

Isolation of mononuclear cells



Contents

01	Introduction	3	• Notes	11
02	Ficoll-Paque density gradient media	5	• Troubleshooting inadequate performance	12
	• The separation principle	6	• Ficoll-Paque PLUS	13
	• A recommended standard method	7	• Ficoll-Paque PREMIUM products	13
	• Equipment and solutions required but not provided	8	• Properties of mononuclear cells isolated by the Ficoll-Paque method	14
	• Preparation of reagents	8	• Further applications of Ficoll-Paque PLUS and Ficoll-Paque PREMIUM	15
	• Preparation of the sample	9	• Applications using Ficoll-Paque PREMIUM 1.084	16
	• Procedure for isolation of mononuclear cells	9	• Applications using Ficoll-Paque PREMIUM 1.073	16
	• Washing the cell isolate	10	• Availability and storage	16
	• Procedure for isolation of granulocytes	10		
03			References	17
04			Ordering Information	19

01

Introduction

Ficoll-Paque™ products are sterile, ready-to-use density gradient media for isolating mononuclear cells in high yield and purity from small or large volumes of human peripheral blood, using a simple and rapid centrifugation procedure based on the method developed by Bøyum (1). Ficoll-Paque products can also be used to prepare purified mononuclear cells from sources other than human peripheral blood.

Ficoll-Paque products from Cytiva are:

- Sterile (moist heat sterilization), endotoxin tested (< 0.12 EU/ml) solutions of Ficoll™ PM400 and sodium diatrizoate available in different densities
- Recommended for small- or large-scale isolation of viable mononuclear cells in high yield from whole human peripheral blood, bone marrow, and umbilical cord blood
- Subjected to rigorous quality control testing, which ensures reproducible performance from batch to batch
- Supplied in bottles sealed with a rubber septum closure, which facilitates aseptic withdrawal of solution
- Available in convenient pack sizes: 6×100 ml and 6×500 ml. Detailed instructions for use are included with each pack
- Stable for at least 3 yr when stored at 4°C to 30°C and protected from light.

Separation of normal whole human peripheral blood by the procedure recommended in this booklet for Ficoll-Paque PLUS and Ficoll-Paque PREMIUM typically yields a mononuclear cell preparation with:

- $60 \pm 20\%$ recovery of the mononuclear cells present in the original blood sample
- $95 \pm 5\%$ mononuclear cells
- $> 90\%$ viability of the separated cells
- Maximum 5% granulocytes
- Maximum 10% erythrocytes

Ficoll-Paque PLUS (see page 13) is intended for research use only and has a density of 1.077 ± 0.001 g/ml at 20°C .

In October 2005, Cytiva launched Ficoll-Paque PREMIUM. This product (see page 13) is based on Ficoll-Paque PLUS but is manufactured within a quality management system certified to ISO 13485, and is intended for the preparation of human mononuclear cells for clinical research and cell therapy manufacturing applications. Ficoll-Paque PREMIUM, with a density of 1.077 ± 0.001 g/ml, can be incorporated into existing procedures where Ficoll-Paque PLUS is used, enabling the isolation of human mononuclear cells from bone marrow, peripheral blood, and umbilical cord blood.

Ficoll-Paque PREMIUM 1.084 and Ficoll-Paque PREMIUM 1.073 (see page 16), have densities of 1.084 ± 0.001 g/ml and 1.073 ± 0.001 g/ml, respectively. They are both manufactured within a quality management system certified to ISO13485 as for the Ficoll-Paque PREMIUM product. These newer density products were launched in December 2007.

Ficoll-Paque PREMIUM 1.073 can be used when isolating lower density human mononuclear cells, for example mesenchymal stromal/stem cells or monocytes. The higher density lymphocytes and granulocytes will sediment through Ficoll-Paque PREMIUM 1.073 to the bottom of the tube, thereby enriching the lower density cells at the interface. Ficoll-Paque PREMIUM 1.073 has been found superior to Ficoll-Paque PREMIUM for isolating mesenchymal stem cells from human bone marrow (56).

Ficoll-Paque PREMIUM 1,084 can be used for preparation of cell fractions including higher density human mononuclear cells or for isolating lymphocytes that form rosettes with autologous red blood cells (15). It can also be used for separating blood cells from mice and rats, since the lymphocytes in rodents have a slightly higher average density than lymphocytes in humans (50, 51).

02

Ficoll-Paque density gradient media

Ficoll-Paque media products are aqueous solutions containing Ficoll PM400 and sodium diatrizoate with calcium disodium ethylenediamine-tetraacetic acid. Ficoll PM400 is a synthetic high molecular weight (M_r 400 000) polymer of sucrose and epichlorohydrin which is readily soluble in water. The molecules of Ficoll PM400 are highly branched, approximately spherical, and compactly coiled with a Stokes' radius of about 10 nm. Ficoll PM400 has a low intrinsic viscosity (17 ml/g) compared with linear polysaccharides of the same molecular weight (cf. dextran M_r 400 000:intrinsic viscosity (η)/49 ml/g) and solutions of Ficoll PM400 have low osmotic pressures.

Sodium diatrizoate is a convenient compound to use with Ficoll PM400 since it forms solutions of low viscosity with high density. Sodium diatrizoate (M_r 635.92) is the sodium salt of 3,5-diacetamido-2,4,6-triodobenzoic acid.

Since sodium diatrizoate is light-sensitive, all Ficoll-Paque products must be stored protected from light. The function of sodium diatrizoate in Ficoll-Paque products is to provide the optimal density and osmolarity necessary for the efficient removal of other cells from the mononuclear cells.

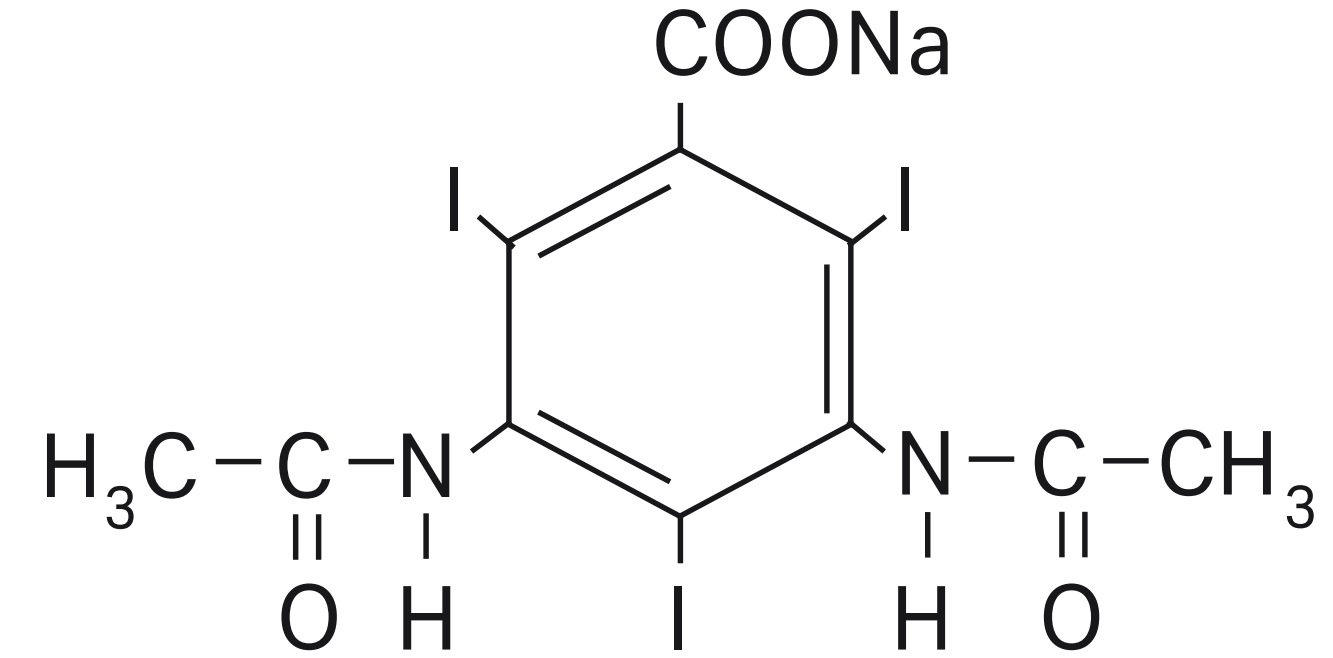
Ficoll-Paque products are supplied as sterile solutions in a bottle with a rubber septum closure. To maintain sterility, aseptic techniques should be used when withdrawing solution and the rubber septum closure should not be removed.

