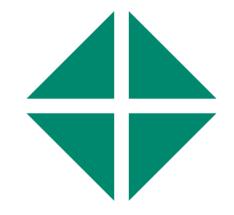


# **UNICORN** start 1.3

**User Manual** 



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# 1 Introduction

# About this user manual

This user manual provides the instructions needed to install and operate UNICORN $^{\text{TM}}$  start 1.3, and analyze chromatography run data.

# In this chapter

Section		See page
1.1	Important user information	6
1.2	User documentation	7
1.3	UNICORN start 1.3 overview	8

# 1.1 Important user information

# Purpose of UNICORN start 1.3 User Manual

The purpose of this user manual is to provide a guide describing system control, method editor and result evaluation functions of UNICORN start 1.3.

The manual covers the following topics:

- · System control
- · Create and edit methods
- View, evaluate and present results
- · Administration and database management
- Troubleshooting

Refer to the ÄKTA™ start Operating Instructions (29027057) for further information on how to perform a chromatography run.

**Note:** The UNICORN start 1.3 User Manual does not describe all functions or every

command in all panes and dialogs of the user interface. Refer to the UNICORN start 1.3 Online Help for more information about commands that

are not described in this manual.

### Notes and tips

**Note:** A Note is used to indicate information that is important for trouble-free and

optimal use of the product.

**Tip:** A tip contains useful information that can improve or optimize your

procedures.

## **Typographical conventions**

Software items are identified in the text by **bold italic** text. A colon separates menu levels, thus  $File \rightarrow Open$  refers to the Open command in the File menu.

Hardware items are identified in the text by **bold** text (e.g., **Power** switch).

Text entries that UNICORN start 1.3 generates or that the user must type are represented by a monotype type face (e.g.,  $C:\$  Program Files (x86) \Cytiva \UNICORN\UNICORN start 1.3).

### **Prerequisites**

The following prerequisites must be fulfilled before you can use this manual the way it is intended:

- You need to have a general understanding of how your PC/laptop and Microsoft® Windows® work.
- You need to understand the general concepts of liquid chromatography.

# 1.2 User documentation

### **User documentation**

From the *Help* menu in UNICORN start 1.3 or on the UNICORN start 1.3 DVD, the following user documentation is available.

Document	Main contents
UNICORN start 1.3 User Manual	Overview and detailed descriptions of the different features in UNICORN start 1.3. Instructions on how to use the software. Workflow descriptions for common operations.
	Dialog descriptions and user instructions, context-linked from all software features.
UNICORN start 1.3 Online Help	Dialog descriptions for UNICORN start 1.3 (accessible from the <i>Help</i> menu only).

The user documentation listed in the table below is available on the ÄKTA start User Documentation CD. A display help is available from the Instrument Display.

Documentation	Main contents
ÄKTA start Operating Instructions	Instructions needed to install, operate and maintain ÄKTA start in a safe way.
ÄKTA start Display Help	Dialog descriptions of the functionality menu for ÄKTA start (only accessible from the Instrument Display).
ÄKTA start Maintenance Manual	Detailed instrument and module descriptions, including instructions needed to maintain and troubleshoot ÄKTA start.
ÄKTA start System Cue Card	A condensed guide to prepare and run chromatographic techniques on ÄKTA start.
ÄKTA start Maintenance Cue Card	A condensed guide to handling routine maintenance operations and troubleshooting ÄKTA start.

# 1.3 UNICORN start 1.3 overview

UNICORN start 1.3 is a complete software package for:

- Control, editing methods and monitoring chromatography runs on ÄKTA start.
- Evaluation and analysis of the results generated from ÄKTA start.

# Illustration of the system

The illustration below shows the  $\ddot{\text{A}}$ KTA start System with UNICORN start 1.3 installed on a computer.



Part	Description
1	ÄKTA start (instrument).
2	Frac30 (Fraction collector).
3	UNICORN start 1.3 (software installed on a computer).

## **UNICORN start 1.3 modules**

The four modules of UNICORN start 1.3 and their main features are listed below.

Module	Features
System Control	Provides an intuitive and easy-to-use interface to control ÄKTA start.
	Performs and monitors quick start or user defined chromatography method runs.
	Performs and monitors chromatography runs from pre- defined or user-defined methods.
	Performs system performance method runs.

Module	Features
Method Editor	Provides the flexibility to automate the chromatography runs.
	Allows creation of methods from predefined chromatography templates like <i>Affinity</i> , <i>Gel Filtration</i> , <i>Ion Exchange</i> and <i>Desalting</i> .
	Gives flexibility to create a customized method by dragging and dropping the chromatography phases such as <i>Prime and Equilibration</i> , <i>Sample Application</i> , <i>Elution and Fractionation</i> , etc.
	Allows exporting methods to a USB memory stick for importing them into ÄKTA start.
Evaluation	Allows viewing and presentation of results, including creation of PDF reports.
	Allows various evaluation operations on curves and chromatograms including comparison, peak integration etc.
	Allows importing results from ÄKTA start and exporting results to other formats.
Administration	Allows administration of the UNICORN start 1.3 database for backup, restore, archive and retrieve operations.
	Allows reviewing of UNICORN start 1.3 and system logs.

# 2 Installation of UNICORN start 1.3

# In this chapter

Section		See page
2.1	Computer requirements	11
2.2	Language and regional settings for UNICORN start on Windows 10	13
2.3	Installation overview	17
2.4	Additional software installed by UNICORN start 1.3	18
2.5	Installation procedure	20
2.6	Uninstall UNICORN start 1.3	44
2.7	Removing additional software components	47

# 2.1 Computer requirements

# **Installation requirements**

Before starting the installation procedure, make sure that the following requirements are met for the computer (PC or laptop):



#### **IMPORTANT**

UNICORN start 1.3 is tested using an US-English operating system version. **Using other language versions of the operating system may cause errors.** 



#### **IMPORTANT**

Changing the default font and font size on Windows may cause problems in the UNICORN start 1.3 user interface.



#### **IMPORTANT**

A screen resolution of 1280x1024 or higher is recommended. Parts of the UNICORN start 1.3 user interface may not be displayed properly using a lower specification.



#### **IMPORTANT**

Windows power save features should be turned off to avoid conflicts with system operations.

Component	Specification
Operating System	Windows 10 Professional, Enterprise & Educational x64 (English US).
RAM	4 GB (minimum)
Minimum free disk space	12 GB (minimum)
Display	<ul><li>Laptop: 14" (minimum)</li><li>Desktop: 17" (minimum)</li></ul>

Component	Specification
Processor	Laptop: Dual Core with 3 GHz clock speed (minimum)     Desktop: Dual Core with 2.5 GHz clock speed (minimum)
USB port	• 3.0 (recommended) • 2.0
Other	DVD drive     Network interface card

#### Note:

- UNICORN start 1.3 is not tested for operation in a mixed Windows XP/ Windows 7/Windows 10 environment. It has only been tested and verified with Windows 10.
- If UNICORN 7 is installed on the computer then UNICORN start 1.3 installation might not be successful and might not function correctly.
- UNICORN start 1.3 is tested using US-English operating system version.
   Using other language versions of the operating system may cause errors.
   For more information, please refer to Section 2.2 Language and regional settings for UNICORN start on Windows 10, on page 13.
- A screen resolution of 1280x1024 or higher is recommended. Parts of the UNICORN start 1.3 user interface may not be displayed properly using a lower specification.
- Changing the default font and font size on Windows may cause problems in the UNICORN start 1.3 user interface.
- The Windows basic color scheme is recommended.
- Windows power save features should be turned off to avoid conflicts with system operations.
- UNICORN start 1.3 is not compatible with the Windows 10 feature High DPI Awareness which allows the graphical user interface to be scaled. The interface scale must remain at 100% scale to avoid issues with clipping and misalignment of parts of the UNICORN user interface:
  - 1. On the desktop, right-click UNICORN start, and click Properties.
  - 2. On the Compatibility tab, click Change high DPI settings.
  - 3. Select the **Override high DPI scaling behavior** check box.
  - 4. In the Scaling performed by list, click System (Enhanced).
  - 5. Click **OK**, and then **Apply**.
  - 6. Right-click UNICORN start and click Run as administrator.

# 2.2 Language and regional settings for UNICORN start on Windows 10

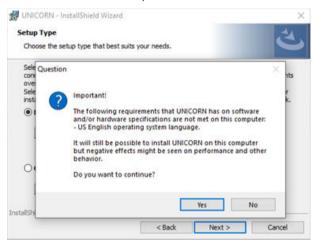
#### Information

For Windows 10 versions, e.g. 1803, or higher, (see *note on page 16*, for how to check Windows version) there have been changes in how culture and language settings are handled. This might cause problems on UNICORN start 1.3, and previous versions. To prevent any unwanted behavior, make sure that the culture format is set to "English (United States)" for the Current User, Welcome screen, and New user accounts. If the "English (United States)" language pack is not available on the computer, it must be installed (see below).

Note:

A warning dialog pop-up message may appear during installation of UNICORN start that the language is not set correctly.

If the settings are correct, the warning message can be ignored, and the installation can proceed as normal.



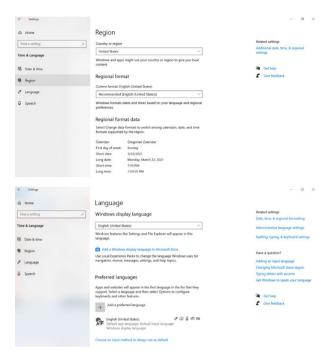
# How to fix language problem for UNICORN start 1.3

# **Update culture settings**

 Change language and region to "English (United States)", see below, for examples. If English (United States) is not available, the language pack needs to be installed first. See *Install language pack*, on page 16.

#### 2 Installation of UNICORN start 1.3

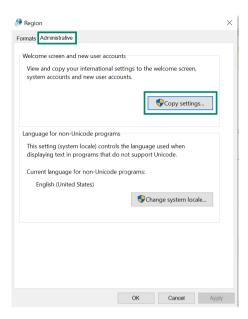
2.2 Language and regional settings for UNICORN start on Windows 10



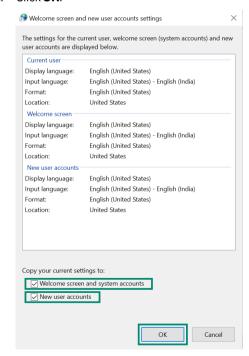
- 2. Set culture format to "English (United States)" and specify that the newly set language applies to all new users and the system user.
  - a. Go to Control Panel.
  - b. In the View by drop-down box, select Small icons.
  - c. Click Region.



- d. Select Format "English (United States)".
- e. Click the Administrative tab.
- f. Click Copy settings....



- g. Check Welcome screen and system accounts.
- h. Check New user accounts.
- i. Click OK.



# Install language pack

#### Offline, manually:

1. Download the offline language pack from the Microsoft official website.

**Note:** The language pack must be for the correct Windows version (e.g. 1803, or higher).

 Install the language pack running the command "lpksetup" (press Windows key + R).

### Online, via System Settings:

Add the language from:

Settings  $\rightarrow$  Time & Language  $\rightarrow$  Region & Language  $\rightarrow$  Add Language (Windows 10 1803),

or

Settings → Time & Language → Language → Add Language (Windows 10 1809).

**Note:** To check the Windows version, type "**run**" in the Windows search bar, then type "**winver**" in the Run App.

# 2.3 Installation overview

# **Installation summary**

The table below provides an overview of the UNICORN start 1.3 installation procedure for setting up a UNICORN start 1.3 workstation.

Stage	Description	
1	Installation of additional software necessary to run UNICORN start 1.3.	
	See Section 2.4 Additional software installed by UNICORN start 1.3, on page 18.	
2	Installation of UNICORN start 1.3.	
	See Section 2.5 Installation procedure, on page 20.	

# Do not copy the DVD-ROM or decompress the files



#### **IMPORTANT**

Do not run the installation from a network drive.



#### **IMPORTANT**

Do not attempt to decompress the files using any other file decompression utility.

UNICORN start 1.3 is supplied on a DVD-ROM. Files on the DVD-ROM are compressed and the installation cannot be performed by simply copying the files onto the local hard drive. However, the installation files can be copied to a hard drive and the installation may be run from the hard drive instead. Do not run the installation from a network drive.

During the installation procedure, the required folder structure is created on the hard drive and the files are decompressed. Do not attempt to decompress the files using any other file decompression utility.

# 2.4 Additional software installed by UNICORN start 1.3

## **Required software**

The following additional software will be installed, if not found, before the installation of UNICORN start 1.3 begins:

- MadCap™ HelpViewer
- Microsoft .NET Framework 3.5 SP1
- Microsoft .NET Framework 4.0
- Microsoft .NET Framework 4.7 Full
- Microsoft Visual C++® 2008 SP1 Redistributable Package (x86)
- Microsoft Visual C++ 2010 SP1 Redistributable Package (x86)
- Microsoft Visual C++ 2010 SP1 Redistributable Package (x64)
- Microsoft Visual C++ 2012 Redistributable Package (x86)
- Microsoft Visual C++ 2012 Redistributable Package (x64)
- Microsoft Visual C++ 2013 Redistributable Package (x86) or (x64)
- Microsoft Core XML Services (MSXML) 6.0
- Microsoft SQL Server®
- Microsoft SQL Server 2019 Express SP3 (x64)
- Windows Installer 4.5 for Windows Vista and Server 2008 (x86)
- · Windows Driver Package Cytiva Sweden AB

The installation will be initiated automatically by the UNICORN start 1.3 installation program. Any of the listed software that is already installed will not be included in this installation step. You may need to restart the computer several times to proceed from one application installation to the next.

All software listed above is already installed on computers delivered from Cytiva.

Note:

By default, the software listed above is installed on the computer C:\ drive. However, you can manually change to another computer drive e.g., D:\ drive.

If the available space on the drive is insufficient, the installation may fail and the installation program will attempt to repeat the installation after each restart. Make sure that enough space is available on the drive before starting the installation. The actual required space will vary depending on what is previously installed on the computer, and subsequent selections in the UNICORN start 1.3 installation. However, the installation program needs a total of 12 GB of free space to initiate the installation.

## Required disc space



#### **IMPORTANT**

Make sure that enough space is available on the drive before starting the installation.

The installation program needs 12 GB of free space to initiate the installation. If the available space on the drive is insufficient, the installation may fail and the installation program will attempt to repeat the installation after each restart.

Make sure that enough space is available on the drive before starting the installation. The actual required space will vary depending on what is previously installed on the computer, and subsequent selections in the UNICORN start 1.3 installation.

## The help viewer application

As part of the UNICORN start 1.3 installation, a special viewer for the UNICORN start 1.3 Online Help is also installed. This application, the *MadCap HelpViewer*, is accessed from the UNICORN start 1.3 user interface either by clicking the help buttons in the dialogs, by pressing the **F1** key or by selecting *Help* from the menu bar.

# 2.5 Installation procedure

# In this section

Section		See page
2.5.1	Install UNICORN start 1.3	21
2.5.2	Configure e-license	30
2.5.3	Upgrade UNICORN start	36

## 2.5.1 Install UNICORN start 1.3

#### Introduction



#### **IMPORTANT**

Make sure that enough space is available on the drive before starting the installation.

The installation of UNICORN start 1.3 includes all software components necessary to operate UNICORN start 1.3 and a connected ÄKTA start instrument.

The components that are installed are described in the table below.

Software component	Characteristics
UNICORN start 1.3 client	UNICORN client software for the workstation, including the four UNICORN start 1.3 modules: <b>Administration</b> , <b>Method Editor</b> , <b>System Control</b> and <b>Evaluation</b> .
UNICORN start 1.3 database	An instance of Microsoft SQL Server Express Edition that stores the results and methods created by the UNICORN start 1.3 modules.
Instrument server	A Windows service necessary to control and manage an instrument that is connected to the workstation.

#### Instructions



#### **IMPORTANT**

Make sure that enough space is available on the drive before starting the installation.

