

Streptavidin Mag Sepharose

Mag Sepharose

Instructions for Use

Streptavidin Mag Sepharose™ is available in the following pack sizes (instructions for use are included in both the packs):

- Streptavidin Mag Sepharose, 10% medium slurry, 2 × 1 mL
- Streptavidin Mag Sepharose, 10% medium slurry, 5 × 1 mL

Note: *1 mL medium slurry is sufficient for 20 reactions according to the recommended immunoprecipitation protocol (see [Chapter 5 Protocol for immunoprecipitation, on page 10](#)). 1 mL of 10% (v/v) medium slurry contains 100 µL of magnetic beads.*

Streptavidin Mag Sepharose products contain magnetic beads that enable strong binding of biotinylated biomolecules. Two protocols are included in this instruction; one for capturing and eluting biotinylated proteins and another for enrichment of proteins by immunoprecipitation.

Streptavidin Mag Sepharose is intended for research use only, and should not be used in any clinical or in vitro procedures for diagnostic purposes.

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1 Principle

Streptavidin Mag Sepharose is an affinity chromatography medium with high affinity for biotinylated proteins and other biotinylated biomolecules. The medium consists of streptavidin ligand coupled to magnetite-containing Sepharose beads.

The binding of streptavidin to biotin is one of the strongest known non-covalent biological interactions. Hence, denaturing conditions are generally required for the efficient elution of biotinylated biomolecules. Alternatively, biotinylated biomolecules bound to Streptavidin Mag Sepharose can be used to capture interacting target substances such as proteins. Impurities are removed by washing, and the enriched target protein is eluted using less harsh elution conditions.

Streptavidin Mag Sepharose provides flexible enrichment of biomolecules. The enrichment process can be easily scaled just by varying the volume of the Mag Sepharose beads.

For simplified handling, Mag Sepharose products can be used together with a magnetic rack, for example, MagRack 6 for sample volumes up to 1.5 mL or MagRack Maxi for sample volumes up to 50 mL (see [Chapter 2 Advice on handling, on page 3](#)).

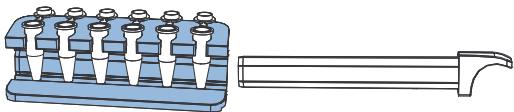
2 Advice on handling

Note: *Streptavidin Mag Sepharose is intended for single use only.*

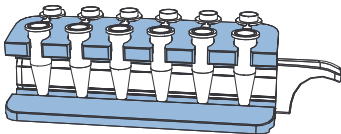
General magnetic separation

The protocols recommended in this instruction use 1.5 mL Eppendorf tubes and MagRack 6 (see [Chapter 4 Protocol for capturing and eluting biotinylated proteins, on page 8](#) and [Chapter 5 Protocol for immunoprecipitation, on page 10](#)).

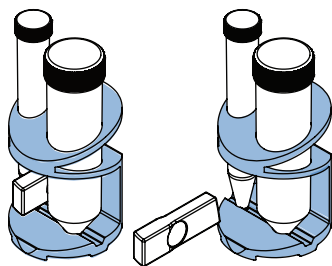
1. **Remove the magnet before adding the liquid.**



2. **Insert the magnet before removing the liquid.**



When using volumes above 1.5 mL, MagRack Maxi, designed for sample volumes up to 50 mL, is recommended.



For volumes more than 50 mL, the beads can be spun down using a swing-out centrifuge.

Dispensing the medium slurry

Step	Action
1	Vortex the medium slurry to homogeneity before dispensing.
2	Pipette out the required amount of medium slurry into the appropriate tube immediately after resuspension.
3	Repeat the resuspension step between each pipetting step.

Handling the liquid

Step	Action
1	Remove the magnet from the magnetic rack, before adding liquid to the tube.

Step	Action
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| 2 | Resuspend the beads by vortexing or by inverting the tube manually, after adding the liquid. |
| 3 | Use the magnetic rack with the magnet inserted for each liquid removal step. |
| 4 | Remove the liquid from the lid of the vial using a pipette, if needed. |

Incubating

Step	Action
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|---|---|
| 1 | Resuspend the Mag Sepharose beads well in solution by mixing end-over-end or by using a benchtop shaker, during incubation. |
| 2 | Longer incubation times might be necessary when purifying larger volumes of samples, samples having low affinity or samples with very low concentrations. |

Note: *The samples can generally be incubated at room temperature. However, they can be incubated at +4°C also, if specifically recommended for the sample.*

3 Preparing buffers and samples

Recommended buffers

Use the listed recommended buffers. Use any of the alternative buffers for optimization.