The separation principle

Mononuclear cell isolation using Ficoll-Paque separation media is based on methodology established through the extensive studies of Bøyum (1, 2, 3) and investigations carried out in our own laboratories.

Defibrinated or anticoagulant-treated blood is diluted with an equal volume of a balanced salt solution, layered carefully over Ficoll-Paque product (without intermixing) and centrifuged for 30 to 40 min. Differential migration of cells during centrifugation results in the formation of layers containing different cell types:

- The bottom layer contains erythrocytes, which have been aggregated by Ficoll PM400 and therefore sediment completely in the Ficoll-Paque density gradient media layer
- The layer immediately above the erythrocyte layer contains mostly granulocytes, which at the osmotic pressure of the Ficoll-Paque media solution, attain a density high enough to migrate through the Ficoll Paque media layer
- At the interface between the plasma and the Ficoll-Paque layer, mononuclear cells are found together with other slowly sedimenting particles (e.g., platelets) with low density



Several factors contribute to the success of this separation. On centrifugation, cells in the blood sample sediment towards the blood/Ficoll-Paque media interface, where they come in contact with the Ficoll PM400 present in Ficoll-Paque products. Red blood cells are efficiently aggregated by this agent at room temperature. Aggregation increases the rate of sedimentation of the red cells, which rapidly collect as a pellet at the bottom of the tube, where they are well separated from mononuclear cells. Granulocytes also sediment to the bottom of the Ficoll-Paque media layer. This process is facilitated by an increase in their densities caused by contact with the slightly hypertonic Ficoll-Paque media. Thus, on completion of centrifugation, both granulocytes and red blood cells are found at the bottom of the tube, beneath the Ficoll-Paque product.

Lymphocytes, monocytes, and platelets are not dense enough to penetrate into the Ficoll-Paque media layers having densities of 1.077 and 1.084 g/ml. These cells therefore collect as a concentrated band at the interface between the original blood sample and the Ficoll-Paque products having densities of 1.077 and 1.084 g/ml. This banding enables the mononuclear cells to be recovered with high yield in a small volume with little mixing with the Ficoll-Paque media. When Ficoll-Paque PREMIUM 1.073 is used, some lymphocytes with densities >1.073 will enter the Ficoll-Paque media layer and the resulting cell preparation will be enriched for lower density cells like mesenchymal stromal/stem cells and monocytes. Washing and centrifuging the harvested cells subsequently removes platelets, any contaminating Ficoll-Paque media, and plasma. The resulting cell suspension then contains highly purified, viable lymphocytes, monocytes, mesenchymal stromal/stem cells, and is suitable for further studies.

A recommended standard method

Cell separation using Ficoll-Paque products can be carried out over a wide range of blood sample volumes. With its high yield, this method can be adapted to the processing of very small amounts of blood, such as may be obtained from children. For maximum reproducibility of separation it is recommended that a standardized procedure be used. The following procedure has been evaluated in our laboratories with Ficoll-Paque PLUS and is recommended for separation of normal blood samples. Simple changes can easily be made to suit a particular centrifugation system. The same procedure is recommended when separating cells using Ficoll-Paque PREMIUM, Ficoll-Paque PREMIUM 1.084, and Ficoll-Paque PREMIUM 1.073.

To standardize the technique, blood volume and diameter of the centrifuge tube should be chosen first. These factors determine the height of the blood sample in the tube and consequently the centrifugation time. Increasing the height of the blood sample in the tube increases red cell contamination. The separation is, however, not appreciably affected by changing the diameter of the tube. Hence a larger volume can be separated with the same degree of purification in a tube of larger diameter if the height of the blood sample in the tube and the separation time are kept constant.

The yield and degree of purity of the mononuclear cells depend to a considerable extent on the efficiency of red cell removal.

When erythrocytes in whole blood are aggregated, some mononuclear cells are trapped in the clumps and therefore sediment with the erythrocytes. This tendency to trap mononuclear cells is reduced by diluting the blood. Dilution gives a better yield of mononuclear cells and reduces the size of the red cell clumps. Aggregation of erythrocytes is enhanced at higher temperatures (37°C), which consequently decreases the yield of mononuclear cells. At lower temperatures (4°C); however, the rate of aggregation is decreased but the time of separation is increased, which also decreases the yield of mononuclear cells. A compromise temperature of 18°C to 20°C gives optimal results.

Preparation of reagents

Diluent and washing solution

The balanced salt solution for dilution of the blood and cell washing can be made according to the instruction below. Other diluents and washing fluids such as isotonic Ca²⁺/Mg²⁺ free phosphate buffered saline (e.g. Dulbecco's PBS), salt solutions (e.g., Hank's) or cell culture media (e.g., RPMI 1640) may also be used.

Balanced salt solution

To prepare the balanced salt solution, mix one volume stock solution A with nine volumes stock solution B and sterilize. At least 20 ml for each sample should be processed. Other sterile standard salt solutions may also be used.

Ficoll-Paque product

Warm the Ficoll-Paque density gradient media to 18°C to 20°C before use. For samples larger than 3 ml, see Notes on page 11.

Equipment and solutions required but not provided

1. Sterile balanced salt solution or other standard salt solutions (see *Preparation of reagents*).
 2. Centrifuge with swing-out rotor (brake should be off).
 3. Sterile centrifuge tubes and pipettes.
 4. Sterile needles and syringes.
 5. Red blood cell lysis solution of choice (if isolating granulocytes).

Solution A		Concentration (g/l)
Anhydrous D-glucose	5.5 × 10 ⁻³ M (0.1%)	1.0
CaCl ₂ ·2H ₂ O	5.0 × 10 ⁻⁵ M	0.0074
MgCl ₂ ·6H ₂ O	9.8 × 10 ⁻⁴ M	0.1992
KCl	5.4 × 10 ⁻³ M	0.4026
TRIS	0.145 M	17.565
Dissolve in approximately 950 ml of distilled water and add concentrated HCl until the pH is 7.6 before adjusting the volume to 1 l.		