To install UNICORN start 1.3, go through the following steps:

#### Step Action

- Depending on how you have purchased the **UNICORN start** software, do one of the following:
  - a. Download the software from the ESD portal by logging in with your activation ID or username and password.
     or

**b.** Insert the installation DVD in the DVD drive.

The UNICORN start 1.3 installation starts.

#### Tip:

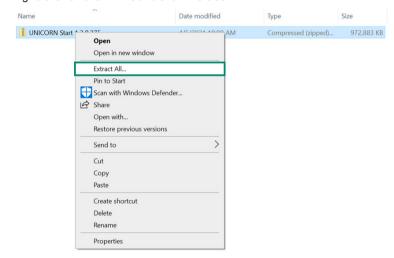
If the DVD drive is not set up to allow automatic start, browse the DVD contents and double click the file Setup.exe to initiate the installation.



#### **IMPORTANT**

Do not click on the .msi file to start the installation. The .msi file does not check the pre-required software for the installation to succeed.

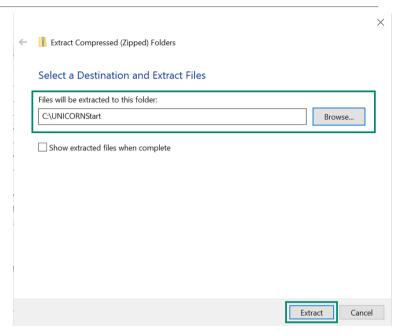
- 2 If installing from a ZIP file:
  - a. Right-click on the ZIP file and click Extract All....



b. Select a destination in C: \UNICORNStart to avoid SQL path complexity and click *Extract*.

#### Note:

The user needs to create the UNICORNStart folder.



c. Navigate to the folder and click the UNICORNstartinstaller:

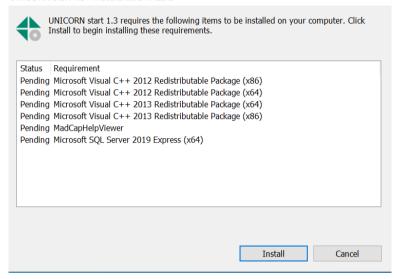


If you see this warning message, click the relevant **Yes** or **No** option:



If there are any required software components to be installed (as mentioned in Section 2.4 Additional software installed by UNICORN start 1.3, on page 18), the following dialog appears:

UNICORN start 1.3 - InstallShield Wizard

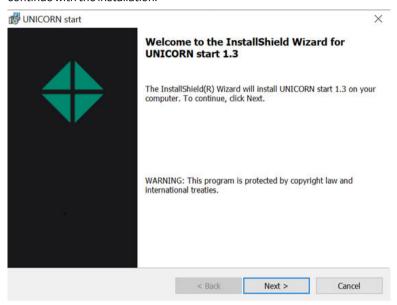


#### Note:

Multiple restarts of the computer may occur during the installation process of these software components.

If the installation does not start automatically after a restart, please manually restart the installation as described in step 1.

The **UNICORN start 1.3 InstallShield Wizard** window opens. Click **Next** to continue with the installation.



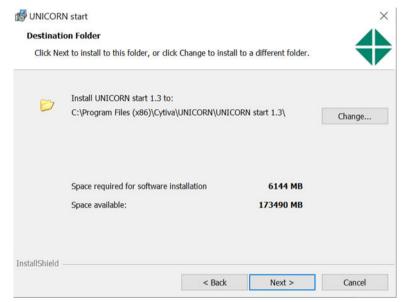
- 5 Read the license agreement carefully.
  - a. Select I accept the terms in the license agreement.
  - b. Click Next to proceed.



6 Click **Next** to install UNICORN start 1.3 in the default folder, or click **Change** to install in a different folder.

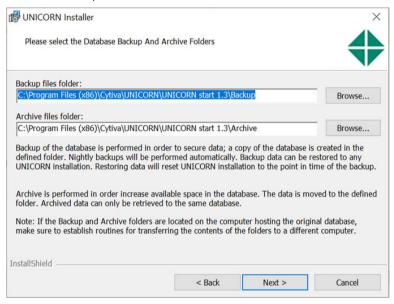
#### Note:

By default, UNICORN start 1.3 suggests C:\Program Files (x86)\Cytiva\UNICORN\.



7 **a.** Enter locations for **Backup** and **Archive** folders.

#### b. Click Next to proceed.



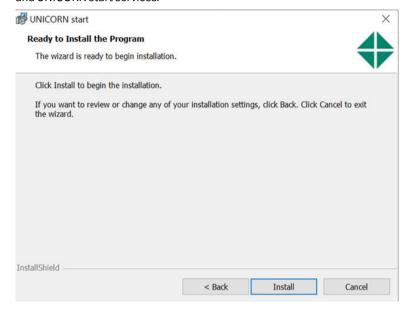
#### Note:

To ensure data safety, it is recommended that the backup and archive folders are copied or moved at regular intervals to another server computer or some other storage media.

#### Tip:

The UNICORN Configuration Manager is located at Start → Programs → Cytiva. This dialog can be used to edit the backup and archive information after the installation.

Click Install to begin installation of UNICORN start 1.3, ÄKTA start driver, and UNICORN start services.

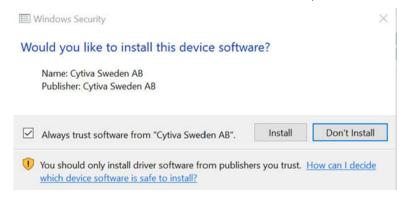


#### Note:

The SQL Server software is installed on the C:\ drive by default. If the available space for this is insufficient, the installation may fail. The required space varies depending on what is previously installed on the computer but at least 2 GB should be available to ensure that the installation is completed correctly.

#### Note:

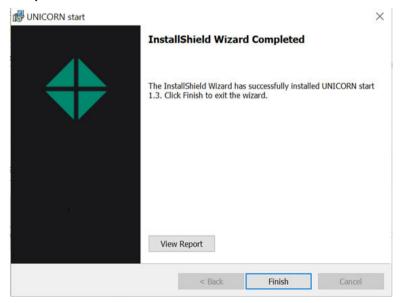
If a Windows Security warning is displayed when the ÄKTA start USB communication driver is installed, click the **Install** button to proceed.



9 Proceed by configuring the e-license, see Section 2.5.2 Configure e-license, .



# 10 Click the *Finish* button in the *UNICORN start InstallShield Wizard Completed* window.



#### Note:

The installation report can be viewed by clicking the **View Report** button.

# 2.5.2 Configure e-license

#### Introduction



#### **IMPORTANT**

Make sure that your PC/laptop is connected to the internet before initiating e-license configuration.

After completing a UNICORN start 1.3 installation, the software prepares to configure an e-license for UNICORN start 1.3.

Note:

Make sure that your PC/laptop is connected to the internet before initiating e-license configuration. An internet connection needs to be established to access the Cytiva e-licensing web site. For details about retrieving an e-license, see Generate a UNICORN license file, on page 31.

## Initiate license configuration

Follow the instructions below to perform the final installation step and initiate the elicense configuration.

#### Step Action

1 In the Configure e-License dialog:



Click the Copy to Clipboard button to copy the Ethernet address.

2.5.2 Configure e-license

Step	Action

#### Note:

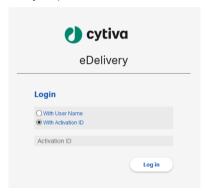
UNICORN start 1.3 attempts to select the right Ethernet address from the available set of network interface cards. If you anticipate the selected network configuration to change in this computer, please choose a more permanent Ethernet address from the list.

2 Click the hyperlink in the dialog to proceed to the e-licensing web site.

### Generate a UNICORN license file

## Step Action

- 1 Go to the web address http://www.cytiva.com/eDelivery.
- For easy access, we recommend you login using the **Activation ID** delivered with your purchase.



3 Open the License Activation & Entitlements tab and click List Entitlements.



- 2.5 Installation procedure
- 2.5.2 Configure e-license

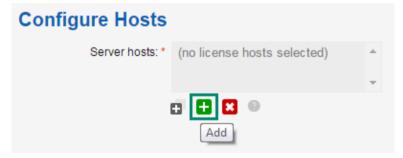
4 Select your entitlement in the list, open the **Actions** tab and click **Activate**.



5 Enter values for fields marked with \* and click **Next**.



6 Under **Configure Hosts**, click ■.



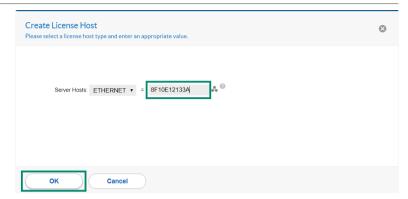
#### Result:

The Create License Host window appears.

7 Enter your Ethernet ID (physical address) in the **Server Hosts** field. Use no spaces or dashes, just the numbers and letters. For example: 8F10E12133A. Then click **OK**.

#### Note:

Use the MAC address of the Ethernet port.



#### Result:

This populates the **Server hosts** field.

8 Click Next.



9 If the **Activation ID** has more licenses, enter the number of licenses to generate under **Fulfill count** and click **Next**.



- 2.5 Installation procedure
- 2.5.2 Configure e-license

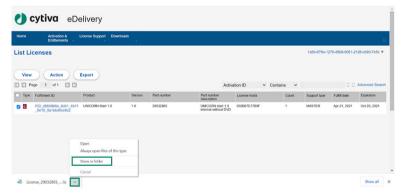
10 Review the license information details and then click **Generate**.



11 Select your product in the list, click **View**, and select **Save To File** or **Email license**.



12 If saving to file, the license file may appear at the bottom left of your browser. Click the arrow to open the drop-down list and click **Show in folder**.



13 Cut and paste the license file to a dedicated e-license folder on your computer.

### Locate and connect the e-license

The table below describes how to connect and confirm the e-license in the UNICORN start 1.3 **Configure e-License** dialog.

- a. Click the **Browse** button to locate the license file and
  - **b.** Click the *Open* button to add the search path to the file in the *e-License Configuration* dialog.



2 Click the **Configure e-License** button.

#### Result:

A confirmation dialog opens, showing that the configuration has been successful.

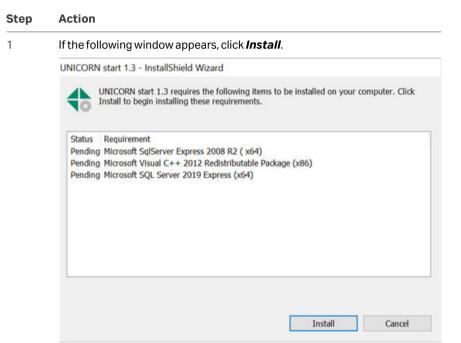
3 Click OK in the confirmation dialog, and Close in the e-License Configuration dialog to complete the process.

# 2.5.3 Upgrade UNICORN start

#### Introduction

The UNICORN start installation program will automatically identify if a previous installation is present on the computer when you attempt to install UNICORN start. You can choose between canceling the upgrade or upgrading to the newer version.

## **Upgrade instructions**



#### Note:

The computer may restart multiple times during the installation process of these software components.

If the installation does not start automatically after a restart, please manually restart the installation.

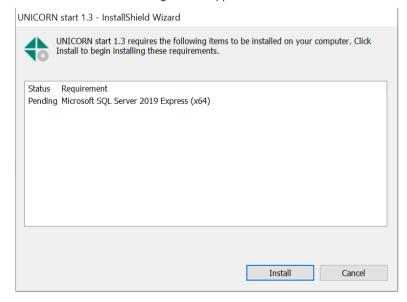
2 Make sure that a backup of the old UNICORN start 1.2 database has been made. If no backup has been made, all data will be lost. Click **Yes** to continue the installation.



3 Click **Yes** and wait for the system to restart.



4 After the restart, the following window appears. Click *Install*.

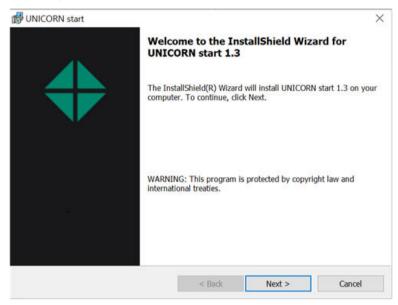


- 2.5 Installation procedure
- 2.5.3 Upgrade UNICORN start

Result:

The InstallShield Wizard for UNICORN start 1.3 window appears.

5 Click **Next**.



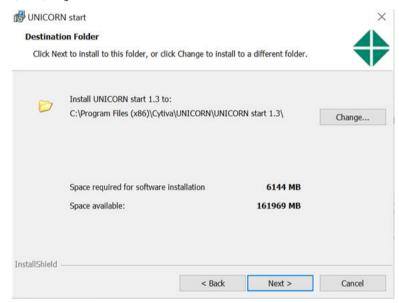
To proceed, select I accept the terms in the license agreement and click Next.



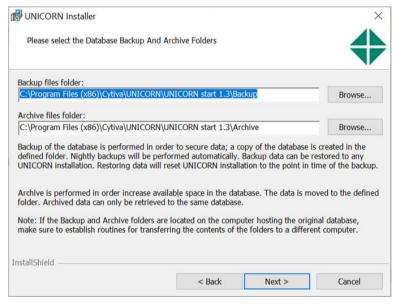
7 To install UNICORN start 1.3 in the default folder, click **Next**. To install in a different folder, click **Change**.

#### Note:

By default, UNICORN start 1.3 suggests C:\Program Files (x86)\Cytiva\UNICORN.



8 Make sure locations for backup and archive folders are correct. Then click Next to proceed.

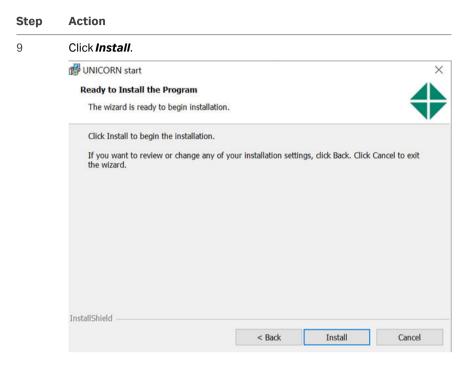


#### Note:

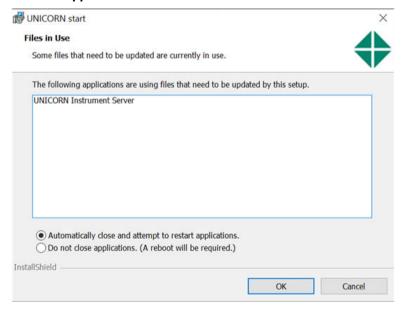
To ensure data safety, it is recommended that the backup and archive folders are copied or moved at regular intervals to another server computer or some other storage media.

#### Tip:

The UNICORN Configuration Manager is located at Start →Programs →Cytiva. This dialog can be used to edit the backup and archive information after the installation.

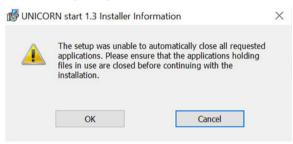


10 If the following window appears, select Automatically close and attempt to restart applications and then click OK.



- 2.5 Installation procedure
- 2.5.3 Upgrade UNICORN start

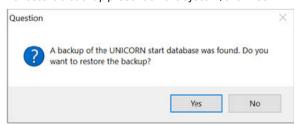
11 If the following dialog box appears, click **OK**.



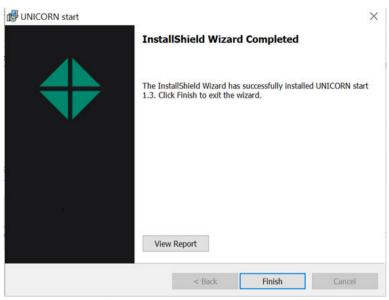
- 12 Click **Yes** in the **UNICORN start 1.3** information message.
- 13 Configure the e-License. See Section 2.5.2 Configure e-license, on page 30 for more information.