Note: Use high-purity water and chemicals for buffer preparation.

Table 1. Recommended buffers for capturing and eluting biotinylated proteins

Buffer	Composition
Binding buffer	Tris-buffered saline (TBS) (50 mM Tris-HCl, 150 mM NaCl), pH 7.5
Washing buffer	TBS, 2 M urea, pH 7.5
Elution buffer	2% SDS

Table 2. Recommended buffers for immunoprecipitation

Buffer	Composition
Binding buffer	TBS, pH 7.5
Washing buffer	TBS, 2 M urea, pH 7.5
Elution buffer	0.1 M glycine-HCl, 2 M urea, pH 2.9

Table 3. Alternative buffers

Buffer	Composition
Washing buffer	<ul style="list-style-type: none">• TBS, pH 7.5 (mild wash)• 0.1 M triethanolamine, 0.5 M NaCl, pH 9.0
Elution buffer	<ul style="list-style-type: none">• 0.1 M glycine, pH 2.5 to pH 3.1• 0.1 M citric acid, pH 2.5 to pH 3.1• 2% SDS• 0.1 M ammonium hydroxide, pH 10.0 to pH 11.0

- Streptavidin HP Buffer kit is available for easy preparation of buffers (see [Chapter 8 Ordering information, on page 15](#)).
- Purity of the sample might be improved by adding detergents or salts to the wash buffer.

Preparing the sample

Step	Action
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| 1 | Adjust the sample to the composition and pH of the binding buffer. pH can be adjusted by either diluting the sample with binding buffer or by buffer exchange using PD-10 MiniTrap™ G-25 or HiTrap™ Desalting columns (see Chapter 8 Ordering information, on page 15). |
| 2 | Clarify the sample before applying it to the beads, if needed. |
| 3 | Inhibiting protease activity in the sample prevents degradation of the target protein. A Protease Inhibitor Mix from Cytiva is available (see Chapter 8 Ordering information, on page 15). |

4 Protocol for capturing and eluting biotinylated proteins

This protocol is suitable for capturing and eluting biotinylated proteins. The first part of the protocol (1-3) captures biotinylated proteins from the sample and can be used for depletion.

Step	Action
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1	Preparing the Mag Sepharose beads
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| | <ul style="list-style-type: none">a. Mix the medium slurry thoroughly by vortexing. Dispense 100 μL of the homogenous medium slurry into an Eppendorf tube.b. Place the Eppendorf tube in the magnetic rack (see Chapter 2 Advice on handling, on page 3).c. Remove the storage solution. |
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2	Equilibrating
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| | <ul style="list-style-type: none">a. Add 500 μL binding buffer and resuspend the medium.b. Remove the liquid. |
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3	Applying the sample
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| | <ul style="list-style-type: none">a. Add 300 μL of sample. If the sample volume is less than 300 μL, dilute to 300 μL with binding buffer.b. Resuspend the medium and incubate for 30 minutes with slow end-over-end mixing or by using a benchtop shaker.c. Remove the sample. |
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4	Washing (perform this step three times totally)
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| | <ul style="list-style-type: none">a. Add 500 μL washing buffer and resuspend the medium.b. Remove the liquid. |
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Step	Action
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5	Eluting biotinylated proteins
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- Add 100 μ L elution buffer.
- Resuspend the medium and incubate at 95°C to 100°C for five minutes.
- Remove and collect the eluted fraction. The collected fraction contains the main part of the protein. If needed, repeat the elution.

Note:

The streptavidin-biotin bond can be broken efficiently only by harsh denaturing conditions. Hence, dissociation of biotin from streptavidin will denature both—the biotinylated protein and streptavidin, causing a leakage of the streptavidin monomers.

5 Protocol for immunoprecipitation

This protocol is suitable as a starting point for enrichment of target proteins using immobilized antibodies.

Step	Action
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1	Preparing the Mag Sepharose beads
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- Mix the medium slurry thoroughly by vortexing. Dispense 50 μ L of the homogenous medium slurry into an Eppendorf tube.
- Place the Eppendorf tube in the magnetic rack (see [Chapter 2 Advice on handling, on page 3](#)).
- Remove the storage solution.

Step	Action
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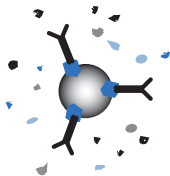
2	Equilibrating
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- Add 500 μ L binding buffer and resuspend the medium.
- Remove the liquid.



3	Binding of biotinylated antibody
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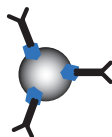
- Add 300 μ L of biotinylated antibody solution (~0.2-0.4 mg/mL). If the sample volume is less than 300 μ L, dilute to 300 μ L with the binding buffer.
- Resuspend the medium and incubate for 30 minutes with slow end-over-end mixing or by using a benchtop shaker.
- Remove the liquid.