Solution B		Concentration (g/l)
NaCl	0.14 M	8.19
To prepare the balanced salt solution, mix 1 volume of solution A with 9 volumes of solution B. Prepare the solution freshly each week. Other standard salt solutions may be used.		

Preparation of the sample

Fresh blood should be used to ensure high viability of isolated mononuclear cells. Prepare the sample at 18°C to 20°C.

1.

To a 10 ml centrifuge tube add 2 ml of defibrinated- or anticoagulant-treated blood and an equal volume of balanced salt solution (final volume 4 ml).
2.

Mix the blood and buffer by inverting the tube several times or by drawing the mixture in and out of a pipette.

Procedure for isolation of mononuclear cells

1.

Invert the Ficoll-Paque media bottle several times to ensure thorough mixing.

For withdrawal of Ficoll-Paque media by syringe:
Snap-off the polypropylene cap. Disinfect the stoppers with an alcohol swab by rubbing the stopper firmly for several seconds and allow them to dry prior to use. Insert the syringe needle through the septum (Fig 1).

For withdrawal of Ficoll-Paque media by pipette:
Remove the snap-off polypropylene cap. Lift the aluminum ring. Pull off the metal seal. Remove the silver ring. Remove the rubber closure. Using aseptic techniques, withdraw the required volume of Ficoll-Paque media.
2.

Add Ficoll-Paque media (3 ml) to the centrifuge tube.
3.

Carefully layer the diluted blood sample (4 ml) onto the Ficoll-Paque media solution (Fig 3).
Important: When layering the sample do not mix the Ficoll-Paque media solution and the diluted blood sample.
4.

Centrifuge at 400 g for 30 to 40 min at 18°C to 20°C (brake should be turned off).
5.

Draw off the upper layer containing plasma and platelets using a sterile pipette, leaving the mononuclear cell layer undisturbed at the interface (Fig 4 and Fig 5). The upper layer, which contains the plasma, may be saved for later use.
6.

Transfer the layer of mononuclear cells to a sterile centrifuge tube using a sterile pipette.



Fig 1.



Blood sample
Ficoll-Paque media

Fig 3.

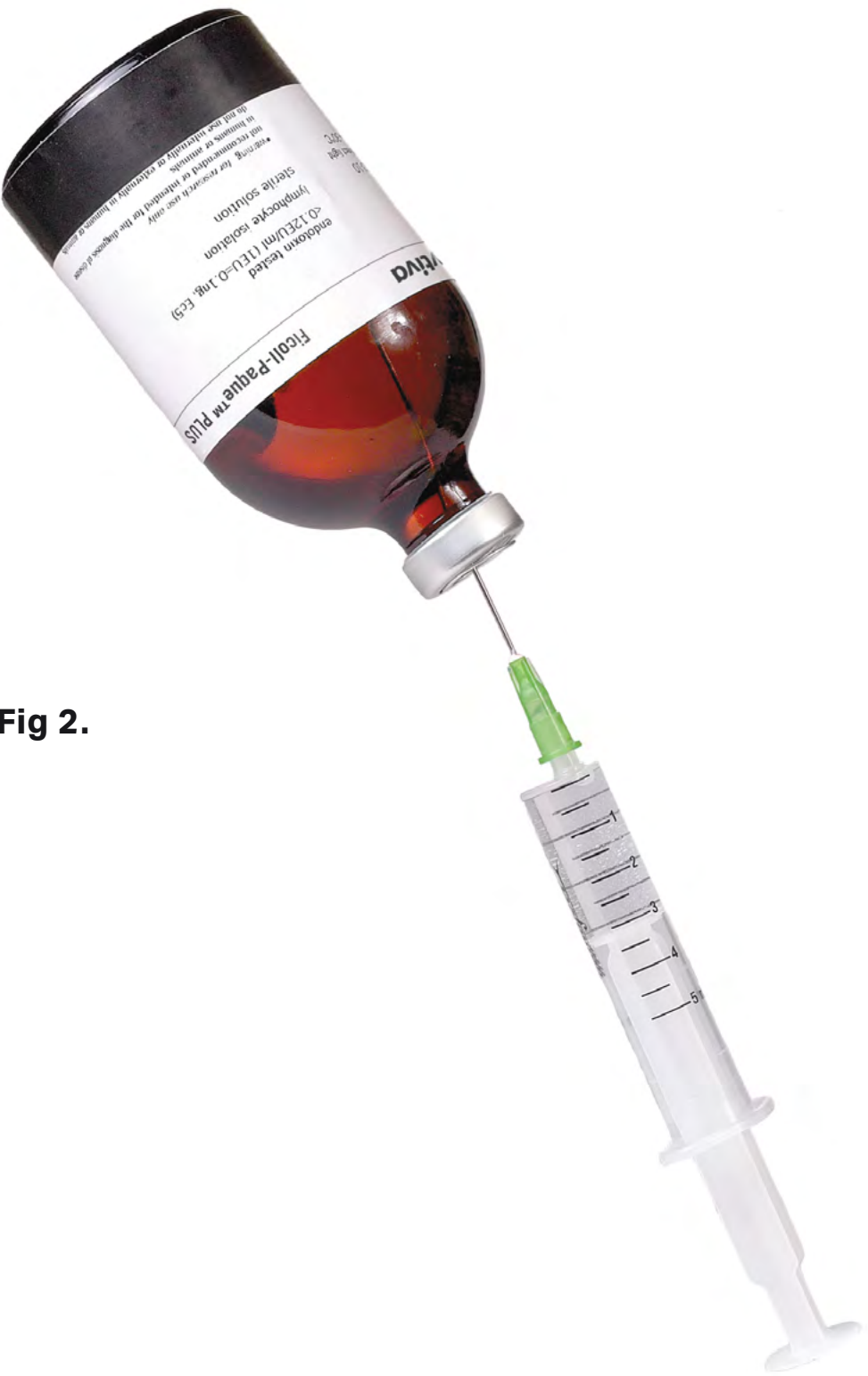


Fig 2.



Plasma
Mononuclear cells
Ficoll-Paque media
Granulocytes, Erythrocytes

Fig 4.



Upper layer removed
Mononuclear cells
Ficoll-Paque media
Granulocytes, Erythrocytes

Fig 5.

Washing the cell isolate

1. Estimate the volume of the transferred mononuclear cells. Add at least 3 volumes (~ 6 ml) of balanced salt solution to the mononuclear cells in the centrifuge tube.
2. Suspend the cells by gently drawing them in and out of a pipette.
3. Centrifuge at 400 to $500 \times g$ for 10 to 15 min at 18°C to 20°C .

Note: A centrifugation at high speed increases the mononuclear cell recovery. However, if it is important to also get rid of platelets a lower centrifugation speed is recommended (60 to $100 \times g$).

4. Remove the supernatant.
5. Resuspend the mononuclear cells in 6 to 8 ml balanced salt solution.
6. Centrifuge at 400 to $500 \times g$ (or 60 to $100 \times g$ for removal of platelets) for 10 min at 18°C to 20°C .
7. Remove the supernatant.
8. Resuspend the cell pellet in media appropriate for the application.