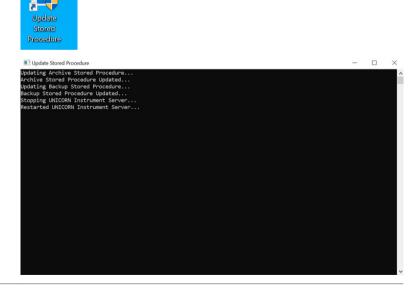
To restore a backup present on the system, click **Yes**.



#### 15 Click *Finish*.



16 Run the *Update Stored Procedure.exe* icon from the desktop.



## 2.6 Uninstall UNICORN start 1.3

#### Introduction

This section describes how to remove UNICORN start 1.3 using the installation program. UNICORN start 1.3 can also be removed using the Windows *Add or Remove Programs* dialog.

#### Remove the installation

To uninstall UNICORN start 1.3, follow the steps described below:

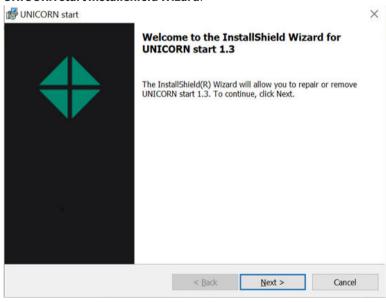
Note:

Before removing UNICORN start 1.3, a backup must be made, otherwise all data will be lost if proceeding. For instructions about how to back up data, see Section 9.2.2 Database maintenance, on page 226.

#### Step Action

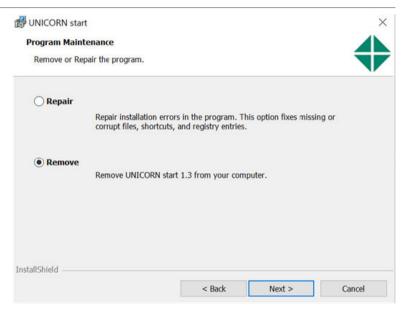
Open the UNICORN start 1.3 installation program **Setup.exe**. This will open the

#### UNICORN start InstallShield Wizard.

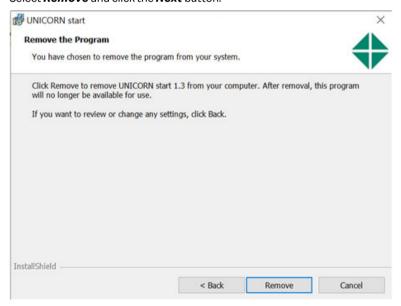


2 Click the **Next** button.

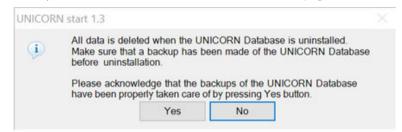
UNICORN start Program Maintenance dialog opens.



#### 3 Select **Remove** and click the **Next** button.

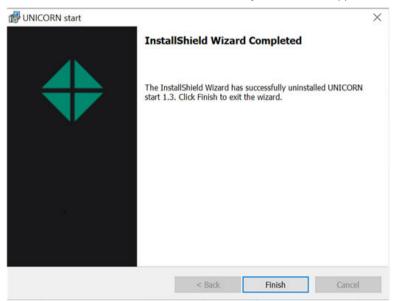


Make sure that a backup of the database has been made. If no backup has been made, all data will be lost if proceeding. For instructions about how to back up data, see Section 9.2.2 Database maintenance, on page 226.



5 Click the **Yes** button to proceed.

The UNICORN start InstallShield Wizard Completed window appears.



6 Click the **Finish** button to complete the uninstallation.

#### Note:

After removing UNICORN start 1.3, some parts will still remain in the installation folder, for example logs and database backups. If desired, these objects must be removed manually.

# 2.7 Removing additional software components

#### Introduction

Some software components required to operate UNICORN cannot be removed automatically by the installation wizard. This is because the same components may be used for other purposes as well as for UNICORN start 1.3. The components may also have been updated after the original installation, and UNICORN start 1.3 can no longer determine if they were part of the installation package. If any of the components listed below are not in use by other applications, they need to be removed manually:

- MadCap HelpViewer
- Microsoft .NET Framework 3.5 SP1
- Microsoft .NET Framework 4.0
- Microsoft .NET Framework 4.7 Full
- Microsoft Visual C++ 2008 SP1 Redistributable Package (x86)
- Microsoft Visual C++ 2010 SP1 Redistributable Package (x86)
- Microsoft Visual C++ 2010 SP1 Redistributable Package (x64)
- Microsoft Visual C++ 2012 Redistributable Package (x86)
- Microsoft Visual C++ 2012 Redistributable Package (x64)
- Microsoft Visual C++ 2013 Redistributable Package (x86) or (x64)
- Microsoft Core XML Services (MSXML) 6.0
- Microsoft SQL Server 2008 R2 Express SP2 (x86) or (x64)
- Microsoft SQL Server 2019 Express SP3 (x86) or (x64)
- Windows Installer 4.5 for Windows Vista and Server 2008 (x86)
- · Windows Driver Package Cytiva Sweden AB

#### Note:

The UNICORN start 1.3 instance of the Microsoft SQL Server needs to be uninstalled manually. Follow the instructions described in the table below to uninstall the Microsoft SQL Server.

# Remove additional software components

The table below describes how to remove the components manually.

Step	Action
1	Open the Windows <b>Control Panel</b> .
2	Double-click the <i>Add or Remove Programs</i> icon.  Result: The <i>Add or Remove Programs</i> dialog opens. All programs installed on the
	client computer are listed.

Step	Action
3	Select the software component from the list and click the <b>Change/Remove</b> button.
4	Follow the instructions in the dialogs that opens, until the software is removed.

#### **Un-installation of SQL-Server 2019**

- 1. Uninstall Microsoft SQL Server 2019 (64-bit) from the Control Panel.
- 2. Delete the Microsoft SQL Server folder from Program Files and Program Files (x86).

**Note:** Make sure there is no SQL Server installed on the PC before deleting these two folders.

# 3 Getting started

# In this chapter

Section		See page
3.1	Launch UNICORN start 1.3	50
3.2	Help functions	56

## 3.1 Launch UNICORN start 1.3

## Start and log on

Follow the instructions below to start and log on to UNICORN start 1.3.

#### Step Action

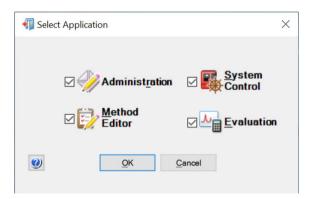
• Double-click the UNICORN start 1.3 icon on the desktop,



or

Select Start →All Programs →Cytiva →UNICORN start 1.3
 →UNICORN start 1.3.

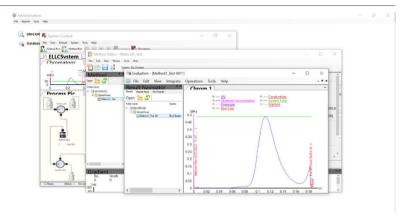
Result: The **Select Application** dialog opens.



2 In the **Select Application** dialog, select the module(s) to be opened.

Result:

The selected UNICORN start 1.3 modules open.



#### 3 Select System Control.

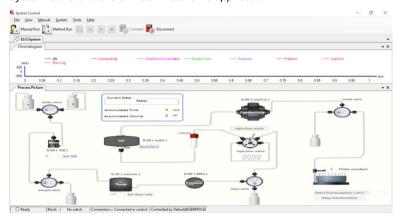
4 Click Connect to the ÄKTA start instrument.

#### Result:

UNICORN start 1.3 connects to the ÄKTA start.

#### Note:

Since the **UNICORN Instrument Server** takes time to sync with the instrument, as soon as the connection is established between the Instrument and the PC the system will disconnect. Make sure to unplug and then plug-in the USB cable (check the status of the Instrument display, only **Settings and Service** should be **Enabled**). Now click the **Connect to System** tab in the System Control of the UNICORN start 1.3 application.





#### **IMPORTANT**

Make sure that the computer which is used for running UNICORN start 1.3 is connected to the ÄKTA start instrument using the USB cable. Check the connection status on the standalone display.

#### Start and evaluate a Manual run

Follow the instructions below to start a Manual run using UNICORN start 1.3.

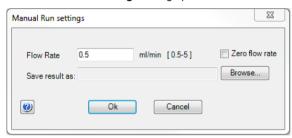
#### Step Action

1



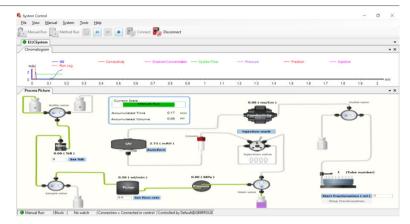
in the toolbar to start a manual run.

The Manual Run settings dialog opens.



- 2 Enter the desired **Flow Rate** as ml/min.
- 3 Click the **OK** button to start the manual run.

The run starts and **System Control** shows the current flow path as well as the real time chromatogram.

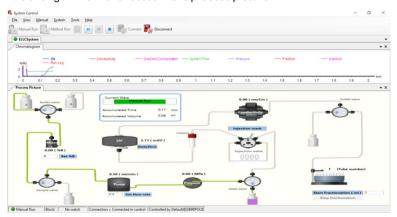


4 Select the upper radio button to change the position of the wash valve.



#### Result:

The change in flow is reflected in the process picture.



End the run by clicking the buttor

Result:

The run stops.

6 Open the **Evaluation** module by selecting it on the Windows task bar or by selecting **Tools** → **Evaluation**.

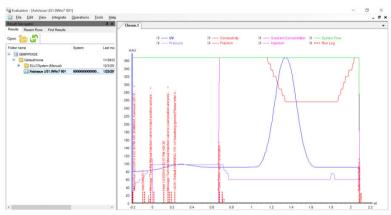
Result:

The **Evaluation** module opens.

7 Open the **Result Navigator** by clicking the icon or the **Result Navigator** icon.

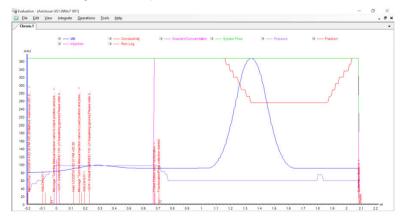
Result:

The Result Navigator opens.



8 Double-click the most recent run to open the chromatogram.

The chromatogram result opens.



Step	Action
9	Click the icon to save a PDF result and select a location to save the file.  The PDF result is generated and saved in the location selected above.

#### **Exit UNICORN start 1.3**

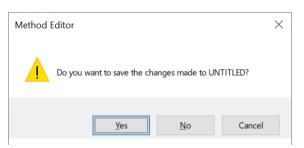
To exit UNICORN start 1.3, select  $File \rightarrow Exit \, UNICORN$ . This can be performed from any of the UNICORN start 1.3 modules.



Result: All open UNICORN start 1.3 modules close.

Note:

If an updated method or result is open and not saved during exit or log off from UNICORN start 1.3, a warning is displayed. Click **Yes** to save, **No** to exit without saving, or **Cancel** to stay logged on.



# 3.2 Help functions

#### The help viewer application

As part of the UNICORN start 1.3 installation, a special viewer for the online help is installed. This application, the *MadCap HelpViewer*, can be accessed from the UNICORN start 1.3 user interface either by clicking help buttons in dialogs, by pressing the **F1** key or by selecting *Help* from the menu bar.

By default, this application places a shortcut icon on the desktop, which can be used to open the online help without logging on to UNICORN start 1.3. However, if the desktop icon is not present, this application can also be accessed from **Start**  $\rightarrow$ **All Programs**  $\rightarrow$ **MadCap Software**  $\rightarrow$ **MadCap HelpViewer V6.** 

To start the UNICORN start 1.3 online help:

## Step Action 1 Double-click the shortcut icon. MadCap HelpView... 2 Choose File → Open. 3 Browse to the folder UNICORN\Documentation\Help. 4 Select the file Manual.mchelp. 5 Click the **Open** button. Result: The online help portal page opens.

#### Note:

Sometimes Windows may need to register **MadCap HelpViewer** as the application for files of the type .mchelp before it recognizes calls from help buttons in UNICORN start 1.3. If that happens, open the viewer manually as described above.

# 4 System Control

# In this chapter

Section	on	See page
4.1	Overview	58
4.2	Connect to system	61
4.3	Start a run	62
4.4	Monitor a run	67

## 4.1 Overview

#### Introduction

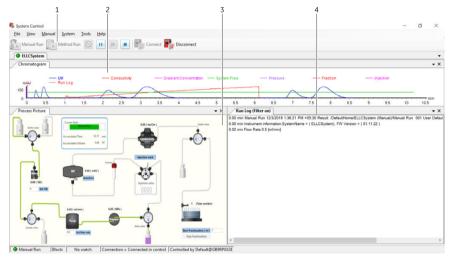
The  ${\it System Control}$  module is used to start, monitor and control a chromatography run.

Runs can be performed in two different ways, as listed below:

- Using a predefined method template.
- Performing the chromatography run by providing manual instructions.

## System Control user interface

The illustration below shows the different panes and areas of the **System Control** module.



**Note:** The status bar at the bottom of the window shows the current connection and control status for the displayed system.

Pane	Function
1	Toolbar: Includes buttons and icons for system control.
2	<b>Chromatogram</b> : Illustrates data as curves such as UV absorbance (mAU), Conductivity (mS/cm), Flow rate (mI/min), Pressure (mPa), Gradient (%B), Run log and Fraction.
3	<b>Process Picture</b> : Illustrates the current flow path.
4	Run Log: Displays the current state of the system.

# **Toolbar icons in the System Control** module

The table below describes the toolbar icons in the **System Control** module.

Icon	Function
Method Run	Starts a method run.  Note:
Start a Method Run	It is mandatory to select a method from the <b>Method</b> Navigator dialog to start the run.
Manual Run	Starts a manual run.
Start a Manual Run	
Hold	Temporarily holds the run, with currently set flow rate, valve positions, and B concentration.
Pause	Temporarily pauses the run by stopping the pump.
Continue	Continues the run that has been paused
End	Terminates the ongoing run.
Connect	Connects the software to the instrument.
Disconnect	Disconnects the software from the instrument.

## Select panes to display

The table below describes how to display panes:

# In the System Control menu bar, select View. Select the panes to display. View Manual System Tools Help Toolbar Status Bar Chromatogram Process Picture

Selected panes are displayed on the screen.

Run Log

Refresh

Restore to Default

#### Instrument states

Information about the status of instrument connections and chromatography runs is displayed as *Current State* in the chromatogram.

F5

# 4.2 Connect to system

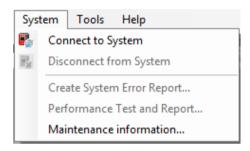
# Connect the ÄKTA start system to UNICORN start 1.3

Note:

Since the **UNICORN Instrument Server** takes time to sync with the instrument, as soon as the connection is established between the Instrument and the PC the system will disconnect. Make sure to unplug and plug in the USB cable from the PC (check the status of the Instrument display, only **Settings and Service** should be **Enabled**). Now click the **Connect to System** tab in the System Control of the UNICORN start 1.3 application.

Click on the icon to connect UNICORN start 1.3 to the ÄKTA start instrument.

Alternatively, navigate to  $\textbf{System} \rightarrow \textbf{Connect to System}$  in the System Control module.



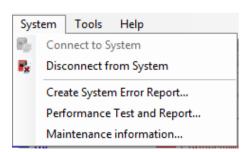
#### **Disconnect from system**

Click on the



icon to disconnect UNICORN start 1.3 from the ÄKTA start instrument.

Alternatively, navigate to  $\textbf{System} \rightarrow \textbf{Disconnect from System}$  in the System Control module.



