein and analyse it. If batch format is used, centrifuge the suspension at 500 × g for 5 minutes to pellet the Glutathione Sepharose High Performance and carefully transfer the eluate to a tube. The eluate will contain the protein of interest, while the GST portion of the tagged protein and the PreScission Protease will remain bound to the Glutathione Sepharose High Performance.

PreScission Protease cleavage of eluted GST-tagged protein

Assumption: 8 mg GST-tagged protein bound/mL medium

Step	Action
1	Remove the reduced glutathione from the eluate using a quick buffer exchange on HiTrap Desalting, a PD-10 column or HiPrep™ 26/10 Desalting depending on sample volume, or dialyse against PreScission cleavage buffer.
2	Add 1 µL (2 U) of PreScission Protease for each 100 µg of tagged protein in the eluate. If the amount of tagged protein in the eluate has not been determined, add 80 µL (160 units) of PreScission Protease for each mL of Glutathione Sepharose High Performance bed volume. (8 mg tagged protein bound/ mL medium).

Step	Action
3	Incubate at +4°C for 4 hours.
4	Once digestion is complete, apply the sample to washed and equilibrated Glutathione Sepharose High Performance to remove the GST moiety of the tagged protein and the PreScission Protease from the protein of interest.
5	Incubate for 20 to 30 min. at room temperature.
6	Sediment the medium by centrifugation at 500 × g for 5 min. The protein of interest will be found in the supernatant.

Thrombin

Thrombin, M_r 37 000.

Thrombin cleavage buffer: PBS, pH 7.3.

Preparation of thrombin solution:

Step	Action
1	Dissolve 500 U thrombin in cold 500 µL PBS, pH 7.3 (1 U/µL).
2	Swirl gently to dissolve.
3	Freeze as 80 µL aliqouts and keep at –80°C.

Thrombin cleavage of GST-tagged protein bound to the column/bulk medium

Assumption: 8 mg GST-tagged protein bound/mL medium

Step	Action
1	Follow steps 1 to 3 under *Column purification, on page 8*.
2	Prepare the thrombin mix: For each mL of Glutathione Sepharose High Performance bed volume, prepare a mixture of 80 µL (80 units) of thrombin and 920 µL of PBS, pH 7.3 (8 mg GST-tagged protein bound/mL medium).
3	Load the thrombin mix onto the column. Seal the column. If batch format is used, add the thrombin mixture to the Glutathione Sepharose High Performance pellet. Gently shake or rotate the suspension.
4	Incubate at room temperature (22°C to 25°C) for 2 to 16 hours.
5	Following incubation, wash the column with approx. 3 bed volumes of PBS, pH 7.3. Collect the eluate in different tubes to avoid dilution of the tagged protein and analyse it. If batch format is used, centrifuge the suspension at 500 x g for 5 minutes to pellet the Glutathione Sepharose High Performance and carefully transfer the eluate to a tube. The eluate will contain the protein of interest and thrombin, while the GST portion of the tagged protein will remain bound to the Glutathione Sepharose High Performance.

Thrombin cleavage of eluted GST-tagged protein

Assumption: 8 mg GST-tagged protein bound/mL medium

Step	Action
1	Add 10 µL (10 units) of thrombin solution for each mg of tagged protein in the eluate. If the amount of tagged protein in the eluate has not been determined, add 80 µL (80 U) of thrombin solution for each mL of Glutathione Sepharose High Performance bed volume from which the tagged protein was eluted. (8 mg tagged protein bound/mL medium).
2	Incubate at room temperature (22°C to 25°C) for 2 to 16 hours.
3	Once digestion is complete, GST can be removed by first removing glutathione using a quick buffer exchange on HiTrap Desalting, a PD-10 column or HiPrep 26/10 Desalting depending on sample volume, or by dialysis against PBS, pH 7.3. Follow this by applying the sample to washed and equilibrated Glutathione Sepharose High Performance.
4	Incubate for 20 to 30 min. at room temperature.
5	Sediment the medium by centrifugation at 500 × g for 5 min. The supernatant will contain the protein of interest and thrombin, while the GST portion of the tagged protein will remain bound to the Glutathione Sepharose High Performance.

Factor Xa

Factor Xa, M_r 48 000.

Note: *Factor Xa consists of two subunits linked by disulfide bridges. As glutathione can disrupt disulfide bridges, it should be removed from the sample prior to the cleavage reaction. Factor Xa cleavage buffer: 50 mM Tris-HCl, 150 mM NaCl, 1 mM $CaCl_2$, pH 7.5*

Preparation of factor Xa solution:

Step	Action
1	Dissolve 400 U factor Xa in 400 µL cold water (1 U/µL).
2	Swirl gently to dissolve.
3	Freeze as 80 µL aliqouts and keep at –80°C.