Procedure for isolation of granulocytes

1. Perform density gradient centrifugation using Ficoll-Paque media as described to the right in Procedure for isolation of mononuclear cells, steps 1 to 6.
2. Draw off the upper layer of Ficoll-Paque media using a sterile pipette, leaving the white cell layer of granulocytes above the red blood cell layer undisturbed.
3. Collect the thin white cell layer of granulocytes with a pipette and transfer to a sterile centrifuge tube.
4. Resuspend the cells in at least five volumes of balanced salt solution and centrifuge at $400 \times g$ for 15 min.
5. Lyse remaining red blood cells with any red blood cell lysis solution of choice.
6. Centrifuge the granulocytes at 400 to $500 \times g$ for 10 to 15 min at 18°C to 20°C .
7. Remove the supernatant.
8. Resuspend the granulocytes in 6 to 8 ml balanced salt solution.
9. Centrifuge at 400 to $500 \times g$ for 10 min at 18°C to 20°C .
10. Remove the supernatant.
11. Resuspend the cell pellet in media appropriate for the application.

Notes

Anticoagulants: Heparin, EDTA, citrate, acid citrate dextrose (ACD), and citrate phosphate dextrose (CPD) may be used as anticoagulants for the blood sample. Defibrinated blood requires no anticoagulant. Defibrination, however, results in a lower mononuclear cell yield and may cause increased contamination by red cells (3). It also causes selective loss of monocytes. Bøyum has found that a slightly purer mononuclear cell preparation is obtained using EDTA instead of heparin as anticoagulant (3). It has also been noted in the purification of mononuclear cells from sources other than peripheral blood that addition of heparin may cause gelling of cell suspensions (4). Citrate-stabilized blood may result in better quality RNA and DNA than other anticoagulants, and produce a higher yield of mononuclear cells. Heparin affects T-cell proliferation and binds to many proteins. EDTA is good for DNA based assays, but influences Mg^{2+} concentration and causes problems for cytogenetic analysis (5). It has also been shown that the RNA yield is higher in EDTA-blood than in heparin-blood (6).

Blood volume: Larger volumes of blood may be processed with the same efficiency of separation by using centrifuge tubes of increased diameter while maintaining approximately the same heights of Ficoll-Paque media (2.4 cm) and blood sample (3.0 cm) as in the standard method described above. Increasing the tube diameter does not affect the separation time required.

Blood sample storage: Blood samples should be free of clots and processed as soon as possible after collection to ensure optimal results. Delays in processing the blood can result in loss of viability, lower cell recoveries and more contaminating granulocytes and/or erythrocytes. Indeed, storage for 24 h at room temperature has been reported to result in reduced lymphocyte yield, altered expression of surface markers, and reduced response to mitogenic stimulation (5, 7, 8).

Dead cell removal: Dead cells are removed during cell isolation using Ficoll-Paque PLUS (9, 10, 58).

Density and temperature: The temperature at which density gradient separations are carried out are naturally affected by room temperature, centrifuge temperature, temperature of the density gradient media, and temperature of the liquid sample. There can, at times, also be confusion regarding what is meant by room temperature (e.g., in Europe it can be 20°C and in North America > 20°C). It is known that the densities of Ficoll-Paque products decrease with increase in temperature. For instance Ficoll-Paque PREMIUM (1.077 g/ml) exhibits a density of 1.0772, 1.0767, and 1.0758 at 20°C, 22°C, and 25°C, respectively.

Pathological blood samples: The standard method described above has been developed for the purification of mononuclear cells from peripheral blood of normal, healthy, human donors. Different results may be obtained with samples taken from donors with infections or other pathological conditions, such as cancer (see *Further applications of Ficoll-Paque PLUS and Ficoll-Paque PREMIUM*, page 15).

Platelet removal: If it is important to remove all platelets from the mononuclear cell fraction a second centrifugation in a 4% to 20% sucrose gradient layered over Ficoll-Paque PLUS can be applied. This procedure will effectively remove any platelet contamination (11). Platelets will remain at the top of the sucrose gradient and mononuclear cells will sediment through the sucrose gradient to the top of the Ficoll-Paque PLUS layer. Alternatively, the platelets may be removed by aggregation with adenosine-5'-diphosphate (ADP) before separating the mononuclear cells (12).

Troubleshooting inadequate performance

If used according to the recommended standard procedure, Ficoll-Paque PLUS and Ficoll-Paque PREMIUM may be expected to give trouble-free isolation of human peripheral blood mononuclear cells with results as shown on page 4. As mentioned earlier, Ficoll-Paque PREMIUM 1.073 and Ficoll-Paque PREMIUM 1.084 will lead to isolation of cell preparations having slightly different density subsets of mononuclear cells. However, deviations in certain experimental parameters may lead to poor results and the troubleshooting chart given here is intended to assist in the rapid identification and correction of the problem causing reduced performance.

Deviation in performance	Likely source of problem	Comments
Increased red blood cell and contamination of the lymphocytes.	A. Temperature too low.	The densities of all Ficoll-Paque products are greater at low temperature (see <i>Notes</i> , Density and temperature) and red blood cells are aggregated less well, so granulocytes and red blood cells are prevented from entering the Ficoll-Paque media layer. Raise the temperature to 18°C to 20°C.
	B. Centrifugation speed too slow and/or centrifugation time too short.	Adequate time and g-force must be used to ensure complete sedimentation of non-lymphoid cells.
Low yield and viability of mononuclear cells.	Temperature too high.	Ficoll-Paque products are less dense at high temperature and some lymphocytes may penetrate into the Ficoll-Paque media layer. Cell viability may also be affected. Reduce the temperature to 18°C to 20°C.
Low yield of mononuclear cells with normal viability.	Blood not diluted 1:1 with balanced salt solution; unusually high hematocrit.	The high cell density results in large numbers of lymphocytes being trapped by red blood cell aggregates. Dilute the blood sample further.
Low yield of mononuclear cells with increased granulocyte contamination.	Vibration of the centrifuge rotor, leading to stirring of the gradient.	Vibration may cause broadening of the mononuclear cell band and mixing with the underlying cells. Check that the rotor is properly balanced. Choose rotor speed to avoid natural resonant frequencies.
Low yield of mononuclear cells, low viability, and contamination by other cell types.	Sample contains cells with abnormal densities; densities different from those in normal human blood.	May be encountered with pathological blood samples, non-human blood samples, old blood samples, or samples from sources other than peripheral blood. Percoll™ PLUS, a medium for density gradient centrifugation, may be more suitable than Ficoll-Paque products for such separations.

Ficoll-Paque PLUS

Ficoll-Paque PLUS consists of a mixture of Ficoll PM400 and sodium diatrizoate at a density of 1.077 g/ml, which is optimized for the isolation of mononuclear cells from human peripheral blood. It has also been used in many other applications as described below. For over 30 years it has been trusted to deliver consistent performance and reproducible results in research laboratories. Ficoll-Paque PLUS as well as the Ficoll-Paque PREMIUM products are sterile. For the purpose of ensuring sterility, all Ficoll-Paque products are subject to terminal moist heat sterilization.

Ficoll-Paque PREMIUM products

Ficoll-Paque PREMIUM (density 1.077 g/ml) is a density gradient media used as a reagent in cell therapy applications for the preparation of human mononuclear cells. Based on Ficoll-Paque PLUS, it was developed for both large- and small-scale purification of mononuclear cells from human peripheral blood using a simple and rapid centrifugation technique developed by Bøyum (1). Ficoll-Paque PREMIUM 1.084 and Ficoll-Paque PREMIUM 1.073, have densities of 1.084 ± 0.001 g/ml and 1.073 ± 0.001 g/ml, respectively. These may be used when higher or lower densities than the standard 1.077 g/ml are required.