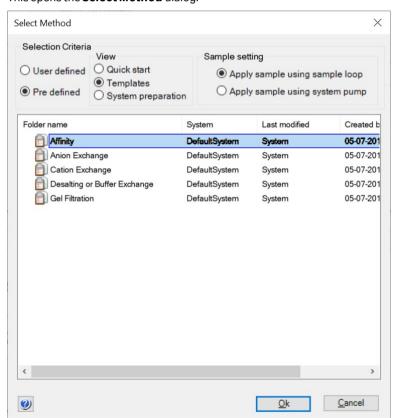
#### 4.3 Start a run

#### Start a method run

Follow these steps to start a method run:



This opens the **Select Method** dialog.

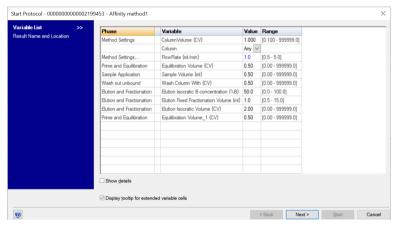


- Select the desired Selection criteria: User defined or Pre defined.
- For **User defined**, find and select a suitable method in the folder pane.

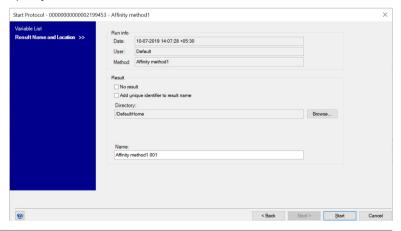
  For **Pre defined**, select **View** and find a suitable method in the folder pane, and then select **Sample setting**:

- a. View:
  - Quick start
  - Templates
  - · System preparation
- b. Sample setting:
  - · Apply sample using sample loop
  - Apply sample using system pump
- 4 Click the **OK** button to start the selected method run.

This opens the **Start protocol** dialog.



- 5 Review the *Variable List* and change method parameters if required, and then click *Next* to proceed to the next page.
- 6 Specify a result name and then click **Start** to start the run.



#### Start a manual run

Follow these steps to start a manual run:

#### Action Step 1 Manual Run icon in the toolbar. This opens the Manual Run settings dialog. X Manual Run settings Zero flow rate Flow Rate 0.5 ml/min [0.5-5] Browse... Save result as: (0) 0k Cancel 2 Enter the desired flow rate. 3 Click the **Browse** button and select the location where the result should be saved. Click the **OK** button to start the manual run.

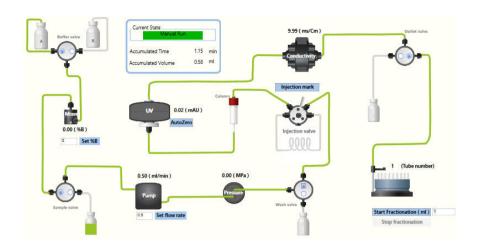
#### **Manual Instructions**

4

To open the *Manual Instructions* dialog, select *Manual* → *Execute Manual* 

The following manual instructions can be performed from the process picture (see the illustration below):

- All valve changes can be executed with a single click.
- % B concentrations can be set.
- Flow rate can be changed.
- Fractionation can be started or stopped.
- Autozero can be performed; an injection mark can be set.



# Perform manual instructions during a method run

The instruction below is an example of how to manually interact with an ongoing method run. The example shows how to increase the system flow.

#### Step Action

- 1 Open the *Manual Instruction* dialog according to the above instruction.
- 2 In the *Instructions* list, select the instruction group *Pumps* and instruction *Flow rate*.
- 3 Enter a new value for **Flow rate**.



- 4 To execute multiple instructions at the same breakpoint, select and edit an instruction and then click *Insert*. Repeat the same steps to include multiple instructions.
- 5 To refresh the parameter fields during a method run, check the **Auto update of parameters during run** checkbox.
- 6 To execute the instructions, click the **Execute** button.

#### 4 System Control

4.3 Start a run

#### Note:

The first instruction only takes flow rate and gradient parameters and starts the run with all valves in the default position regardless of queued instructions. Multiple instructions at the same breakpoint can only be executed after the manual run has started.

During a sample application with the sample valve, buffer valve changes or gradient instructions are not recommended, as the buffer valve is in sample position.

#### 4.4 Monitor a run

#### Introduction

This section describes the data shown in **System Control** during a run, and the procedure to customize the view of the different panes. It also describes how to enable alarm and error notifications.

You can follow the ongoing method run in the **System Control** module.

- Selected curves are shown in the *Chromatogram* pane.
- The current flow path is shown in the **Process Picture** pane.

To find an overview of the **System Control** user interface, see Section 4.1 Overview, on page 58.

#### **Open the Customize dialog**

To customize displayed information and data in the different panes:

• Select **Customize** from the **Tools** menu in the **System Control** toolbar.

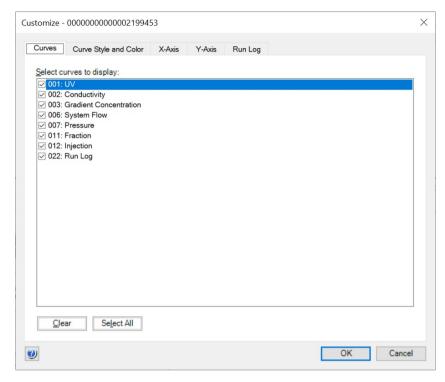


or

Right-click in the different panes (except *Process Picture*) and select *Customize*.
 Result: The *Customize* dialog is opened.

#### 4 System Control

#### 4.4 Monitor a run



**Note:** Further information about the settings in the **Customize** dialog can be found in the Online Help.

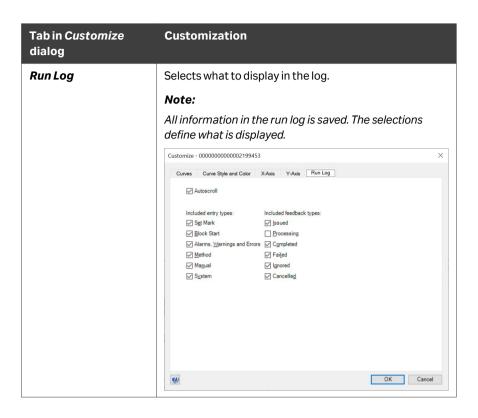
# **Customize Chromatogram**

The **Chromatogram** pane displays registered curves during the run.

Tab in Customize dialog	Customization
Curves	Choose curves to be displayed
Curve style and color	Customize the appearance of the displayed curves
X-axis	Choose base unit (time or volume) and set axis scale
Y-axis	Set scale for the different curves and select which axes to display

### **Customize Run Log**

The *Run Log* pane displays registered actions during the run.



# Select and zoom a curve in the chromatogram

The table below shows how to select a curve and to zoom in the chromatogram.

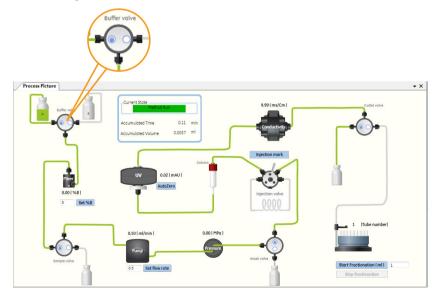
Command	Instruction
Select a curve	<ul> <li>Position the mouse marker over a curve name:         The curve line becomes bold to facilitate the identification of the desired curve.     </li> <li>Click the curve:         The corresponding legend text is shown in bold and the Y-axis scale changes to the unit applicable for the curve.     </li> </ul>

Command	Instruction
Zoom	Click and hold the left mouse button with the pointer positioned in the top left corner of the area you want to zoom in on.
	Drag the pointer downwards and to the right. A dotted rectangle marks the selected area. Select the area you want to zoom in on, and release the left mouse button.
	(The text <b>Zoomed mode</b> is shown in the chromatogram)
	To reset zoom, right-click and select <b>Reset zoom</b> .
	To reset the last zoom action, right-click and select <i>Undo zoom</i> .
Right-click menu options	Right-click and select options and actions. See <i>Online Help</i> for information.

# Illustration of the Process Picture pane

The **Process Picture** pane shows the current flow path during the run. Color indication is applied and real-time data from monitors are shown.

The following illustration shows an example of a process picture. In this example, the orange marking indicates where you can shift between options by clicking the radio button for e.g., Buffer A or B.



The table describes how different colors indicate the current state of the flow paths; open or closed.

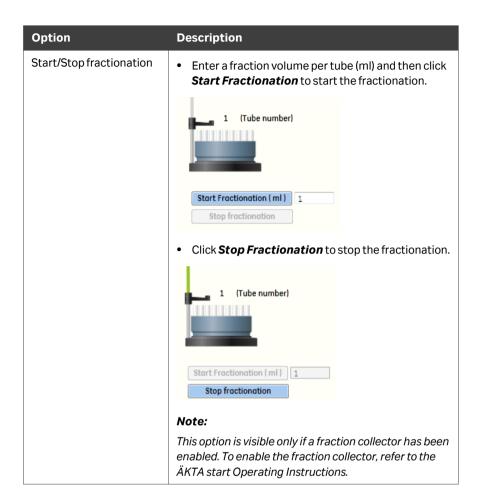
Color	Indication
Green	Open flow path
Gray	Closed flow path

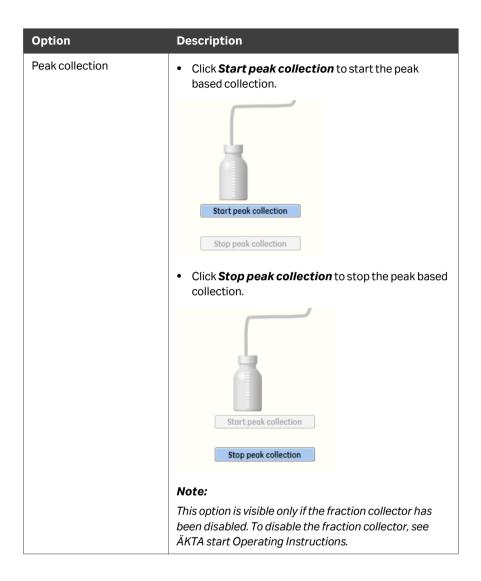
# **Options in the Process Picture pane**

The following table describes how to change valve positions, set values, start/stop fractionation and start/stop peak collection in the **Process Picture** pane.

Option	Description	
Change valve positions	Click the radio buttons to change between the valve positions.	
Set % B concentration	Enter a value for setting the B concentration and then click <b>Set</b> %B.  Output  Set %B	
Set flow rate	Enter a value for flow rate and then click <b>Set flow rate</b> to set the flow rate.  0.50 (ml/min)  Pump  0.5 Set flow rate	

Option	Description
Introduce Injection mark	Click the <i>Injection mark</i> button to introduce an injection mark in the chromatogram.  Injection mark Injection valve
Preform UV baselining	Click the <b>AutoZero</b> button to perform UV baselining.





## 5 Create a method

## **About this chapter**

This chapter contains a general introduction to creating and editing methods using UNICORN start 1.3. It also describes overall method options, how to print methods, and how to import/export methods. Descriptions of the predefined methods and phases supplied with the software are included as well. The chapter contains the following sections:

## In this chapter

Section	n	See page
5.1	Overview	76
5.2	Create and edit methods	83
5.3	Predefined methods and phases	88
5.4	Fraction collection	97
5.5	Import and export methods	107
5.6	Column handling	112

## 5.1 Overview

#### About this section

This section gives an introduction to the **Method Editor** module of UNICORN start 1.3. It also gives a brief description of the **Method Editor** interface and describes the concept of methods in UNICORN start 1.3.

For information on how to create, open and edit methods and import/export methods, see Section 5.2 Create and edit methods, on page 83.

#### In this section

Section	Section Se	
5.1.1	Method Editor	77
5.1.2	Methods in UNICORN start 1.3	80

#### 5.1.1 Method Editor

#### Introduction

The **Method Editor** module is used to:

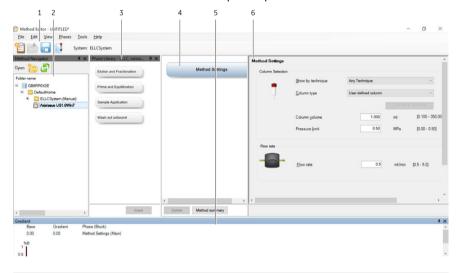
- · Create and edit methods.
- · Copy, save and delete methods.

**Method Editor** also assists the user in optimizing runs and allows handling of column types and user defined columns. For more information, see the following sections.

#### **Method Editor interface**

The **Method Editor** interface consists of different panes that allows the user to get an overview of the method, a deep dive into each phase of a method and what methods and phases the database contains.

The *Method Editor* interface with all the optional panes is illustrated next.



Pane	Description	
1	Toolbar (optional pane): Shows the toolbar icons.	
2	<b>Method Navigator</b> (optional pane): Shows all the user folders and methods that are available in the database.	
3	Phase Library (optional pane): Contains all available phases.	
4	Method outline: Shows the phases included in the open method.	
5	<b>Gradient</b> (optional pane): Shows the programmed gradient and break points for included phases.	

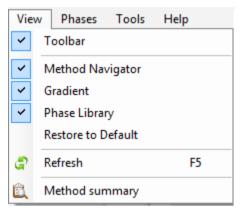
Pane	Description
6	<b>Phase properties</b> : Shows the settings for the highlighted phase in the method outline.

### **Display optional panes**

The optional panes in **Method Editor** are displayed by selecting them in the **View** menu.

To restore the appearance of **Method Editor** and display the default panes, select **Restore to Default** in the **View** menu. Then, the **Toolbar**, **Gradient** and **Phase Library** are displayed. The appearance of the optional panes can also be controlled using the auto hide function (see below for more information).

The illustration below shows the **View** menu with the default panes selected.



#### Method Navigator auto hide

The **Method Navigator** may be displayed in the left portion of the **Method Editor** module, or the **Auto Hide** function can be selected by clicking the pin symbol in the top right-hand corner of the pane.

Pin direction	Function
Auto Hide	Auto Hide is off. Click the pin symbol to turn the function on.
Auto Hide	Auto Hide is on. Click the pin symbol to turn the function off.

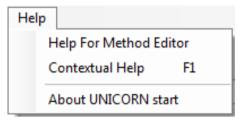
If **Auto Hide** is selected, the **Method Navigator** opens automatically when the mouse pointer is placed over its tab. It remains open as long as the mouse pointer remains over the pane. The pane closes automatically when the pointer is moved outside the pane.

# Getting help on the toolbar and panes in Method Editor

The table below describes how to find detailed information about the toolbar and the different panes in *Method Editor* by opening the online help.

#### Step Action

1 To display detailed information about the toolbar and different panes in the **Method Editor** interface, select **Help → Help For Method Editor**.



#### Result:

The online help opens and displays the *Method Editor* help start page.

To display help for a specific pane, click in the pane and press the **F1** key. Result:

The online help page describing that pane is opened.

#### 5.1.2 Methods in UNICORN start 1.3

#### **About methods**

The program instructions for a chromatography run are defined in a method. These instructions are specific for each instrument configuration and follow certain syntactical and hierarchical rules.

In a predefined method, each phase reflects a step in the chromatography run, for example, equilibration or sample application. Several settings are available for each type of phase. By building methods in this way, methods are easily created and edited.

See Section 5.2 Create and edit methods, on page 83 for information about creating and editing methods in **Method Editor**.

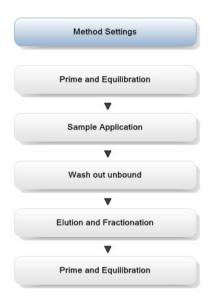
The illustration below shows the phases in a predefined method in the method outline pane (left) and the corresponding settings for the highlighted phase in the phase properties pane (right).



#### **Method structure**

A method always starts with the **Method Settings** phase. This phase contains the general settings that affect the rest of the method. If **Column type** is changed for a predefined method, UNICORN start 1.3 automatically calculates the correct settings for column volume, flow rate, and pressure limits that will be used throughout the method. Subsequent phases reflect steps included in the chromatography run.

A predefined method with the different phases in the method outline pane is illustrated next.



## Working with methods

It is recommended to create and edit methods using phase properties. Phases can easily be dragged-and-dropped into the method outline from the **Phase Library** and the phases are easily rearranged. Settings for each phase are set in the phase properties pane.

