

Strep(II)-tagged protein purification ÄKTAprime Plus

Preparing the buffers

- Use high purity water and chemicals.
- Filter all buffers through a 0.45 µm filter before use.

Binding buffer (port A1):

100 mM Tris-HCl, 150 mM NaCl, 1 mM EDTA, pH 8.0

Elution buffer (port B):

100 mM Tris-HCl, 150 mM NaCl, 1 mM EDTA, 2.5 mM desthiobiotin, pH 8.0

Prepare at least 500 mL of each eluent.

Preparing the sample

Step Action

- 1 Adjust the sample to composition of binding buffer by:
 - diluting the sample in binding buffer or
 - by buffer exchange using HiTrap[™] Desalting or HiPrep[™] 26/10 Desalting.
- 2 Pass the sample through a 0.45 µm filter.

Preparing the system

Step Action

1 Place the inlet tubing from port A1 (8-port valve) in the binding buffer and the tubing from port B (2-port valve) in the elution buffer.

2 Place the three brown waste tubings in waste.

- 3 Connect the column between port 1 on the injection valve (7port valve) and the UV flow cell (see Ordering information on next page for suitable columns).
- 4 Fill the fraction collector rack with 18 mm tubes¹ (minimum 10) and position the white plate on the fractionation arm against the first tube.
- 5 Connect a sample loop large enough for your sample between port 2 and 6 on the injection valve. Use a syringe to manually fill the loop.

Note:

If a Superloop $^{\rm M}$ is needed, additional information is supplied in the instructions for Superloop.

Selecting Application Template and starting the method

Step Action

2

3

- 1 Check the communication to PrimeView[™]. At the lower right corner of the screen the text **Controlled By** →**prime** should be displayed.
 - Use the arrow and **OK** buttons to move in the menu tree until you find **Affinity Purification any HiTrap**.



Enter the sample volume and press **OK** to start the template. **Note:**

If a 5 mL column is preferred, see cue card on p.36.

Theoretical gradient in **Affinity Purification any HiTrap** Application Template.



Total separation time = 47 min + sample application time

¹ The number of tubes to insert in the fraction collector varies with the sample volume. Fill the fraction collector with 20 tubes + one tube/mL sample. For example, if the sample volume is 10 mL, fill the fraction collector with 20 + 10 = 30 tubes. However, note that the maximum capacity of the fraction collector is 95 tubes, limiting the sample volume to 75 mL.

Typical result

Sample:	Clarified lysate of <i>E. coli</i> expressing	
	Strep(II)-tagged protein.	
Column:	StrepTrap [™] HP 1 mL	
Binding buffer (port A1):	100 mM Tris-HCl, 150 mM NaCl, 1 mM EDTA, pH 8.0	
Elution buffer (port B):	100 mM Tris-HCl, 150 mM NaCl, 1 mM EDTA, 2.5 mM desthiobiotin, pH 8.0	



Troubleshooting

High backpressure:

Column clogged – Clean the column according to instructions. Make sure the sample has been centrifuged and/or filtered through a 0.45 μm filter.

System clogged – Replace the column with a piece of tubing. Check pressure. If backpressure > 0.3 MPa, clean system according to manual.

No binding:

- Regenerate the column with 3 column volumes (CV) water, 3 CV 0.5 M NaOH, 3 CV water, 5 CV binding buffer before starting the run.
- Check that the correct column is used.
- Check that the inlet tubing from each buffer is connected to the correct inlet port.
- Check that the composition and pH of the buffers are correct.
- Check that the sample has been adjusted to the binding buffer conditions.
- Check that your sample contains target protein.

No elution:

- Check that the inlet tubing from each buffer is connected to the correct inlet port.
- Check that the composition and pH of the buffers are correct.
- Use alternative elution conditions according to the column instructions.
- Check that your sample contains target protein.

Ordering information

Product

	Quantity	Product code
StrepTrap HP	5×1mL	28907546
HiTrap Desalting	5 × 5 mL	17140801
	100 × 5 mL ¹	11000329
HiPrep 26/10 Desalting	1 (53 mL)	17508701
	4 (53 mL)	17508702
Superloop 10 mL	1	18111383
Superloop 50 mL	1	18111384
Superloop 150 mL	1	18102385

¹ Pack size available by special order

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