The methodology for using Ficoll-Paque PREMIUM 1.084 and Ficoll-Paque PREMIUM 1.073 is the same as for all Ficoll-Paque products described earlier. Ficoll-Paque PREMIUM 1.084 can be used for preparation of cell fractions including higher-density human mononuclear cells, and for separating blood cells from mice or rats. Ficoll-Paque PREMIUM 1.073 can be used when preparing lower density mononuclear cells, for example mesenchymal stromal cells or monocytes. The higher density lymphocytes and granulocytes will sediment through Ficoll-Paque PREMIUM 1.073 to the bottom of the tube, thereby enriching the lower density cells at the interface.

Ficoll-Paque PREMIUM products differ from Ficoll-Paque PLUS in that they are manufactured in a controlled environment within a quality management system certified to ISO 13485 and meet USP <1043> 'ancillary materials for cell, gene, and tissue-engineered products', within the responsibilities applicable to a supplier (1). For our customers this means a high quality assurance on product validation and documentation of manufacturing procedures.

Ficoll-Paque PREMIUM products offer the following benefits:

- Manufactured within a quality management system certified to ISO 13485
- Manufactured in accordance to USP <1043> recommendations for ancillary materials
- Sterile (moist heat sterilized), ready-to-use reagent
- Low levels of endotoxin (< 0.12 EU/ml) secured and tested
- Osmolality tested
- RSF (Regulatory Support File) available as support for clinical applications to regulatory agencies
- No cytotoxicity according to USP Method <87> and ISO 10993-5 guidelines

Properties of mononuclear cells isolated by the Ficoll-Paque method

Since its introduction in 1968, the mononuclear cell separation method described by Bøyum (1, 2) has been used in numerous immunological investigations. This widespread adoption indicates the superior results obtained with this technique and its freedom from impairment of lymphocyte function. Nevertheless, certain effects of the separation procedure have been seen and these are noted below, since research situations may arise in which they are of significance.

Separation with Ficoll-Paque PLUS has been reported to lead to adsorption of cytophilic IgG to the mononuclear leukocytes (13), resulting in erroneously high estimates of the number of Ig-bearing lymphocytes and too low estimates of the number of cells bearing Fc receptors. This interference can be avoided by washing the blood cells with balanced salt solution before isolation, thus removing the IgG present in the plasma that gives rise to these artifacts.

Selective loss of a population of lymphocytes that form rosettes with autologous red blood cells has been reported to occur using the standard procedure (14, 15) and evidence was found that this is the result of a specific lymphocyte-red blood cell interaction, not a non-specific trapping (15). This population was found to account for about 6% of the lymphocytes initially present in the blood sample and could be recovered almost quantitatively by resuspending the red cell pellet in medium and recentrifuging over a gradient of slightly higher density than normal (i.e. 1.083 g/ml) (15).

Slight differences in phenotype compositions have been seen between fresh samples of whole blood and samples purified using Ficoll-Paque PLUS (16, 17). Fresh samples of whole blood show lower percentages of CD4+, CD19+, and CD4+CD45RA+ cells, and higher percentages of CD8+ and CD4+CD29+ subsets than fresh lymphocyte purified on a Ficoll-Paque PLUS gradient (15).

Ficoll-Paque density gradient separation (FDS) was associated with significantly higher percentages of CD3+/CD62L+ and CD3+/CD11b+ lymphocytes in young children and adults alike, while the percentages of CD3+/CD54+ cells from adults was not affected by FDS. The percent expression of CD54, CD62L, and CD11b on T cells from both children and adults were significantly higher following FDS, with a greater increase in CD11b expression on T cells from young children, reaching adult levels (16).

Lymphocytes separated by the Bøyum procedure have been reported (18) to show enhanced stimulation in mixed lymphocyte cultures as compared with lymphocytes in "leukocyte-rich plasma" (not exposed to Ficoll-Paque PLUS). This enhanced reaction was postulated to depend at least partially on the removal in the Ficoll-Paque PLUS method of neutrophils that appear otherwise to have a suppressive effect on the mixed lymphocyte reaction (18).

Further applications of Ficoll-Paque PLUS and Ficoll-Paque PREMIUM

A great many modifications and extensions of the method have come into use following the introduction of the technique described by Bøyum in 1968 and its subsequent widespread adoption. For example, monocytes (which are recovered in the mononuclear cell fraction, using the standard procedure described in this booklet) can be removed, if desired, by incubating the blood sample with iron (or iron carbonyl) before separation on Ficoll-Paque PLUS. The monocytes phagocytose the iron particles and become denser, with the result that they sediment through the Ficoll-Paque PLUS layer on centrifugation and collect in the red blood cell pellet at the bottom of the tube (3).

An important and widely used extension of the original technique is its application, in combination with selective rosetting (clustering), to the isolation of lymphocyte subclasses. Selective rosetting is presented in RosetteSep™ products marketed by StemCell Technologies Inc. (19, 20, 21, 59). These products contain tetrameric antibody complexes (TAC) that crosslink unwanted cells to multiple red blood cells present in the sample, forming immunorosettes. Upon centrifugation over Ficoll-Paque product, the immunorosettes sediment to the bottom of the tube together with the red blood cells. Alternatively, the purified lymphocytes obtained by the standard procedure (with or without monocyte removal) can be incubated with an excess of sheep red blood cells (ratio of red blood cells to lymphocytes at least 50:1), whereupon the T lymphocytes spontaneously form rosettes with the sheep red blood cells. On centrifugation for a second time over Ficoll-Paque PLUS, the T lymphocyte rosettes sediment to the bottom of the tube together with the excess red blood cells, leaving the other (non-rosetting) lymphocytes at the interface (3).

Such techniques for the separation of lymphocyte subclasses, as well as the standard method for isolating the entire lymphocyte population, have been widely applied to studies of lymphocyte functions and surface markers in disease states as compared to normal controls. Caution is, however, necessary in applying the Ficoll-Paque PLUS technique to pathological blood specimens, since it has been found that the resulting mononuclear cell layer may be contaminated with immature granulocytes in patients with certain infections (22), and particularly cancer (9, 23). In the latter case, elevated numbers of monocytes may also be present (24).

Ficoll-Paque PLUS has been used with success to separate cells from a variety of sources other than peripheral blood, even though its properties have been optimized specifically for blood mononuclear cell isolation. Thus, separation over

Ficoll-Paque PLUS facilitated detection and identification of malignant cells in abdominal and pleural fluids (23, 25). Separation on Ficoll-Paque PLUS has also been reported to assist in establishing cultures of amniotic fluid cells and to facilitate their subsequent cytogenic analysis (26). Ficoll-Paque PLUS and Ficoll-Paque PREMIUM have also been used for the isolation of cord blood derived hematopoietic stem cells, cord blood derived pluripotent stem cells for regenerative medicine clinical applications, and bone marrow mesenchymal stem cells (27, 28, 60, 61). Ficoll-Paque products are also suitable for granulocyte isolation (29, 30, 31).