The phase properties settings for the **Method Settings** phase in a predefined method are illustrated next.



#### **Method types**

UNICORN start 1.3 provides several predefined methods for different chromatography techniques, and for preparation and maintenance of the instrument (e.g., preparation and cleaning of the system and columns). Customized methods may be created, by dragging and dropping the phases from the phase library followed by setting the parameters. The phase *Method Settings* is mandatory to all methods.

See Section 5.2 Create and edit methods, on page 83 for information about how to create new methods.

The table below gives a general description of the different method types.

Method	Description
Predefined Method	Predefined methods include several relevant phases appropriate for the purification or maintenance to be performed. These predefined methods can be used as they are, or with adjusted settings as needed.
	See Section 5.3 Predefined methods and phases, on page 88 for descriptions of the predefined methods provided with the software.
Customized Method	Customized methods include the mandatory phase <i>Method Settings</i> . Other phases are then added by the user from the Phase Library pane and settings adjusted as needed.

## **Predefined phases**

UNICORN start 1.3 provides several predefined phases.

Predefined phases (for example, *Prime and Equilibration*) can be used when building or editing methods in *Method Editor*. A predefined phase contains all the necessary instructions needed for a particular phase. All predefined phases can easily be viewed in the *Phase Library* pane.

See  $Section 5.3.1 \ Definitions$ , on page 89 for descriptions of the predefined phases supplied with the software.

## 5.2 Create and edit methods

## **About this section**

This section describes how to create and edit methods in UNICORN start 1.3 using the phase properties pane.

## In this section

Section		See page
5.2.1	Working with methods	84
5.2.2	Open a method	86

## 5.2.1 Working with methods

#### Introduction

In UNICORN start 1.3, the predefined methods for ÄKTA start are built up of phases, where each phase corresponds to a step in a chromatography run, with several properties associated with that phase. See *About methods, on page 80* for more information about method structure, definitions, and concepts of methods in UNICORN start 1.3.

Methods can be created and edited by using phases and phase properties.

# Main steps when defining a new method using predefined phases

The main steps when defining a method are:

Step	Description	
1	Create/open a method	
	<ul> <li>Create a predefined method (including a set of phases that may be edited)</li> </ul>	
	or	
	Open an existing method that can be edited and saved with a new name or overwritten.	
2	Build/edit the method outline and/or edit the phase properties for the appropriate phases	
	<ul> <li>Predefined Methods: use as they are, or edit the method outline and/or phase properties.</li> </ul>	
	Opened methods: edit the method outline and/or phase properties.	
3	Save the method.	

# Main steps when defining a customized method

The main steps when defining an empty method are:

Step	Description	
1	Create/open a method	
	• Create a new empty method containing the <b>Method Settings</b> phase.	

Step	Description
2	Build/edit the method outline and edit the phase properties for the phases
	<ul> <li>Add predefined phases to the method (i.e., build the method outline) and edit the phases as appropriate.</li> </ul>
	Rename the phases.
3	Save the method.

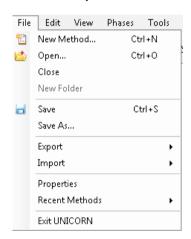
## 5.2.2 Open a method

Follow the instructions to open an existing method in the database:

#### Step Action

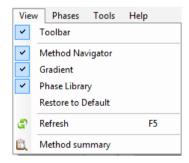
#### 1 In **Method Editor**:

- Click the icon in the toolbar
   or
- Select File → Open



or

Select View → Method Navigator

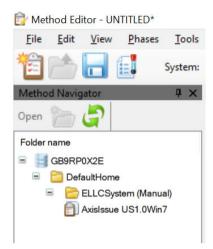


Result:

The **Method Navigator** is displayed.

#### Step Action

2 Select the method to be opened in the **Folder name** column.



- 3 To open the method,
  - Click the Open button located in the toolbar of the Method Navigator pane



or

- Double-click the selected method or
- Right-click on the method name and select **Open** from the context menu

#### Result:

The method is opened and displayed in the method outline pane with the included phases. The phases of the method are still editable using phase properties.

## 5.3 Predefined methods and phases

## In this section

Section	1	See page
5.3.1	Definitions	89
5.3.2	Create a predefined method	92
5.3.3	Edit phase properties	94

#### 5.3.1 Definitions

#### Introduction

A predefined method contains a set of phases, each phase reflecting a specific stage of a chromatography or maintenance run. Additional phases can be selected from the phase library and added to an existing method and phases that are not required can be removed.

The predefined purification methods have default values with suitable running conditions for the chosen column type such as flow and pressure limits. Other settings (e.g., sample application technique, sample volume, elution profile and fractionation) are set in the phase properties pane of the appropriate phases.

This section describes the predefined methods and phases.

## **Predefined purification methods**

**Method Editor** has five predefined methods for different separation techniques. The methods include several relevant phases.

The table below describes the available predefined purification methods and phases that are included.

Predefined purification method	Principle	Included phases
Affinity Chromato graphy (AC)	Separates molecules based on the reversible interaction between the target protein and the specific ligand attached to a chromatography matrix. The steps include equilibration, sample application, wash unbound step to remove unbound non-specific proteins, elution with fractionation followed by re-equilibrating the column.	Method Settings  Prime and Equilibration  Sample Application  Wash out unbound  Elution and Fractionation  Prime and Equilibration

#### 5.3.1 Definitions

Predefined purificatio n method	Principle	Included phases
Anion Exchange Chromato graphy (AIEX)	Based on the reversible interaction between negatively charged protein and positively charged chromatography media. The steps include equilibration with binding buffer, sample application, elution with change in salt concentration (e.g. NaCl) or change in pH of elution buffer and re-equilibration with binding buffer.	Method Settings  Prime and Equilibration  Sample Application  Wash out unbound  Elution and Fractionation  Prime and Equilibration
Cation Exchange Chromato graphy (CIEX)	Based on the reversible interaction between positively charged protein and negatively charged chromatography media. The steps include equilibration with binding buffer, sample application, elution with change in salt concentration (e.g. NaCl) or pH of elution buffer and re-equilibration with binding buffer.	Method Settings  Prime and Equilibration  Sample Application  Wash out unbound  Elution and Fractionation  Prime and Equilibration
Desalting	A gel filtration technique that allows rapid group separation of high molecular weight substances from low molecular weight substances. Small molecules like salt, free labels and other impurities are efficiently separated from the high molecular weight substances of interest. The method involves equilibration followed by sample application, elution and fractionation using single buffer.	Method Settings  Prime and Equilibration  Sample Application  Elution and Fractionation

Predefined purificatio n method	Principle	Included phases
Gel filtration	Also known as size-exclusion chromatography, is a	Method Settings
separates difference The meth sample a	chromatography technique that separates molecules based on differences in the molecular size. The method involves equilibration,	Prime and Equilibration  ▼  Sample Application
	sample application and elution and fractionation.	▼ Elution and Fractionation

## **Predefined phases**

The table below describes the predefined phases.

Phase Name	Description
Method Settings	The first, and mandatory, phase in any method. Defines common parameters used in the subsequent phases.
	The <b>Method Settings</b> phase defines:
	Column selection
	Flow rate
	Pressure limit
	Note:
	Default values for pressure limits and flow rate are given for the Cytiva selected column type.
	Note:
	Some of these options may not be required by certain methods.
Prime and Equilibration	Primes the flow path by filling it with buffers of interest, and equilibrates the column before a purification, or reequilibrates the column after a purification.
Sample Application	Applies sample to the column. Defines the sample application mode technique, sample volume, and handling of the flow through.
Washout unbound	Washes out unbound sample after sample application.
Elution and Fractionation	Elutes the sample from the column. Defines parameters for the elution and fractionation settings.

## 5.3.2 Create a predefined method

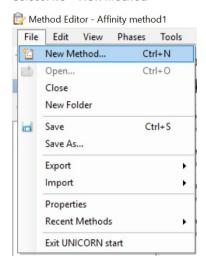
Follow the instructions to create a new method using phases:

#### Step Action

#### 1 In **Method Editor**:

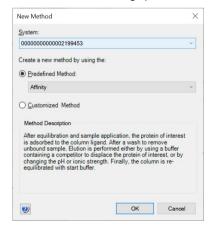


Select File → New Method



#### Result:

The New Method dialog opens.



#### Step Action

- 2 In the **New Method** dialog:
  - a. Select a Predefined Method.
  - b. Click the OK button.

#### Result:

The method outline pane shows the included phases for the chosen method and the phase properties pane shows the default settings for the currently highlighted phase.



## 5.3.3 Edit phase properties

#### Introduction

When editing phase properties, the changes affect either:

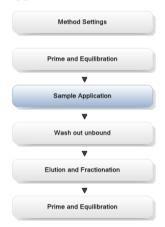
- The whole method, when editing the *Method Settings* phase.
- Only the phase that is being edited, when editing phases other than the *Method* Settings phase.

# Getting help when editing phase properties

Follow the instructions to get help for the properties in a phase:

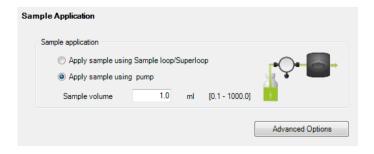
#### Step Action

Select a phase in the method to be edited, for example, Sample
 Application.



#### Result:

The properties for the selected phase are displayed in the phase properties pane.



#### Step Action

#### Note:

Click the **Advanced Options** button to display more phase properties settings.

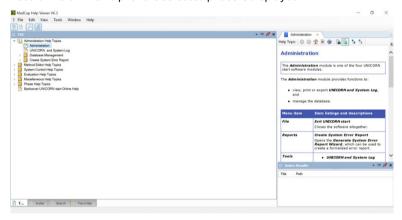
While a sample is injected with the manual injection valve, the system is on hold and the pump is running. Since the system is in manual mode during the sample application phase, it is recommended to be present in front of the system to prevent column dry out.

- 2 Click anywhere in the phase properties area to make it the active area in the software.
- Press the F1 keyboard key.

or

• Select Help → Contextual Help

Result: The Online help for the selected phase is displayed.



# View and edit phases using phase properties

Follow the instructions to edit a method phase in the **Phase properties** pane:

#### Step Action

- 1 Make sure the phase properties pane is selected.
- Select the *Method Settings* phase if the basic settings affecting the whole method need to be edited (e.g., *Column type*, and *Flow rate*). Continue with steps 3 and 4.

or

#### Step Action

- Select any other phase to edit the properties for that specific phase.
   Continue with step 5.
- To edit the properties for the *Method Settings* phase, click *Method Settings* in the method outline pane.

Result:

The phase properties of the *Method Settings* phase are displayed.

4 Edit the settings for the **Method Settings** phase in the phase properties pane as appropriate. If changing **Column type**, UNICORN start 1.3 automatically calculates the correct settings for volume, flow rate, and pressure limits.

#### Note:

Settings in this phase affect the whole method.

#### Note:

Allowed parameter ranges are shown in parentheses beside the text boxes.

The method is updated with the new settings.

5 Select a phase in the method to be edited, for example, Sample Application.

#### Result:

The properties for the selected phase are displayed in the phase properties pane.

- Edit the settings as appropriate.
  - Repeat steps 5 and 6 until the appropriate phases have been edited.

#### Result:

The method is updated with the new settings. The edited settings remain in place while subsequent phases are edited. If the method is closed and not saved, the settings will revert to the earlier values.

7 Save the method.

## 5.4 Fraction collection

## In this section

Section		See page
5.4.1	Introduction	98
5.4.2	Fractionation setup	99
5.4.3	Fractionation without Frac30	100
5.4.4	Fractionation with Frac30	102

#### 5.4.1 Introduction

Collecting the protein fractions during either sample application, wash unbound or elution step, is an important activity during a chromatography run. Some of the predefined phases and methods include options for fraction collection in the phases.

Note:

The Fraction Collector needs to be turned ON from the ÄKTA start Instrument Display for it to be available as an option within UNICORN start 1.3.

Whenever the user enables or disables the Fraction Collector from the ÄKTA start Instrument, communication between the PC and the instrument stops. To reestablish the connection, do the following:

- 1. Enable the Fraction Collector from the ÄKTA start instrument.
- 2. Unplug and then plug in the USB cable.
- 3. Go to the System Control module of UNICORN start 1.3 application and click the **Connect** tab.

#### **Fractionation**

Fractionation settings are available in the phase properties pane in the predefined phases **Sample Application**, **Washout Unbound** and **Elution and Fractionation**. These three phases are included in many of the predefined methods of UNICORN start 1.3. See <u>Section 5.3.3 Edit phase properties</u>, on page 94 for details on how to edit methods and phases.

For each phase, fractions can either be collected with or without fraction collector (using **Outlet Valve**). If there is no risk of sample loss, the eluate may be sent to the waste and not collected.

## 5.4.2 Fractionation setup

## **Options**

The following fractionation options are available based on the instrument configuration in use.

Instrument Configuration	Fractionation options
ÄKTA start, UNICORN start 1.3 and Frac30	<ul> <li>Fixed volume fractionation</li> <li>Peak fractionation</li> <li>Level based</li> <li>Slope based</li> </ul>
ÄKTA start and UNICORN start 1.3	Single Peak collection     Level based     Slope based
ÄKTA start and Frac30	Fixed volume fractionation
ÄKTA start	Collection of elution volume

## **Delay volume handling**

Option	Description
Fractionation with Frac30	Delay volume is collected in the first tube (T1) followed by the rest of the fractions in subsequent tubes.
Fractionation without Frac30	Delay volume is collected in the collection container along with fraction (total collected volume = Delay volume + fraction volume).

For setting the delay volume refer to section *Prepare the fraction collector* in Operating Instructions 29027057.

For detailed instructions on fractionation operations using ÄKTA start, see Operating Instructions 29027057.

## 5.4.3 Fractionation without Frac30

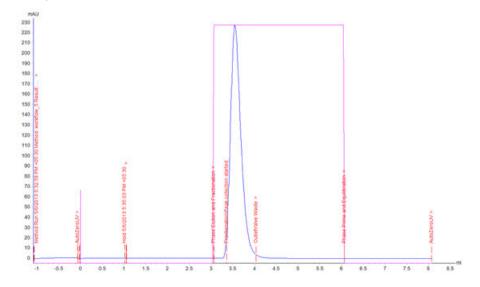
## Single peak collection



Single Peak Collection option is available when Frac30 has been disabled from the instrument. Single Peak Collection can be chosen and configured in the *Elution and Fractionation* phase as depicted above.

Setting	Description
Start level	UV absorbance (mAU) that triggers the start of peak collection.
End level	UV absorbance (mAU) that triggers the end of peak collection.

A representative chromatogram depicting fractionation by setting level [Abs(mAU)] values for collecting fractions from UNICORN start 1.3 using **Outlet Valve** (without Frac30) is shown below.



#### Note:

Make sure to standardize and set the optimized level values and fraction volume in order to collect the peak of interest in a single collection tube. For collecting multiple peaks, make sure to change the collection tube manually after collecting each peak.

- The UV monitor to Outlet Valve volume (0.27 ml) is constant for all instruments.
- Make sure to use recommended length and inner diameter of polyether ether ketone tubing from UV monitor to Outlet Valve to avoid incorrect calculation of delay volume.
- Make sure to update the length and inner diameter of the polyether ether ketone tubing in case the tubing is not of the recommended length and inner diameter.

## 5.4.4 Fractionation with Frac30

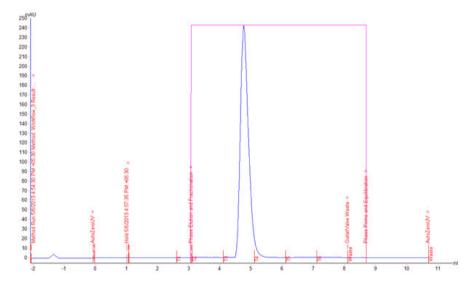
#### Fixed volume fractionation

Fixed volume fractionation can be chosen and configured in the **Sample Application**, **Washout unbound** and the **Elution and Fractionation** phases as depicted.