Step	Action
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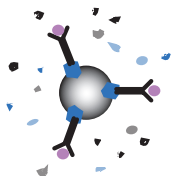
4	Washing (perform this step two times totally)
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- Add 500 μL washing buffer and resuspend the medium.
- Remove the liquid.



5	Binding of the target protein
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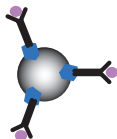
- Add 300 μL of sample. If the sample volume is less than 300 μL , dilute to 300 μL with binding buffer.
- Resuspend the medium and incubate for 60 minutes with slow end-over-end mixing or by using a benchtop shaker.
- Remove the liquid.



Step	Action
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6	Washing (perform this step three times totally)
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| | <ol style="list-style-type: none">Add 500 μL washing buffer and resuspend the medium.Remove the liquid. |
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7	Eluting
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| | <ol style="list-style-type: none">Add 50 μL of elution buffer.Resuspend the medium and incubate for two minutes.Remove and collect the elution fraction. The collected elution fraction contains the main part of the protein. If needed, repeat the elution. |
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6 Optimizing the parameters

The protocols recommended in this instruction (see [Chapter 4 Protocol for capturing and eluting biotinylated proteins, on page 8](#) and [Chapter 5 Protocol for immunoprecipitation, on page 10](#)) are suitable as starting points for most purifications involving biotinylated proteins. However, the optimal parameters for protein enrichment depend on the specific biomolecules used. Hence, optimization may be required to obtain the best results.

Examples of parameters which may require optimization are:

- Amount of beads
- Amount of biotinylated proteins/antibodies
- Amount of protein (antigen) to be enriched
- Incubation time
- Number of washes
- Buffer compositions, pH etc.

Non-specific binding can be minimized by decreasing the amount of Mag Sepharose beads, increasing the amount of biotinylated protein or by optimizing the wash conditions. If proteins are bound non-specifically to the Eppendorf tube, the bead solution can be transferred to a fresh tube during the last wash step.

7 Characteristics

Table 4. Streptavidin Mag Sepharose

Matrix	Highly cross-linked spherical agarose (Sepharose) containing magnetite
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Medium	Streptavidin-coupled NHS activated Mag Sepharose
Ligand	Streptavidin
Binding capacity	> 300 µg biotinylated BSA/mL of medium slurry
Particle size	37-100 µm
Working temperature	Room temperature and at +4°C
Storage solution	20% ethanol, 10% medium slurry
Storage temperature	+4°C to +8°C

8 Ordering information

Products	Quantity	Code No.
Streptavidin Mag Sepharose	2 × 1 mL 10% medium slurry	28985738
Streptavidin Mag Sepharose	5 × 1 mL 10% medium slurry	28985799

Related products	Quantity	Code No.
MagRack 6	1	28948964
MagRack Maxi	1	28986441
His Mag Sepharose Ni	2 × 1 mL 5% medium slurry	28967388
His Mag Sepharose Ni	5 × 1 mL 5% medium slurry	28967390
His Mag Sepharose Ni	10 × 1 mL 5% medium slurry	28979917
NHS Mag Sepharose	1 × 500 µL 20% medium slurry	28944009
NHS Mag Sepharose	4 × 500 µL 20% medium slurry	28951380
Protein A Mag Sepharose	1 × 500 µL 20% medium slurry	28944006

Related products	Quantity	Code No.
Protein A Mag Sepharose	4 × 500 µL 20% medium slurry	28951378
Protein A Mag Sepharose Xtra	2 × 1 mL 10% medium slurry	28967056
Protein A Mag Sepharose Xtra	5 × 1 mL 10% medium slurry	28967062
Protein G Mag Sepharose	1 × 500 µL 20% medium slurry	28944008
Protein G Mag Sepharose	4 × 500 µL 20% medium slurry	28951379
Protein G Mag Sepharose Xtra	2 × 1 mL 10% medium slurry	28967066
Protein G Mag Sepharose Xtra	5 × 1 mL 10% medium slurry	28967070
TiO ₂ Mag Sepharose	1 × 500 µL 20% medium slurry	28944010
TiO ₂ Mag Sepharose	4 × 500 µL 20% medium slurry	28951377
HiTrap™ Desalting	5 × 5 mL	17140801
PD MiniTrap™ G-25	50 columns	28918007
Protease Inhibitor Mix	1 mL	80650123
Streptavidin Buffer Kit	1	28913568

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