Ficoll-Paque PLUS can also be used to isolate mononuclear cells from species other than man. In some cases (e.g. cow, goat, and rabbit) it may be necessary to alter the standard procedure to achieve good results (3) and it should be remembered that the density of Ficoll-Paque PLUS (1.077 g/ml), although optimized for the isolation of human mononuclear cells, may not give optimal yield and purity of mononuclear cells from other species. However, isolation methods using Ficoll-Paque PLUS for preparation of mononuclear cells from peripheral blood (PB) and bone marrow (BM) have been described for mouse ((32(PB), 33 (BM)), dog (34 (PB), 35(BM)), monkey (36 [PB], 37 [BM]), cow (38 [PB], 39 [PB]), rabbit (36 [BM], 40 [PB]), horse (41 [PB]), pig (42 [PB]), and even fish (43). Where it is desired to work with solutions of densities other than 1.077 g/ml it may be convenient to use Ficoll-Paque PREMIUM 1.084, Ficoll-Paque PREMIUM 1.073, or the alternative centrifugation media Percoll/Percoll PLUS since iso-osmotic solutions of different densities are very easily prepared with these media, facilitating the optimization of a particular separation. In research studies, separation with Percoll has also been reported to give improved lymphocyte yields and purities in some cases (44 to 47).

Applications using Ficoll-Paque PREMIUM 1.084

Ficoll-Paque PREMIUM 1.084 can be used for isolating higher density cells. It has for instance been shown that the lymphocytes in rodents have a slightly higher average density than lymphocytes in humans (50, 51) and a fraction of these will therefore be lost into the pellet after a standard 1.077 g/ml density gradient centrifugation, contaminating the granulocyte layer and decreasing the mononuclear cell recovery. Dog, bovine, rabbit, mouse, and rat blood cells have successfully been separated using a 1.084 g/ml density gradient media such as Percoll or Ficoll-diatrizoate solutions (52, 53).

It has been reported that selective loss of lymphocytes that form rosettes with autologous blood cells (as described above) may occur by the standard procedure using a 1.077 g/ml Ficoll-Paque products (14, 15) and these lymphocytes have been shown to be recovered almost quantitatively by resuspending the red cell pellet in medium and recentrifuging over a gradient of slightly higher density than normal (i.e., 1.083 g/ml) (15). Thus, Ficoll-Paque PREMIUM 1.084 could potentially be used for this application, although this remains to be shown.

Applications using Ficoll-Paque PREMIUM 1.073

Ficoll-Paque PREMIUM 1.073 can be used for enriching lower density cells such as monocytes from peripheral blood, or mesenchymal stem cells from bone marrow or placenta (54, 55, 56, 57).

Grisendi *et al.* directly compared Ficoll-Paque PREMIUM and Ficoll-Paque PREMIUM 1.073 for isolating and expanding mesenchymal stem cells from human bone marrow aspirate and convincingly demonstrated that density gradient separation using the lower density Ficoll-Paque PREMIUM 1.073 was associated with an enrichment of mesenchymal stem cell (MSC) subtypes characterized by a higher proliferation potential. Thus, Ficoll-Paque PREMIUM 1.073 can ultimately benefit clinical applications based on these cells. Brooke *et al.* recently described a successful manufacturing process for isolating and expanding placenta-derived human MSC using Ficoll-Paque PREMIUM 1.073 for a clinical trial (57).

Availability and storage

Ficoll-Paque PLUS and Ficoll-Paque PREMIUM are available in packs of 6 × 100 ml (code numbers 17-1440-02 and 17-5442-02 respectively) and 6 × 500 ml (code numbers 17-1440-03 and 17-5442-03 respectively) as sterile (moist heat sterilized), ready-to-use liquids in glass bottles with rubber septum closures. The Ficoll-Paque PREMIUM products of different densities, Ficoll-Paque PREMIUM 1.084 and Ficoll-Paque PREMIUM 1.073, are available in packs of 6 × 100 ml (code numbers 17-5446-02 and 17-5446-52 respectively). The different densities of each Ficoll-Paque product is clearly reflected in the colors of the polypropylene caps where density 1.077 g/ml is shown in blue, density 1.084 g/ml in red, and density 1.073 g/ml in gray.

Full instructions for use are included with each pack. Ficoll-Paque products should be stored between 4°C and 30°C protected from light, under which conditions they will maintain sterility and stability for 3 yr.