Fixed fractionation volume is the volume that will be collected in every fraction per tube.



A representative chromatogram depicting fractionation using fixed volume fraction collection from UNICORN start 1.3 with Frac30 is shown below.



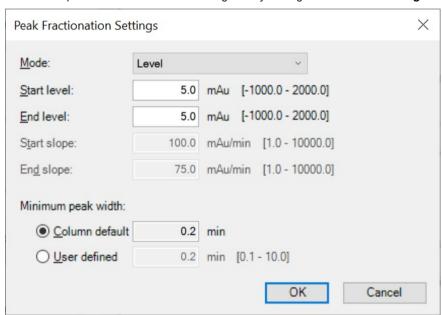
**Note:** Make sure to set fraction volumes that suit the column being used, and to fill adequate number of collection tubes.

#### Peak fractionation: Level based

Peak fractionation can be chosen and configured in the *Elution and Fractionation* phase as depicted.



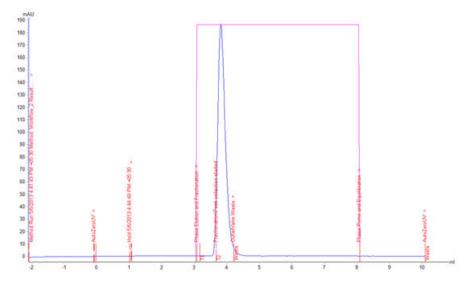
Level based peak fractionation can be configured by editing the **Peak Frac Settings**.



Setting	Description
Start level	UV absorbance (mAU) that triggers the start of peak fractionation.
End level	UV absorbance (mAU) that triggers the end of peak fractionation.
Column default	Selecting this option displays the default minimum peak width based on the column type selected in Method Settings.
User defined	To choose a custom value for the minimum peak width.

#### 5.4.4 Fractionation with Frac30

A representative chromatogram depicting fractionation by setting level [Absorbance (mAU)] values for collecting fractions from UNICORN start 1.3 with Frac30 is shown below.



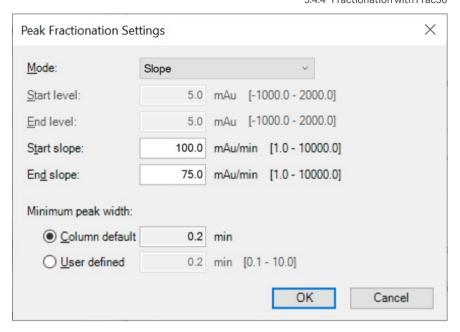
**Note:** Make sure to standardize and set the optimized level values and fraction volume in order to collect the peak of interest in a single collection tube.

## Peak fractionation: Slope based

Peak fractionation can be chosen and configured in the *Elution and Fractionation* phase as depicted.



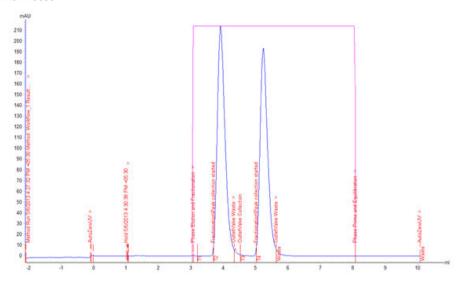
Slope based peak fractionation can be configured by editing **Peak Frac Settings**.



Setting	Description
Startslope	Slope value (mAU/min) that triggers the start of peak fractionation.
End slope	Slope value (mAU/min) that triggers the end of peak fractionation.
Column default	Selecting this option displays the default minimum peak width based on the column type selected in Method Settings.
User defined	To choose a custom value for the minimum peak width.

A representative chromatogram depicting fractionation by setting slope [Abs(mAU)/Time (min)] values for collecting fractions from UNICORN start 1.3 with Frac30 is shown below.

#### 5.4.4 Fractionation with Frac30



#### Note:

- Make sure to standardize and set the optimized slope values and fraction volume in order to collect the peak of interest in single collection tube.
- It is recommended to set lower slope values for collection smaller peaks and larger values for collecting higher peaks so that the peak of interest is collected in a single collection tube. The suitable slope values could be derived from UV curve with the UNICORN start 1.3 differentiate operation available in the **Evaluation** module.

## 5.5 Import and export methods

#### Introduction

UNICORN start 1.3 methods are stored internally in the UNICORN start 1.3 database. It is however possible to export entire methods or individual phases to a zip file on the local computer so that they can be imported again later into the same database installation, or imported into another database installation.

Alternatively, methods or phases can be exported as plain text files or Excel® files, which may be useful for documentation purposes.

## Export a method to another UNICORN start 1.3 database

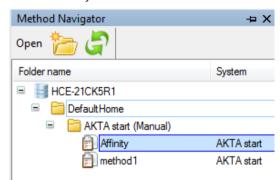
Follow these steps to export a method into a UNICORN start 1.3 database.

#### Step Action

1 In the **Method Navigator** pane, select the method to be exported.

#### Note:

Several methods in the same folder can be selected and exported at the same time. Also, several results can be selected simultaneously by using the **Shift** or **Ctrl** key.

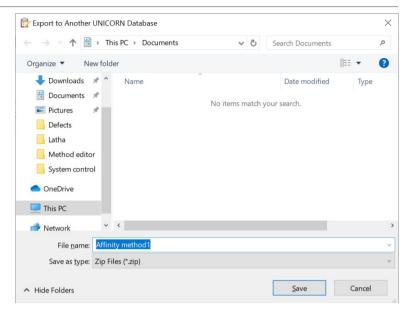


2 Choose File →Export →to UNICORN →Export Method to UNICORN.

Result:

The Export to Another UNICORN Database dialog opens.

#### Step Action



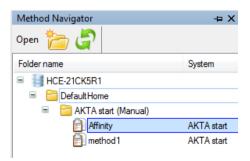
3 Choose a file name and location and click the **Save** button to save the zip file.

## Export a method to a plain text file or to Excel

Follow these steps to export a method as a plain text file, .asc or to Excel.

#### Step Action

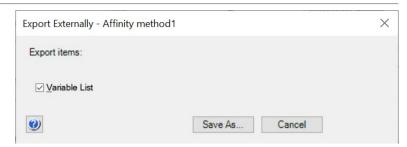
1 In the **Method Navigator** pane, select the method to be exported. It is only possible to export one method at a time to an external file.



2 Choose File → Export → Export Method Externally.

Result:

The Export Externally dialog opens.

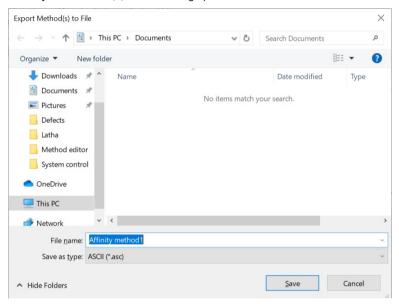


Only Variable list can be exported externally.

- 3 Click the **Save As** button to save the exported variable list.
- 4 To save the text file with the selected information included click the **Save As** button.

Result:

The Export Method(s) to File dialog opens.



- 5 Choose whether to save to an ASCII file or to an Excel file from the Save as type drop-down list.
- 6 Choose a file name and location and click the **Save** button to save the zip file.

### Import a method into UNICORN start 1.3

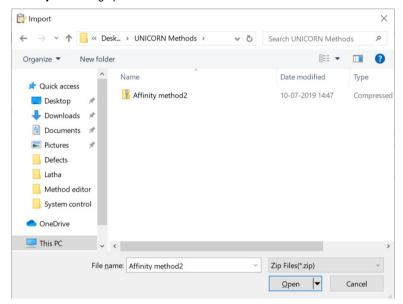
Methods that have previously been exported as zip files can be imported back into UNICORN start 1.3. Plain text files or Excel files can be imported if they contain all the information that UNICORN start 1.3 needs to recreate the method. Follow these steps to import a method.

### Step Action

1 Select File →Import →Import Method.

Result:

The Import dialog opens.

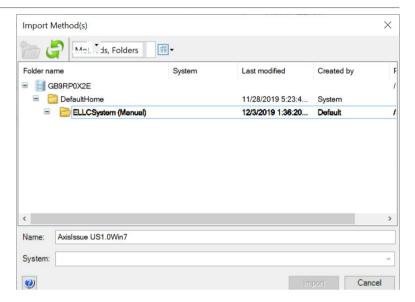


- 2 Browse to the required zip file in the *Import* dialog.
- 3 Select the file and click the *Open* button, or double click on the file name to open it.

### Note:

Several methods in the same folder can be selected and imported at the same time. Several results can be selected simultaneously by using the **Shift** or **Ctrl** key.

Result: The Import Method(s) dialog opens.



- 4 Browse to the required folder in the database and type in a new *Name* if necessary.
- 5 Click the *Import* button to import the method.

### Result:

The imported method is opened in *Method Editor*.

To import several methods, repeat the procedure in the  $\it Import\,Method$  dialog for each method.

### 5.6 Column handling

### Introduction

This section describes how to update predefined column types provided by Cytiva. For details on how to add, edit, delete, import, export, and print specific column types, please refer to *UNICORN start Online Help*.

Updated lists of predefined column types are provided by Cytiva. When a new list is imported, it will replace all the predefined column types in the database with the updated column types. Only predefined column types will be replaced; user defined column types will remain in the database.

**Tip:** For column list updates, please visit www.cytiva.com/unicornstart.

### Import column list into the database

Follow these steps to import a new column list into the database.

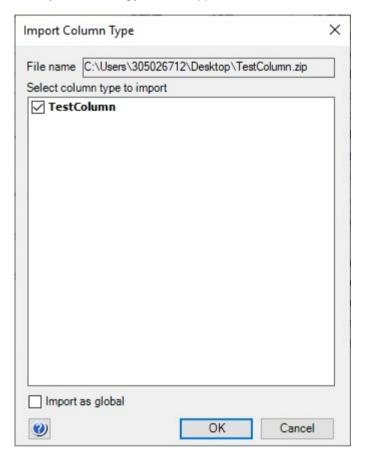
Step	Action
1	Click the column handling icon in the Toolbar.
2	In the <b>Column Type Parameters</b> tab, in the <b>Column Handling</b> dialog, click the <b>Import</b> button.
	<u>I</u> mport
	Result:

The *Import* dialog opens.

3 Locate the zip file with the column list to be imported and click **Open**.

Result:

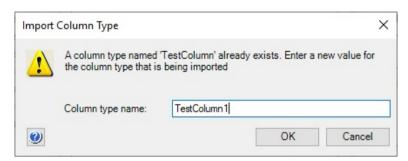
The Import Column Type window appears.



4 Click **OK**.

Result:

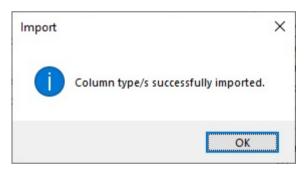
The Import Column Type dialog box appears.



5 Enter a column type name and then click **OK**.

Result:

The *Import* dialog box appears.



6 Click **OK**.

### 6 View and present the results

### **About this chapter**

This chapter describes how to view results, edit and optimize the representation of the results. It contains the following sections:

### In this chapter

Section	on	See page
6.1	Introduction	116
6.2	Evaluation module interface	117
6.3	View and present the results	122
6.4	Optimize the presentation of chromatograms	133

### 6.1 Introduction

A result is automatically generated at the end of a run and contains a complete record of the chromatography run, including system settings, monitored data, and run log. The complete result is saved in the UNICORN start 1.3 database.

UNICORN start 1.3 *Evaluation* module is used to view and analyze the result data and optimize the result presentation, for example, by:

- Integrating peaks.
- Measuring peak properties such as area and height.
- · Comparing curves and chromatograms from different runs.

When a satisfactory evaluation is obtained, a report may be compiled. The report can be anything from a simple print of a chromatogram, showing selected curves, to a comprehensive report containing chromatograms from several results.

### 6.2 Evaluation module interface

### In this section

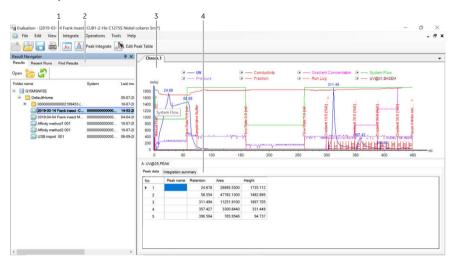
Section		See page
6.2.1	User interface	118
6.2.2	Result navigator	120
6.2.3	Chromatogram	121

### 6.2.1 User interface

The interface consists of a pane where the current result is displayed and, the **Result Navigator**. Both panes are described below.

### **Evaluation module interface**

Different parts of the *Evaluation* module are illustrated below:



Pane	Function
1	Toolbar
2	Result navigator
3	Chromatogram
4	Peak summary table

### Toolbar icons in the Evaluation module

The table below describes the toolbar icons in the **Evaluation** module.

Icon	Function
	Opens the <b>Result Navigator</b> . The <b>Result Navigator</b> is described below.

### lcon **Function** Opens the Add To Compare dialog. You may use the dialog to search for curves to compare in specified folders, results, or chromatograms. This dialog is described in Section 7.4 Compare different runs, on page 193. Note: This dialog can also be opened from the **File** menu. Saves the current result. This icon is used to save the current result that has been edited and not saved yet. Note: This function can also be selected in the **File** menu. Note: To save the result with a different name, select **File** →**Save As.** The **Save As** dialog opens, and the result can be named and saved in the selected folder. Opens the **Print Chromatograms** dialog. This dialog is used for printing the chromatograms on a selected printer. The chromatogram can also be previewed before printing. This dialog is described in Section 6.4.4 Print chromatograms and peak data, on page 145. Note: This dialog can also be opened from the **File** menu. Saves the report as PDF. This opens the Save As dialog where the result can be named and saved in the selected folder. Note: This dialog can also be opened from the **File** menu. Performs peak integration of the selected UV curve. This dialog is described in detail in Section 7.1 Peak integration, on page 149. Note: This dialog can also be opened from the **Integrate** menu. Opens the **Edit Peak Table** dialog. This wizard can be used to edit peak tables.

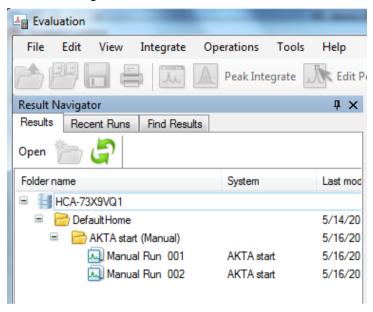
This dialog can also be opened from the **File** menu.

Note:

### 6.2.2 Result navigator

### Illustration of Result Navigator pane

The **Result Navigator** shows all the folders and individual results.



### Result Navigator auto hide

**Result Navigator** may be displayed in the left portion of the **Evaluation** module, or the auto hide function can be selected by clicking the pin symbol in the top right-hand corner of the pane.

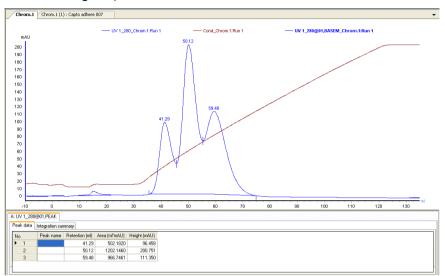
Pin direction	Function
Auto Hide	Auto Hide is off. Click the pin symbol to turn the function on.
Auto Hide	Auto Hide is on. Click the pin symbol to turn the function off.

If auto hide is on, the **Result Navigator** opens automatically when the mouse pointer is placed over its tab. It remains open as long as the mouse pointer remains over the pane. The pane closes automatically when the pointer is moved outside the pane.

### 6.2.3 Chromatogram

### The Chromatogram pane

The **Chromatogram** pane is illustrated below.



The current result contains two chromatograms. The chromatograms are displayed by clicking the corresponding tab. A **Peak Table** is displayed below the chromatogram. **Peak Table** are described in detail in Section 7.1.3 Display peak data, on page 154.