Factor Xa cleavage of GST-tagged protein bound to the column/bulk medium

Assumption: 8 mg GST-tagged protein bound/mL medium

Step	Action
1	Follow steps 1 to 3 under *Column purification, on page 8*.
2	Wash the tagged protein bound Glutathione Sepharose High Performance with 10 bed volumes of factor Xa cleavage buffer.
3	Prepare the factor Xa mix: For each mL of Glutathione Sepharose High Performance bed volume, prepare a mixture of 80 µL (80 units) of factor Xa and 920 µL of factor Xa cleavage buffer. (8 mg tagged protein bound/mL medium).

Step	Action
4	Load the factor Xa mixture onto the column. Seal the column. If batch format is used, add factor Xa mixture to the Glutathione Sepharose High Performance pellet. Gently shake or rotate the suspension.
5	Incubate at room temperature (22°C to 25°C) for 2 to 16 hours.
6	Following incubation, wash the column with approx. 3 bed volumes of factor Xa cleavage buffer. Collect the eluate in different tubes to avoid dilution of the tagged protein and analyse it. If batch format is used, centrifuge the suspension at 500 × g for 5 minutes to pellet the Glutathione Sepharose High Performance and carefully transfer the eluate to a tube. The eluate will contain the protein of interest and factor Xa, while the GST portion of the tagged protein will remain bound to the Glutathione Sepharose High Performance.

Factor Xa cleavage of eluted GST-tagged protein

Assumption: 8 mg GST-tagged protein bound/mL medium

Step	Action
1	Remove reduced glutathione from the eluate using a quick buffer exchange on HiTrap Desalting, a PD-10 column or HiPrep 26/10 Desalting depending on sample volume, or dialyse against factor Xa cleavage buffer.

Step	Action
2	Add 10 µL (10 units) of factor Xa solution for each mg tagged protein in the eluate. If the amount of tagged protein in the eluate has not been determined, add 80 µL (80 units) of factor Xa solution for each mL of Glutathione Sepharose High Performance bed volume from which the tagged protein was eluted. (8 mg tagged protein bound/mL medium).
3	Incubate at room temperature (22°C to 25°C) for 2 to 16 hours.
4	Once digestion is complete, apply the sample to washed and equilibrated Glutathione Sepharose High Performance to remove the GST moiety of the tagged protein.
5	Incubate for 20 to 30 min. at room temperature.
6	Sediment the medium by centrifugation at 500 × g for 5 min. The protein of interest will be found in the supernatant together with factor Xa.

7 Troubleshooting guide

Consult the Handbook 18115758 (1) for more detailed information and pGEX instructions regarding troubleshooting recommendations for expression, fermentation and solubilization.

GST-tagged protein does not bind to Glutathione Sepharose High Performance

- **GST-tagged protein denatured by sonication:** Too extensive sonication can denature the tagged protein and prevent it binding to Glutathione Sepharose High Performance. Use mild sonication conditions during cell lysis. Conditions for sonication must be empirically determined.
- **Add DTT prior to cell lysis and to buffers:** Adding DTT to a final concentration of 1–10 mM may significantly increase binding of some GST-tagged proteins to Glutathione Sepharose High Performance.
- **Test the binding of GST from parental pGEX:** Prepare a sonicate of cells harboring the parental pGEX plasmid and check binding to the matrix. If GST produced from the parental plasmid binds with high affinity, the fusion protein may have altered the conformation of GST, thereby reducing its affinity. Adequate results may be obtained by reducing the temperature used for binding to +4°C, and by limiting column washing.
- **Equilibrate Glutathione Sepharose High Performance before use:** Binding of GST-tagged proteins to Glutathione Sepharose High Performance is not efficient at pH less than 6.5 or greater than 8. Check that the Glutathione Sepharose High Performance has been equilibrated with a buffer 6.5 to 8.0 (e.g. PBS) before the clarified cell lysate is applied.

- **Use fresh Glutathione Sepharose High Performance:** If the Glutathione Sepharose High Performance has already been used several times, it may be necessary to use fresh Glutathione Sepharose High Performance. See also "Cleaning Glutathione Sepharose High Performance".
- **Decrease flow rate during sample load.** See *note on page 8*.