03

References

- Isolation of mononuclear cells and gr anulocytes from human blood. (Paper IV). Bøyum, A., *Scand. J., Clin. Lab. Invest. 21 Suppl*, **97**, 77–89 (1968).
- Isolation of leucocytes from human blood – further observations. (Paper II). Bøyum, A., *Scand. J. Clin. Lab. Invest. 21 Suppl*, **97**, 31–50 (1968).
- Isolation of lymphocytes, granulocytes and macrophages. Bøyum, A., *Scand. J., Immunol. 5 Suppl*, **5**, 9–15 (1976).
- Gel formation with leucocytes and heparin. Almeida, A.P., Beaven, M.A., *Life Sci*, **26**, 549–555 (1980).
- Biological sample collection and processing for molecular epidemiological studies. Holland, N.T., Smith, M.T., Eskenazi, B., Bastaki, M., *Mutation Research*, **543**, 217-234 (2003).
- Collection and storage of Human Blood Cells for mRNA Expression Profiling: A 15-Month Stability Study. Marteau, J., Mohr, S., le Pfister, M., Visvikis-Siest, S., *Clinical Chemistry*, **51**, 1250-1252 (2005).
- Altered lymphocyte markers and blastogenic responses associated with 24 hour delay in processing of blood samples. Kaplan, J., Nolan, D., Ree, A. J., *Immunol. Methods*, **50**, 187–191 (1982).
- Stability of hematological analytes depends on the hematology analyser used: A stability study with Bayer Advia 120, Beckman Coulter LH 750 and Sysmex XE 2100. Imeri, F., Herklotz, R., Risch, L., Arbetsleitner, C., Zerlauth, M., Risch, G.M., Huber, A.R., *Clinica Chimica Acta*, **397**, 68-71 (2008).
- How to optimize multiparameter flow cytometry for leukaemia/lymphoma diagnosis. Paiette, E., *Best Practice & Research Clinical Haematology*, **16**, 671-683 (2003).
- Dendritic Cells Cross-Dressed with Peptide MHC Class I Complexes Prime CD8 T Cells. Dolan, B.P., Gibbs Jr, K.D., Ostrand-Rosenberg, S. J., *Immunol.*, **177**, 6018-6024 (2006).
- Purification of lymphocytes and platelets by gradient centrifugation. Perper, R.J., Zee, T.W., Mickelson, M.M., *J. Lab. Clin. Med.*, **72**, 842–848 (1968).
- Platelet aggregation technique used in the preparation of lymphocyte suspensions. Vives, J., Parra, M., Castillo, R., *Tissue Antigens*, **1**, 276–278 (1971).
- Quantitation of Fc receptors and surface immunoglobulin is affected by cell isolation procedures using Plasmagel and Ficoll-Hypaque. Alexander, E.L., Titus, J.A., Segal, D.M., *J. Immunol. Methods*, **22**, 263–272 (1978).
- Analysis of the lymphocyte distribution during Isopaque-Ficoll isolation of mononuclear cells from human peripheral blood. Hokland, P., Heron, I., *J. Immunol. Methods*, **32**, 31–39 (1980).
- The Isopaque-Ficoll method re-evaluated: Selective loss of autologous rosette-forming lymphocytes during isolation of mononuclear cells from human peripheral blood. Hokland, P., Heron, I., *Scand. J. Immunol*, **11**, 353–356 (1980).
- Lymphocyte immunnophenotyping by flow cytometry in normal adults Comparison of fresh whole blood lysis technique, Ficoll-Paque separation and cryopreservation. Romeu, M.A., Mestre, M., González, L., Valls, A., Verdaguer, J., Corominas, M., Bas, J., Massip, E., Buendia, E., *J. Immunol. Methods*, **154**, 7-10 (1992).
- Expression of adhesion molecules on T lymphocytes in young children and infants – a comparative study using whole blood lysis or density gradient separation. Lin, S.-J, Chao, H.-C, Yan, D.-C, Huang, Y.-J., *Clin. Lab. Haem*, **24**, 353-359 (2002).
- Reactivity in mixed cultures of mononuclear leucocytes separated on Ficoll-Hypaque. Bain, B., Pshyk, K. Proceedings 7th Leucocyte Culture Conference, (Ed. Daguiillard, F.), *Academic Press*, New York, 29–37 (1973).
- Membrane Surface Nanostructures and Adhesion Property of T lymphocytes Exploited by AFM. Wu, Y., Lu, H., Cai, J., He, X., Hu, Y., Zhao, H., Wang, X., *Nanoscale Res. Lett.*, **4**, 942-947 (2009).
- A role for interleukin-12/23 in the maturation of human natural killer and CD56 T cells in vivo. Guia, S., Cognet, C., de Beaucoudrey, L., Tessmer, M.S., Jouanguy, E., Berger, C., Filipe-Santos, O., Feinberg, J., Camcioglu, Y., Levy, J., Jumaah, S.A., Al-Hajjar, S., Stephan, J., Fieschi, C., Abel, L., Brossay, L., Casanova, J., Vivier, E., *J. Blood*, **111**, 5008-5016 (2008).
- Targeting NF- B activation via pharamacologic inhibition of IKK2-induced apoptosis of human acute myeloid leukemia cells. Frelin, C., Imbert, V., Griessinger, E., Peyron, A-C., Rochet, N., Philip, P., Dageville, C., Sirvent, A., Hummelsberger, M., Berard, E., Dreano, M., Sirvent, N., Peyron, *J. Blood*, **105**, 804-911 (2005).
- Ficoll-separated mononuclear cells from sepsis patients are contaminated with granulocytes. Van den Akker, E.L.T., Baan, C.C., Van den Berg, B., Russcher, H., Joosten, K., Hokken-Koelega, A.C.S., Lamberts, S.W.J., Koper, J.W., *Intensive Care Med*, **34**, 912-916 (2008).
- A novel technique for the enrichment of primary ovarian cancer cells. Chan, J.K., Hamilton, C.A., Anderson, E.M., Cheung, M.K., Baker, J, Husain, A., Teng, N.N., Kong, C.S., Negrin, R.S., *Am. J. Obstet. Gynecol.*, **197**, 507.e1—507.e5, (2007).
- Non-lymphoid cells obtained by the Bøyum technique and their significance in cancer patients. Kluin-Nelemans, J.C., van Helden, H.P.T., *J. Clin. Lab. Immunol.*, **4**, 99–102 (1980).
- Gradient separation of normal and malignant cells. II. Application to *in vivo* tumour diagnosis. Minami, R., Yokota, S., Teplitz, R.L., *Acta. Cytol.*, **22**, 584–588 (1978).
- Enhancement of human amniotic cell growth by Ficoll-Paque gradient fractionation. Chang, H-C., Jones, O.W., Bradshaw, C., *et al.*, *In Vitro.*, **17**, 81–90 (1981).
- Optimization of immunomagnetic separation for cord blood-derived hematopoietic stem cells. Kekarainen, T., Mannelin, S., Laine, J., Jaatinen, T., *BMC Cell Biology*, **7** (2006).
- Prolonged ex vivo culture of human bone marrow mesenchymal stem cells influences their supportive activity toward NOD/SCID-repopulating cells and committed progenitor cells of B lymphoid and myeloid lineages. Briquet, A., Dubois, S., Bekaert, S., Dolhet, M., Beguin, Y., Gothot, A., *Haematologica*, **95(1)**, 47-56 (2010).
- The active translation of MHCII mRNA during dendritic cells maturation supplies new molecules to the cell surface pool. Malanga, D., Barba, P., Harris, P.E., Maffei, A., Del Pozzo, G., *Cell. Immunol.*, **246**, 75-80 (2007).
- Reduced number and function of peripheral dendritic cells in celiac disease. Ciccocioppo, R., Ricci, G., Rovati, B., Pesce, I., Mazzocchi, S., Piancatelli, D., Cagnoni, A., Millimaggi, D., Danova, M., Corazza, G.R., *Clin. and Exp. Immunol.*, **146**, 487-496 (2007).
- Anti-proteinase 3 antibodies (c-ANCA) prime CD14-dependent leukocyte activation. Hattar, K., van Burck, S., Bickenbach, A., Grandel, U., Maus, U., Lohmeyer, J., Csernok, E., Hartung, T., Seeger, W., Grimminger, F., Sibelius, U., *J. Leukocytes Biol*, **78**, 992-1000 (2005).
- Modeling the initiation and progression of Human Acute Leukemia in Mice. Lubin, I., Faktorowich, Y., Lapidot, T., Gan, Y., Eshhar, Z., Gazit, E., Levite, M., Reisner, Y., *Science. New Series*, **252**, 427-431 (1991).
- Noncanonical Wnt11 Signaling Is Sufficient to Induce Cardiomyogenic Differentiation in Unfractionated Bone Marrow Mononuclear Cells. Flaherty, M.P., Abdel-Latif, A., Li, Q., Hunt, G., Ranjan, S., Ou, Q., Tang, X., Johnson, R.K., Bolli, R., Dawn, B., *Circulation*, **117**, 2241-2252 (2008).
- Double-label expression studies of prostacyclin synthase, thromboxane synthase and COX isoforms in normal aortic endothelium. Kawka, D.W., Ouellet, M., Hétu, P., Singerm I.I., Riendeau, D., *Biochimica et Biophysica Acta.*, **1771**, 45–54 (2007).
- Growth of bone marrow stromal cells on small intestinal submucosa: an alternative cell source for tissue engineered bladder. Zhang, Y., Lin, H., Frimberger, D., Epstein, R.B., Kropp, B.P., *BJU International*, **96**, 1120-1125 (2005).
- Expression of mRNA for multiple serotonin (5-HT) receptor types/subtypes by the peripheral blood mononuclear cells of rhesus macaques. Yang, G., Qiu, C., Zhao, H., Liu, Q., Shao, Y., *J. Neuroimmunol.*, **178**, 24-29 (2006).
- Functional Analysis of Neuron-like Cells Differentiated from Neural Stem Cells Derived from Bone Marrow Stroma Cells in vitro. Xu, R., Jiang, X., Guo, Z., Chen, J., Zou, Y., Ke, Y., Zhang, S., Li, Z., Cai, Y., Du, M., Qin, L., Tang, Y., Zeng, Y., *Cell Mol. Neurobiol.*, **28**, 545-558 (2008).
- The bovine lymphoid system: Binding and stimulation of peripheral blood lymphocytes by lectins. Pearson, T.W., Roelants, G.E., Lundin, L.B., *et al.*, *J. Immunol. Methods*, **26**, 271–282 (1979).
- Acid a-naphthyl acetate esterase: presence of activity in bovine and human T and B lymphocytes. Yang, T.J., Jantzen, P.A., Williams, L.F., *J. Immunol.*, **38**, 85–93 (1979).
- The effect of blue light exposure in an ocular melanoma animal model. Di Cesare, S., Maloney, S., Fernandes, B.F., Martins, C., Marshall, J., Anteck, E., Odashiro, A.N., Dawson, W.W., Buriner Jr, M.N., *J. Exp. and Clin. Cancer Res.*, **28:48**, 1-9 (2009).
- Comparative study of six methods for lymphocyte isolation from several mammalian sources and determination of their carbohydrate composition. Hueso, P., Rocha, M., *(Article in Spanish) Rev. Esp. Fisiol.*, **34**, 339–344 (1978).
- Effect of cryopreservation on IL-4, IFNc and IL-6 production of porcine peripheral blood lymphocytes. Li, X., Zhong, Z., Liang, S., Wang, X., Zhong, F., *Cryobiology*, **59**, 322-326 (2009).
- A comparison of the methods used for the separation of fish lymphocytes. Blaxhall, P.C., *J. Fish Biol*, **18**, 177–181 (1981).
- Separation of human peripheral blood monocytes on continuous density gradients of Polyvinylpyrrolidone-coated silica gel (Percoll). Brandslund, I., Møller-Rasmussen, J., Fisker, D., *et al.*, *J. Immunol. Methods*, **48**, 199–211 (1982).
- Efficient separation of human T lymphocytes from venous blood using PVP-coated colloidal silica particles (Percoll). Feucht, H.E., Hadam, M.R., Frank, F., *et al.*, *J. Immunol. Methods*, **38**, 43–51 (1980).
- An improved technique for the isolation of lymphocytes from small volumes of peripheral mouse blood. Mizobe, F., Martial, E., Colby-Germinario, S., *et al.*, *J. Immunol. Methods*, **48**, 269–279 (1982).
- Quantitative and Qualitative Comparative Analysis of Gradient Separated Hematopoietic Cells from Cord Blood and Chemotherapy-Mobilized Peripheral Blood. Sato, J., Kawano, Y.F., Takae, Y., Hirao, A., Makirnoto, A., Okarnoto, Y., Abe, T., Shimokawa, T., Iwai, A., Kurodaa, Y., *Stem Cells*, **13**, 548-555 (1995).
- EC Guide to GMP (Good Manufacturing Practice), annex 1 "Manufacture of Sterile Medicinal Products".
- United States Pharmacopeia. Recommendations for ancillary materials, chapter <1043>
- Lymphocyte differentiation in the rabbit thymus. Leene, W., *et al.*, *Ann. Immunol. (Paris)*, **127**, 911-921 (1976).
- Separation of leucocytes: improved cell purity by fine adjustments of gradient medium density and osmolality. Bøyum, A., *et al.*, *Scand. J. Immunol.*, **34**, 697-712 (1991).
- Isolation of bovine colostral lymphocytes: in vitro blastogenic responsiveness to concanavalin A and bovine rotavirus. Archambault, *et al.*, *Ann. Rech. Vet.*, **19**, 169-174 (1988).
- Characterization and species distribution of high affinity GTP-coupled receptors for human rantes and monocyte chemoattractant protein. Van Riper *et al.*, *JEM*, **177**, 851-856 (1993).
- Cryopreserved mesenchymal stromal cell treatment is safe and feasible for severe dilated ischemic cardiomyopathy. Chin *et al.*, *Cytotherapy*, **12**, 31-37 (2010),
- Mesenchymal stem cells from multiple myeloma patients display distinct genomic profile as compared with those from normal donors, Garayoa *et al.*, *Leukemia*, **23**, 1515-1527 (2009).
- GMP-manufactured density gradient media for optimized mesenchymal stromal/stem cell isolation and expansion, Grisendi *et al.*, *Cytotherapy*, **12**, 466-477 (2010).
- Manufacturing of human placenta-derived mesenchymal stem cells for clinical trials. Brooke *et al.*, *British J. of Haematology*, **144**, 571-579 (2008).
- High gradient magnetic cell separation with MACS, Miltenyi *et al.*, *Cytometry*, **11**, 231-238 (1990).
- Reduced expression of ATP-binding cassette transporter G1 increases cholesterol accumulation in macrophages of patients with type 2 diabetes mellitus. Mauldin, J. P. *et al.*, *Circulation*, **117**, 2785-2792 (2008).
- Defined serum-free culturing conditions for neural tissue engineering of human cord blood stem cells. Hamad A. *et al.*, *Acta. Neurobiol. Exp.*, **69**, 11-23 (2009).
- High purity and yield of natural Tregs from cord blood using a single step selection method, Figueroa-Tentori *et al.*, *J. Immunol. Methods*, **339**, 228-235 (2008).