### **Docking Chromatogram panes**

If a result contains several chromatograms, each chromatogram is displayed in its own pane. The chromatograms can be stacked on top of each other and selected by clicking the corresponding tab. They can also be arranged in various positions using a docking function. The arrangement of chromatograms is described in *Section 7.4.1 Open and compare chromatograms, on page 194*.

### 6.3 View and present the results

### In this section

Section		See page
6.3.1	Open and view results	123
6.3.2	Locate results	129

### 6.3.1 Open and view results

### Introduction

A result file is opened in the **Evaluation** module where all of its content can be analyzed, and reports created. The **Evaluation** module user interface and toolbar icons are described in **Section 6.2.1 User interface**, on page 118.

The result files and their folder locations are shown in the **Result Navigator**. There are different ways to locate a specific result, Section 6.2.2 Result navigator, on page 120 describes this. In this section a preview function to identify a certain result and locate specific curves is described, it is called **Quick View**.

This section also describes how to highlight curves in a chromatogram, read curve values using a marker and save curve data as a **Snapshot**.

How to open specific chromatograms and curves is described in Section 7.4.1 Open and compare chromatograms, on page 194 and Section 7.4.2 Open and compare curves, on page 196.

### Open a result in the *Evaluation* module

There are four ways to open a result from the **Result Navigator**:

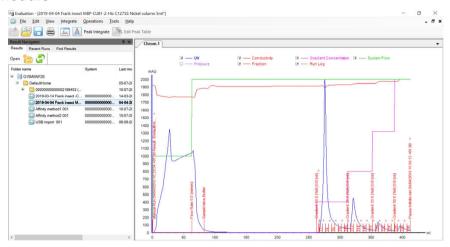
- Select a result and press the Enter key.
- · Double-click a result.
- Right-click a result and select **Open** from the shortcut menu.
- Select a result and click the **Open** toolbar icon in the **Result Navigator**.



Only one result at a time may be opened this way. If a new result is opened, the previous result closes automatically. However, several chromatograms from different results can be opened simultaneously. This is described in Section 7.4.1 Open and compare chromatograms, on page 194.

- 6 View and present the results
- 6.3 View and present the results
- 6.3.1 Open and view results

### Illustration of a result opened in the *Evaluation* module

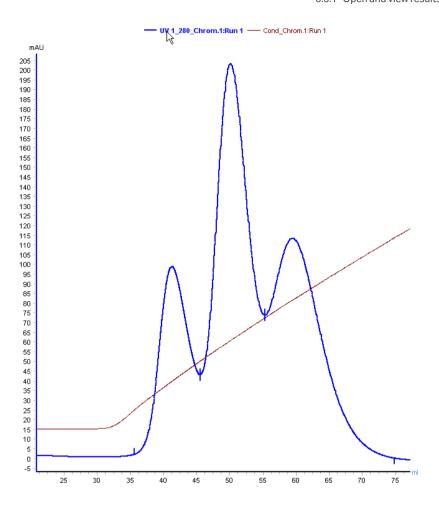


A result may contain several chromatograms and corresponding peak tables. Each chromatogram can be shown by clicking the corresponding tab. The chromatograms can also be displayed simultaneously by selecting **View**  $\rightarrow$ **Tile All Chromatograms**.

### Highlight or select a curve

An individual curve in the chromatogram can be selected or highlighted. The table below describes the differences:

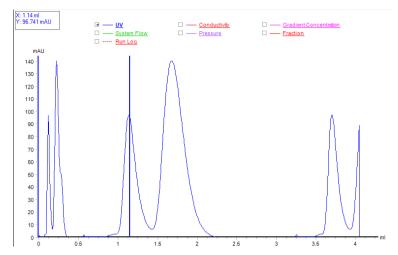
lfyou	Then
hold the mouse pointer over a curve segment	a pop-up box displays the curve name.
hold the mouse pointer over a curve name	the curve and the short line segment in front of the curve name become bold.
click a curve segment or a curve name	the Y-axis shows the values for this specific curve.



### Insert a vertical marker

The vertical marker is used to measure the values for a specific curve position. Right-click in the chromatogram and choose **Vertical marker**. Move the marker along the X-axis and read the X-axis and Y-axis values of the selected curve in the box in the top left corner of the chromatogram.

### 6.3.1 Open and view results



**Note:** The marker measures the curve that currently is selected if several curves are displayed in the chromatogram. The marker has the same color as the selected curve.

### Set marker reference

The vertical marker can be used for more measurements than just the readings from a specific curve position. Follow these steps to use the marker to determine **Delta Peak** and **Mean** Y-axis values.

### Step Action

- Position the marker to where the measurement should begin.
  - Right-click and select **Set vertical marker reference point**.

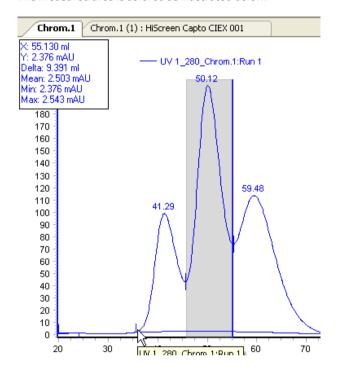
### Result:

The reference point is set to the position shown in the box in the top left corner of the chromatogram.

2 Drag the marker to the point where the measurement should end.

### Result:

The measured area is colored as illustrated below:



3 Read e.g. the **Volume**, **Delta** and **Mean** values from the box:

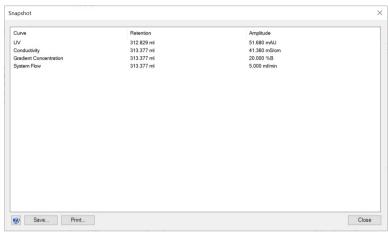
X: 55.130 ml Y: 2.376 mAU Delta: 9.391 ml Mean: 2.503 mAU Min: 2.376 mAU Max: 2.543 mAU

**Note:** If the X-axis has been set to volume base, X will hear equal to the volume for the selected area.

### **Snapshots**

It is possible to take a snapshot of all the curve values at the marker position. Follow these steps to take a snapshot:

- 1 **a.** Position the marker to where the snapshot should be taken.
  - **b.** Right-click and select **Snapshot** from the shortcut menu.



- 2 **a.** Click the **Save** button to save the **Snapshot**.
  - b. Alternatively, click the *Print* button to print the *Snapshot*. The *Print* dialog opens and the *Snapshot* can be printed.

### Note:

Snapshots taken during the method run are saved directly to the result and can be accessed by clicking the **Snapshot** sub-tab in the **Result Information** tab of the **Documentation** dialog.

3 Click the **Close** button to close the **Snapshot** dialog.

**Note:** The snapshot only records the values of the curves that are displayed in the current view.

### 6.3.2 Locate results

### Introduction

In the *Method Editor* and *Evaluation* modules of UNICORN start 1.3 there are navigator panes that can be used to locate and open methods and result files, and to arrange files in folders.

The **Evaluation** module contains a **Result Navigator** pane that is of the same type as the navigator pane in the **Method Editor** module, but it also contains a pane for **Recent Runs** and a specific search pane, **Find Results**. If desired, the **Result Navigator** can also display methods.

### Show the navigator pane

To open and display the navigator panes:

- Choose the *Result Navigator* menu command in the *View* menu.
   Or.
- · Click the



(open result navigator) toolbar icon.

Note:

As described in the general overview of the **Evaluation** module in Section 6.2 Evaluation module interface, on page 117, Auto Hide can be selected for the navigator panes.

### Navigator toolbar

The table below shows the navigator toolbar icons:

Icon	Function
	Opens the selected result.
Open	Note:
	Selected items can also be opened by selecting the <b>File</b> → <b>Open</b> menu command or by double-clicking the file in the navigator pane. The <b>Open</b> command from the right-click shortcut menu can also be used to open selected chromatograms, curves or peak tables.
<b>*</b>	Creates a new folder in the folder that is currently selected.  Note:  This function can also be accessed by selecting the File →New Folder
	menu command or selecting the <b>New Folder</b> command from the right- click shortcut menu.

### 6.3.2 Locate results

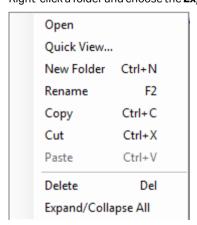
# Performs a Refresh in the navigator pane to update all items to the current status. Note: This function can also be accessed from the View menu. However, note that the Refresh menu command updates both the items in the navigator pane and items open in the module. The navigator toolbar icon updates only the navigator pane. Opens the View Details drop-down list where the following optional information columns may be selected for display in the navigator pane: System Created by Last modified

### **Expand and collapse folders**

When a folder is selected in the **Result Navigator** pane, the subfolders and results are displayed.

If the folder structure is expanded to lower levels, collapse all folders below a selected top folder, to simplify browsing.

• Right-click a folder and choose the **Expand/Collapse All** shortcut.



Result: The folder structure is collapsed to the selected folder level.

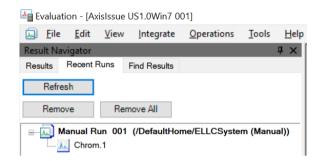
### The Recent Runs pane

The **Recent Runs** pane shows all the available recorded recent runs based on the selected preferences. Follow the instructions below to use **Recent Runs** to locate and open a result.

1 Click the Recent Runs tab in the Result Navigator.

Result:

The **Recent Runs** pane opens.



2 If needed, click the **Refresh** button.

Result:

The **Recent Runs** list is updated with all runs that were performed since the **Result Navigator** was opened the last time.

3 Locate and double-click the desired result.

Result:

The result opens in the **Evaluation** module.

Note:

Click the + signs to view or select individual chromatograms from the results. Individual results can be selected and removed from the list by clicking the **Remove** button. The **Remove all** button clears the whole list. The **Remove** and **Remove all** buttons only clear the list, the results are not deleted.

### Search for results using Find Results

The **Find Results** function in the **Result Navigator** is used to locate results in the available folders.

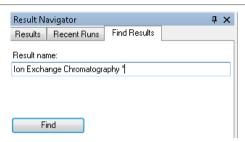
**Note:** There is also a common **Find** function which is available in the **Edit** menu of all modules except the **Administration** module.

The table below describes how to use the *Find Results* function to locate and open a result.

### Step Action 1 Click the Find Results tab. Result: The Find Results pane opens.

6.3.2 Locate results

### Step Action



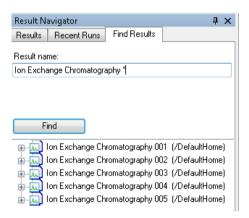
2 Type a result name or part of a result name in the **Result name** text box.

### Note:

The standard wildcard character \* can be used to represent a number of characters before or after the partial result name.

3 Result:

The located results are listed.



4 Double-click the desired result or chromatogram.

Result:

The file or chromatogram opens in the **Evaluation** module.

### Close the navigator pane

To close the navigator pane:

• Click the small cross in the top right-hand corner of the navigator.

Result: The navigator closes.

### 6.4 Optimize the presentation of chromatograms

### In this section

Section		See page
6.4.1	Customize the chromatogram layout	134
6.4.2	Edit curve presentation	137
6.4.3	Change the axes	141
6.4.4	Print chromatograms and peak data	145
6.4.5	Save report as PDF	147

### 6.4.1 Customize the chromatogram layout

### Introduction

The **Customize** dialog box is used to change the chromatogram presentation. The main features of the **Customize** dialog box regarding chromatograms are described in the subsequent sections. Features regarding peak tables are described in Section 7.1 Peak integration, on page 149.

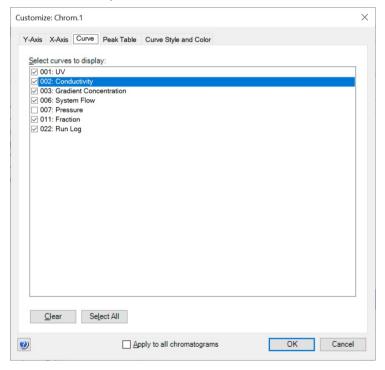
### Instruction

Follow the instructions below to make changes in the **Customize** dialog box:

Step	Action
1	Open a result.
2	Right-click the chromatogram and select <i>Customize</i>
	or
	• Choose <i>Tools</i> → <i>Customize</i> .

Result: The **Customize** dialog box is displayed.

**a.** Carry out the changes on the different tabs to get the desired layout for header, curves, and peak table. For more info, see below.



Step	Action	
	<b>b.</b> Select <i>Apply to all chromatograms</i> if you want to apply the changes made in the <i>Customize:</i> dialog to all open chromatograms.	
4	Click <b>OK</b> to apply the changes.	

### Tabs in the Customize dialog

The table below lists the tabs of the **Customize** dialog and outlines the editing functions of each tab.

Tab	Use this tab to
Y-Axis	Determine how the Y-axis is displayed. This is described in detail in Section 6.4.3 Change the axes, on page 141.
X-Axis	Determine how the X-axis is displayed. This is described in detail in Section 6.4.3 Change the axes, on page 141.
Curve	Select the curves to display in the chromatogram.
Peak Table	Select the <b>Peak Table</b> settings. This is described in detail in Section 7.1.3 Display peak data, on page 154.
Curve Style and Color	Determine how the curves are displayed. This is described in detail in Section 6.4.2 Edit curve presentation, on page 137.

**Note:** The selected settings in all tabs can be applied to all open chromatograms.

### Layout options in the right-click menu

You can also change the chromatogram layout by choosing some menu items from the right-click shortcut menu. The table below describes the layout options available on this menu:

Choose	when you want to
Grid	Add a grid to the background of the chromatogram.

6.4.1 Customize the chromatogram layout

Choose	when you want to
Filter Curves	Add a list of all available curves to the chromatogram, with checkboxes to select the curves to display:  — UV 1 280 Chrom.1:HiScreen Capto CIEX 001  — UV 2 0 Chrom.1:HiScreen Capto CIEX 001
	□ UV 3 0 Chrom.1:HiScreen Capto CIEX 001 □ Cond Chrom.1:HiScreen Capto CIEX 001 □ % Cond Chrom.1:HiScreen Capto CIEX 001 □ Cond B Chrom.1:HiScreen Capto CIEX 001 □ pH hrom.1:HiScreen Capto CIEX 001 □ System flow Chrom.1:HiScreen Capto CIEX 001 □ System pressure Chrom.1:HiScreen Capto CIEX 001 □ Cond temp Chrom.1:HiScreen Capto CIEX 001
	Note:
	Click the arrow buttons to show additional curves that do not fit in the available space.
Select Active Curve Only	Show only the active curve in the chromatogram. (this option is available only when <i>Filter Curves</i> is selected)
Select All Curves	Show all curves in the chromatogram.
	(this option is available only when <b>Filter Curves</b> is selected)
Legend	Show the list of all displayed curve names over the curve area of the chromatogram.
<ul> <li>Time or</li> <li>Volume or</li> <li>Column volume</li> </ul>	Select the unit of the X-axis.

### 6.4.2 Edit curve presentation

### Select curves to display

The *Curve* tab of the *Customize* dialog contains a list of all the curves included in the chromatogram. Select the curves to be displayed in the chromatogram and click the *OK* button.

### **Curve style and color settings**

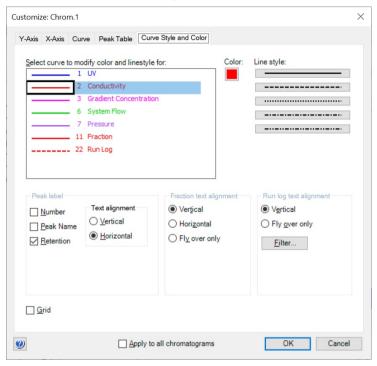
All curves within a chromatogram are represented by a default color and line style. Curves that are imported into the chromatogram or newly created curves are automatically assigned a color and line style. The settings are available in the **Curve Style and Color** tab. The tab also contains settings for peak labels, fraction marks and run log texts.