GST-tagged protein is not eluted efficiently from Glutathione Sepharose High Performance

- **Increase the time used for elution:** Decrease the flow during elution.
- **Increase the volume of elution buffer:** Sometimes, especially after on-column cleavage of tagged protein, a larger volume of buffer may be necessary to elute the tagged protein.
- **Increase the concentration of glutathione in the elution buffer:** The 10 mM recommended in this protocol should be sufficient for most applications, but exceptions exist. Try 50 mM Tris- HCl, 20 to 40 mM reduced glutathione, pH 8.0 as elution buffer.
- **Increase the pH of the elution buffer:** A low pH may limit elution from Glutathione Sepharose High Performance. Increasing the pH of the elution buffer to pH 8 to 9 may improve elution without requiring an increase in the concentration of glutathione used for elution.
- **Increase the ionic strength of the elution buffer:** Adding 0.1 to 0.2 M NaCl to the elution buffer may also improve results.
- **Use fresh elution buffer (reduced glutathione).**

- **Add a non-ionic detergent to the elution buffer:** Non-specific hydrophobic interactions may prevent solubilization and elution of tagged proteins from Glutathione Sepharose High Performance. Adding a non-ionic detergent may improve results. Adding 0.1% Triton X-100 or 2% N-octylglucoside can significantly improve elution of some GST-tagged proteins.

Multiple bands are observed after electrophoresis/ Western Blotting analysis of eluted target protein.

- **M_r 70 000 protein co-purifies with the GST-tagged protein:** The M_r 70 000 protein is probably a protein product of the E. coli gene dnaK. This protein is involved in protein folding in E. coli. It has been reported that this association can be disrupted by incubating the tagged protein in 50 mM Tris-HCl, 2 mM ATP, 10 mM $MgSO_4$, pH 7.4 for 10 min. at +37°C prior to loading on Glutathione Sepharose High Performance.

  Alternatively, remove the DnaK protein by passing the tagged protein solution through ATP-agarose or by ion exchange.
- **Add a protease inhibitor:** Multiple bands may be a result of partial degradation of tagged proteins by proteases. Adding 1 mM PMSF to the lysis solution may improve results. A non-toxic, water-soluble alternative to PMSF is 4-(2- aminoethyl)-benzenesulfonyl fluoride hydrochloride (AEBSF), commercially available as Pefabloc™ SC from Boehringer Mannheim.

Note: *Serine protease inhibitors must be removed prior to cleavage by thrombin or factor Xa. PreScission Protease is not a consensus serine protease and is insensitive to many of the protease inhibitors tested at Cytiva.*

- **Use a protease-deficient host:** Multiple bands may be the result of proteolysis in the host bacteria. If this is the case, the use of a hostdeficient strain may be required (*e.g. lon- or ompT*). *E. coli* BL21 is provided with the pGEX vectors. This strain is *ompT*.
- **Decrease sonication:** Cell disruption is apparent by partial clearing of the suspension and can be checked by microscopic examination. Adding lysozyme (0.1 volume of a 10 mg/mL lysozyme solution in 25 mM Tris-HCl, pH 8.0) prior to sonication may improve results. Avoid frothing as this may denature the fusion protein. Over-sonication can also lead to the co-purification of host proteins with the GST-tagged protein.
- **Include an additional purification step:** Additional bands may be caused by the co-purification of a variety of proteins known as chaperonins, which are involved in the correct folding of nascent proteins in *E. coli*. These include, but may not be limited to: DnaK (M_r ~ 70 000), DnaJ (M_r ~ 37 000), GrpE (M_r ~ 40 000), GroEL (M_r ~ 57 000) and GroES (M_r ~ 10 000). Several methods for purifying GST-tagged proteins from these co-purifying proteins have been described.

- **Cross-adsorb antibody with E. coli proteins:** Depending on the source of the anti-GST antibody, it may contain antibodies that react with various *E. coli* proteins that may be present in your tagged protein sample. Cross-adsorb the antibody with an *E. coli* sonicate to remove anti-*E. coli* antibodies from the preparation. Anti-GST antibody from Cytiva has been cross-adsorbed against *E. coli* proteins and tested for its lack of non-specific background binding in Western Blots.

Incomplete cleavage of GST-tagged proteins

- The PreScission Protease, thrombin or factor Xa to fusion protein ratios are incorrect: Check the amount of tagged protein in the digestion. Note that the capacity of Glutathione Sepharose High Performance for GST is > 10 mg/mL medium. In most purifications, however, the matrix is not saturated with tagged protein.

  Ratios: PreScission protease, at least 10 units/mg tagged protein.

  Thrombin, at least 10 units/mg tagged protein. One cleavage unit of thrombin from Cytiva digests ≥ 90% of 100 µg of a test tagged protein in 16 hours at +22°C.

  Factor Xa, at least 1% (w/w) tagged protein. For some tagged proteins, up to 5% factor Xa can be used. The optimum amount must be determined empirically.

  In some cases, a **tagged protein** concentration of 1 mg/mL has been found to give optimal results. Adding ≈ 0.5% SDS (w/v) to the reaction buffer can significantly improve factor Xa cleavage with some tagged proteins. Various concentrations of SDS should be tested to find the optimum concentration.