04

Ordering Information

Selected products for cell science from Cytiva

Product		Quantity	Code no.
Ficoll-Paque PREMIUM (density: 1.077 g/ml)	Intended for the preparation of human mononuclear cells from blood and bone marrow for clinical applications	6 × 100 ml	17-5442-02
		6 × 500 ml	17-5442-03
Ficoll-Paque PREMIUM 1.073 (density: 1.073 g/ml)	For enrichment of lower density human mononuclear cells like mesenchymal stromal/stem cells or monocytes in the preparation of cells for clinical applications	6 × 100 ml	17-5446-52
Ficoll-Paque PREMIUM 1.084 (density: 1.084 g/ml)	For isolating higher density human mononuclear cells in the preparation of cells for clinical applications and for separating blood cells from mice or rats	6 × 100 ml	17-5446-02
Ficoll-Paque PLUS	A density gradient medium for isolation of lymphocytes for research use from whole human peripheral blood	6 × 100 ml	17-1440-02
		6 × 500 ml	17-1440-03
Ficoll PM400	A hydrophilic sucrose polymer of high molecular weight for the preparation of density gradient centrifugation media for research use	100 g	17-0300-10
		500 g	17-0300-50
Percoll	A PVP coated, silica-based centrifugation medium for the isolation of cells, subcellular particles, and viruses for research use	250 ml	17-0891-02
		1 l	17-0891-01
Percoll PLUS	A silane-coated, silica-based centrifugation medium for the isolation of various cell types for clinical research applications	250 ml	17-5445-02
		1 l	17-5445-01

These and other products are available in the [cytiva.com/contact](https://www.cytiva.com/contact) catalog and in individual technical booklets, which are available free on request.

cytiva.com

Cytiva and the Drop logo are trademarks of Life Sciences IP Holdings Corporation or an affiliate doing business as Cytiva.

Ficoll, Ficoll-Paque and Percoll are trademarks of Global Life Sciences Solutions USA LLC or an affiliate doing business as Cytiva. Any other trademarks are the property of their respective owners.

© 2025 Cytiva

All goods and services are sold subject to the terms and conditions of sale of the supplying company operating within the Cytiva business. A copy of those terms and conditions is available on request. Contact your local Cytiva representative for the most current information.

For local office contact information, visit cytiva.com/contact

CY13988-16Sep25-HB