- **Increase incubation time and enzyme concentration:** For PreScission Protease, thrombin or factor Xa, increase the reaction time to 20 hours or more if the tagged protein is not degraded by extensive incubation. The amount of enzymes can also be increased.
- **Verify the presence of specific cleavage sites:** Check the DNA sequence of the construct. Compare it with a known sequence and verify that the different specific cleavage sites for the enzyme used have not been altered during the cloning of your tagged protein.

Ensure that cleavage enzyme inhibitors are absent:

- **PreScission Protease:** Buffer exchange or dialyse the tagged protein against 50 mM Tris-HCl, 150 mM NaCl, 1 mM EDTA, 1 mM DTT, pH 7.5 before cleavage.
- Factor Xa: Buffer exchange on HiTrap Desalting, a PD-10 column or HiPrep 26/10 Desalting depending on the sample volume, or dialyse against 50 mM Tris-HCl, 150 mM NaCl, 1 mM $CaCl_2$, pH 7.5.
- **Factor Xa is not properly activated:** Functional factor Xa requires activation of factor X with Russell's viper venom. Activation conditions are a ratio of Russell's viper venom to factor Xa of 1% in 8 mM Tris-HCl, 70 mM NaCl, 8 mM $CaCl_2$, pH 8.0. Incubate at +37°C for 5 min. Factor Xa from Cytiva has been preactivated by this procedure.
- **The first amino acid after the factor Xa recognition sequence is Arg or Pro:** Check the sequence of the tagged partner to be sure that the first three nucleotides after the factor Xa recognition sequence do not code for Arg or Pro.

Multiple bands are observed after electrophoresis analysis of cleaved target protein:

- **Determine when the bands appear:** Test to be certain that additional bands are not present prior to PreScission Protease, thrombin or factor Xa cleavage. Such bands may be the result of proteolysis in the host bacteria.
- Tagged partner may contain recognition sequences for PreScission Protease, thrombin or factor Xa: Check the sequences. See the Handbook 18115758 (1) for details.

8 Reference

1. GST Gene Fusion System Handbook, Cytiva, Code No. 18115758.

9 Ordering information

Product	Quantity	Code No.
Glutathione Sepharose High Performance [1]	25 mL	17527901
Glutathione Sepharose High Performance *Footnote.*	100 mL	17527902

[1] Larger quantities are available. Please contact Cytiva for more information.

Site-Specific Proteases

Product	Quantity	Code No.
PreScission Protease	500 units	27084301
Thrombin	500 units	27084601
Factor Xa	400 units	27084901

Related Products

Product	Quantity	Code No.
GST Detection Module	50 reactions	27459001
Anti-GST Antibody	0.5 mL	27457701
Recombinant Protein Handbook	1	18110502
GST Gene Fusion System Handbook	1	18115758
Affinity Cromatography Handbook, Principles and methods	1	18102229

Prepacked columns		
GSTrap HP	5 × 1 mL	17528101
GSTrap HP	100 × 1 mL[1]	17528105
GSTrap HP	1 × 5 mL	17528201
GSTrap HP	5 × 5 mL	17528202
GSTrap HP	100 × 5 mL*Footnote.*	17528205
HiTrap Desalting	5 × 5 mL	17140801
HiTrap Desalting	100 × 5 mL*Footnote.*	11000329
PD-10 Desalting Column	30	17085101
HiPrep 26/10 Desalting	1 × 53 mL	17508701
HiPrep 26/10 Desalting	4 × 53 mL	17508702

[1] Special pack delivered on specific customer order.

Empty lab-scale columns		
Tricorn 5/20 column, 5 mm i.d., max 0.55 mL bed volume or 2.8 cm bed height	1	18116308
Tricorn 5/50 column, 5 mm i.d., max 1.1 mL bed volume or 5.8 cm bed height	1	18116309
Tricorn 10/20 column, 10 mm i.d., max 2.2 mL bed volume or 2.8 cm bed height	1	18116313
Tricorn 10/50 column, 10 mm i.d., max 4.5 mL bed volume or 5.8 cm bed height	1	18116314

Empty lab-scale columns		
Tricorn 10/100 column, 10 mm i.d., max 8.5 mL bed volume or 10.8 cm bed height	1	18116315
XK 16/20 column, 16 mm i.d., max 30 mL bed volume or 15 cm bed height	1	18877301
XK 26/20 column, 26 mm i.d., max 65 mL bed volume or 12.5 cm bed height	1	18100072

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A license for commercial use of GST gene fusion vectors must be obtained from Chemicon International Incorporated, 28820 Single Oak Drive, Temecula, California 92590, USA.

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