### Change the color and style of a curve

The table below describes how to change the color and style of a curve:

Step	Action	
1	Open the <b>Customize</b> dialog.	
2	Click the <b>Curve Style and Color</b> tab.	
3	a. Select the curve to change, from the list.	
	b. Click the <i>Color</i> button to open the <i>Color</i> dialog and choose a color from the palette.	

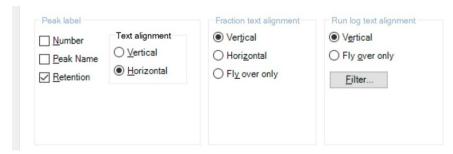
c. Click a button to choose a Line style.



- **d.** Repeat step 3 to modify other curves.
- 4 Click the **OK** button to apply the changes.

### Align Fraction and Run log texts

Both *Fraction* and *Run log* text can be set to be displayed either when the mouse pointer is positioned over a fraction mark (*Fly over only*) or at all times. The *Fraction* text can be aligned both vertically and horizontally. Default setting for both text types is *Vertical*. The *Vertical* selection for *Run log* text is the setting to show the text at all times.



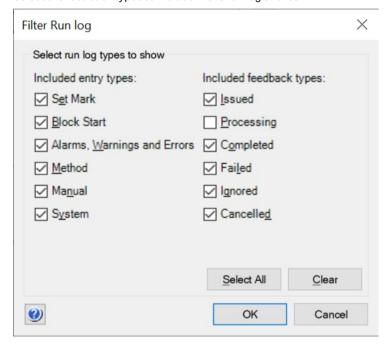
### **Filter Run log information**

4

Follow the instructions below to select the run log entry and feedback types to show in the chromatogram:

## Step Action Open the Customize dialog. Click the Curve Style and Color tab. Click the Filter button in the Run log text alignment field. Filter... Result: The Filter Run log dialog opens.

- a. Select the entry types to show in the chromatogram.
  - **b.** Select the feedback types to include in the run log entries.



c. Click the OK button.

Result:

The Filter Run log dialog closes.

5 Click the **OK** button in the **Customize** dialog to apply the changes.

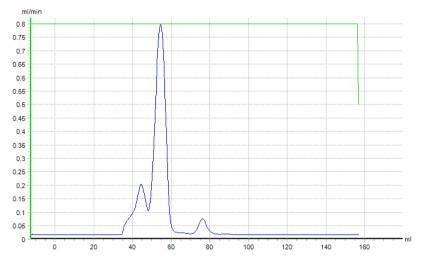
6 View and present the results

6.4 Optimize the presentation of chromatograms

6.4.2 Edit curve presentation

### Add a grid

It is possible to add a grid to the background of the chromatogram by selecting the *Grid* checkbox.



**Note:** This can also be accessed by right-clicking in the chromatogram and selecting **Grid** from the shortcut menu.

### 6.4.3 Change the axes

### Introduction

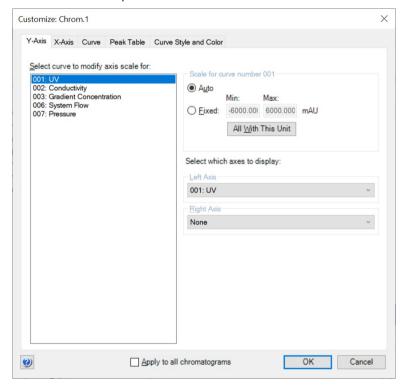
This section describes how to change the chromatogram axes. These changes are made in the **Y-Axis** and **X-Axis** tabs of the **Customize** dialog.

### Change and fix the Y-axis

Follow the instructions below to change and fix the scale of the curves in the chromatogram Y-axis:

### Step Action

- Click the Y-Axis tab.
- 2 **a.** Select the appropriate curve from the list.
  - b. Select the Fixed option in the Scale for curve number field.



**a.** Type the desired minimum and maximum values.

Step

Ac	Action		
b.	Click the <i>All With This Unit</i> button if you want other curves with the same Y-axis units as the current scaled curve to be similarly scaled.		
	Note:		
	The values will only be applied to existing curves. They will not be applied to new curves created after this function was last used.		
Cli	ck the <b>OK</b> button to apply the changes.		

### Select Y-axes to display

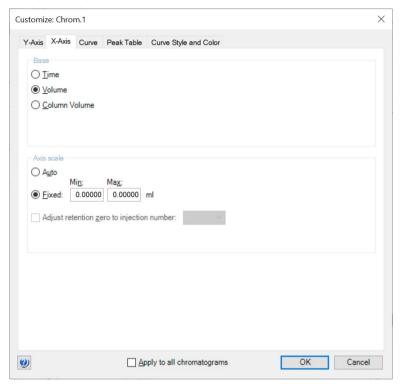
Follow the instructions below to select the Y-axes to display in the chromatogram.

Step	Action
1	Click the <b>Y-Axis</b> tab.
2	<ul> <li>Select a curve for the left Y-axis in the <i>Left Axis</i> drop-down list.</li> <li>To add a second Y-axis, proceed with step 3. If not, jump to step 4.</li> </ul>
3	Select a second curve from the <b>Right Axis</b> drop-down list.
4	Click the <b>OK</b> button to apply the changes.

### Change and fix the X-axis

Follow the instructions below to change and fix the scale of the X-axis in the *Customize* dialog:

Click the X-Axis tab.



- 2 Select the appropriate option in the **Base** field:
  - a. Time
  - b. Volume
  - c. Column Volume

### Note:

Some calculated curves, for example baselines, exist in only one base and might seem to disappear when the base is changed. Curves are collected in time and recalculated for display in volume. Thus, switching the base between **Time** and **Volume** can slightly alter the resolution of the displayed curves.

- Click the *Fixed* option in the *Axis scale* field to set the axis limits manually.
  - **b.** Type the desired minimum and maximum values.

### 6.4.3 Change the axes

### Step Action

**c.** If desired, select the **Adjust retention zero to injection number** checkbox and choose an injection number from the drop-down list.

### Note:

This sets the time/volume to zero at the selected injection mark. The time and volume before selected injection will become negative values.

4 Click the **OK** button to apply the changes.

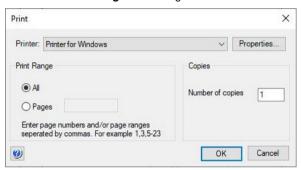
## 6.4.4 Print chromatograms and peak data

#### Introduction

This section describes how to print the chromatograms that are open in the **Evaluation** module.

## The Print Chromatograms dialog box

The **Print Chromatograms** dialog is illustrated below.



#### Instructions

Follow the instructions below to print active chromatograms.

#### Step Action

- Open all chromatograms to be printed.
- Select File →Print.

or



Result: The Print Chromatograms dialog opens.

- Click the **OK** button to open the **Print** dialog and proceed with step 6.
  - Proceed with step 4, to preview and edit the layout.

#### Note:

Click and drag the chromatogram tabs to change the order of the chromatograms on print.

4 Click the **Preview** button.

- 6.4 Optimize the presentation of chromatograms
- 6.4.4 Print chromatograms and peak data

## Step Action

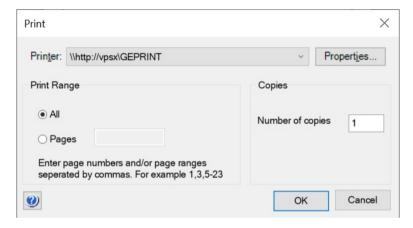
Result:

The Customize Report window opens.

Select File →Print.
 or

Click the toolbar icon.

Result: The Print dialog opens.



#### Note:

It is possible to print directly from the **Customize Report** window or click the **Exit** button to return to the **Print Chromatogram** dialog.

- 6 **a.** Select the print range and number of copies.
  - **b.** If necessary, click the **Properties** button to open the printer properties dialog where the settings for the printer can be changed.
  - **c.** Click the **OK** button to print the chromatograms.

## 6.4.5 Save report as PDF

To save the report as PDF, select *File* → *Report As PDF*.

Result: The report is created as a PDF file and saved in the location specified in the dialog.

**Note:** A PDF report can also be generated by clicking the



(icon) in the toolbar of the Evaluation module.

# 7 Evaluate and compile the results

## **About this chapter**

This chapter describes how to perform several editing and evaluation operations on the results, for example:

- Peak integrations
- Baseline operations (editing and optimization)
- · Fraction and peak operations
- · Comparison of different runs

This chapter also describes how to import and export results.

For information about how to view results, see *Chapter 6 View and present the results, on page 115*.

## In this chapter

Section		See page
7.1	Peakintegration	149
7.2	Baseline operations	171
7.3	Fraction and peak operations	187
7.4	Compare different runs	193
7.5	Rename folders, results, chromatograms, curves and peak tables	204
7.6	Save results	205
7.7	Import and export results	206

# 7.1 Peak integration

#### About this section

Peak integration is used to identify and measure several curve characteristics, including peak areas, retention time and peak widths.

## In this section

Section	n	See page
7.1.1	Perform a peak integration	150
7.1.2	Baseline calculation	153
7.1.3	Display peak data	154
7.1.4	Edit the integration parameters	157
7.1.5	Integrate part of a curve	169
7.1.6	Exclude peaks	170

- 7.1 Peak integration
- 7.1.1 Perform a peak integration

## 7.1.1 Perform a peak integration

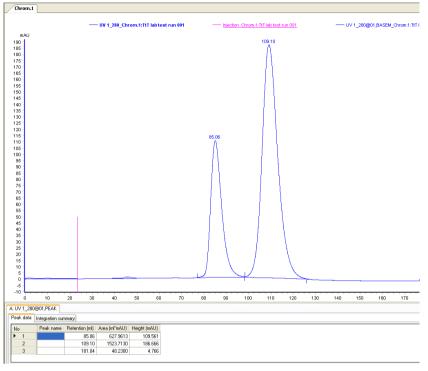
Follow the instructions below to perform a basic peak integration.

Step	Action
1	Open a result file in the <b>Evaluation</b> module.
<ul> <li>Choose Integrate → Peak Integrate.</li> <li>or</li> <li>Click the</li> <li>toolbar icon.</li> </ul>	
	Result: Peak integration of the UV curve is performed.

## **Peak integration results**

The peak table is displayed below the active chromatogram. The start point and end point of each peak are marked by vertical marks, *drop-lines*, in the chromatogram. The peaks are automatically labelled according to what is selected in the *Curve Style and Color* tab of the *Chromatogram Layout* dialog box. A summary of the integration and the settings used can be viewed by clicking the *Integration summary* tab.

The result after a peak integration is illustrated below:



Note: Peak tables can be copied from one chromatogram to another by selecting

Edit →Copy between chromatograms. The copied peak table name has
the default suffix COPY.

## Edit peak integration data

#### Step Action

1 Click the



(icon) or select *File* → *Integrate* → *Edit Peak Table*.

Result:

The Edit Peak Table dialog opens.

- 2 Edit different parameters of the curve from the **Edit Peak Table** dialog:
  - Section 7.1.2 Baseline calculation, on page 153
  - Section 7.1.4 Edit the integration parameters, on page 157
  - Section 7.1.6 Exclude peaks, on page 170
  - Section 7.2 Baseline operations, on page 171

## 7 Evaluate and compile the results

- 7.1 Peak integration
- 7.1.1 Perform a peak integration

Step	Action
	Result:
	The peaks are integrated and updated automatically.

#### 7.1.2 Baseline calculation

#### Introduction

The first step when integrating peaks is to calculate a baseline. A correct baseline is important for accurate calculation of the peak areas. The baseline is per default calculated using the morphological algorithm, when a one-click peak integration is performed. This section describes the options available to calculate baselines manually.

## **Baseline options**

UNICORN start 1.3 offers several options to create an accurate baseline:

- Classical algorithm
- Morphological algorithm
- Zero baseline

#### The Calculate baseline function

Automatic calculation of the baseline can be performed using the **Classical** or **Morphological** algorithm, the latter being the default one. In most cases, the measurement is very accurate.

#### Zero baseline

The baseline will appear as a straight line at zero absorbance.

- 7 Evaluate and compile the results
- 7.1 Peak integration
- 7.1.3 Display peak data

## 7.1.3 Display peak data

#### Introduction

There are several options available to display and edit peak data and peak table contents. This section describes some of these options. Other options are described in Section 7.1.4 Edit the integration parameters, on page 157.

#### Peak characteristics

The peak retention times and several other peak characteristics are calculated automatically. Follow the instructions below to display other peak characteristics.

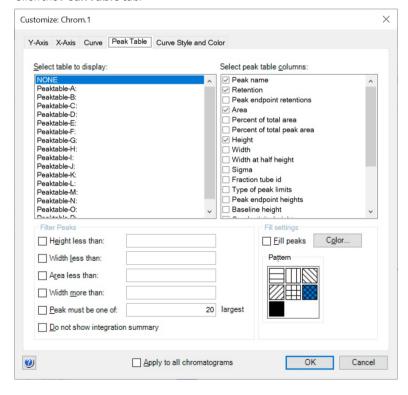
#### Step Action

- a. Right-click in the active chromatogram.
  - **b.** Select **Customize** from the shortcut menu.

Result:

The Customize dialog opens.

2 Click the **Peak Table** tab.



Step	Action
3	a. Select options from the <b>Select peak table columns</b> list.
	<b>b.</b> Click the <b>OK</b> button.
	Result:
	The selected items are displayed in the peak table.

## How to filter peaks

Peaks can be excluded from display in a peak table. Follow the instructions below to filter the peaks:

Step	Action
1	<ul> <li>Right-click in the active chromatogram or peak table.</li> <li>Select <i>Customize</i> from the shortcut menu.</li> </ul>
	Result: The <b>Customize</b> dialog opens.
2	Click the <b>Peak Table</b> tab.
3	<ul> <li>Click the check boxes in the <i>Filter Peaks</i> field to select the filter criteria.</li> <li>Specify filter values.</li> <li>Click the <i>OK</i> button.</li> </ul>

## Filter peaks or reject peaks

The table below describes the major differences in the effects of filtering peaks compared to excluding the peaks by rejection.

Filter peaks	Reject peaks
excludes the peaks from display,	permanently excludes peaks from the integration,
does not exclude the peaks from the calculation of the total peak area,	excludes the peaks from the calculation of the total peak area,
can be reversed.	cannot be reversed.

## **Measurement options**

It is possible to determine the coordinates of any point on a curve and to obtain values for retention and peak height. This is a useful tool for many other functions, such as for measuring the parameters used in baseline calculations.

Coordinates can be obtained in two ways:

- 7 Evaluate and compile the results
- 7.1 Peak integration
- 7.1.3 Display peak data
  - Through direct measurement using the **Vertical marker**. This is described in Section 6.3.1 Open and view results, on page 123.
  - From peak table data.

## 7.1.4 Edit the integration parameters

#### Introduction

Once a peak table has been generated based on an appropriate baseline, it is possible to split or join peaks and to manually adjust the peak start and end points. The peaks are then automatically renumbered, and the peak values recalculated.

These changes can be made in the *Edit Peak Table* dialog. In this dialog, peak names that may be used as labels in the chromatogram can also be added.

## Open the peak table for editing

Follow the instructions below to open the peak table for editing. The editing options are described below the instructions.

Step	Action	
1	Select <i>Integrate</i> → <i>Edit Peak Table</i> .	
	Result:	
	The <b>Edit Peak Table</b> dialog opens.	
	Note:	
	If there are several peak tables in the result, the dialog opens for the peak table tab that is selected. The name of the baseline on which the peak table is based is displayed in the dialog.	
2	Perform the changes (described in the instructions below).	
3	Click the <b>OK</b> button.	
	Result:	
	The <b>Save Edited Peak Table</b> dialog opens. The dialog displays a suggested name and location for the peak table.	
4	Confirm the name and location and click the <b>OK</b> button.	

The **Edit Peak Table** dialog is opened immediately if **Save and Edit Peak** 

## The Edit Peak Table dialog

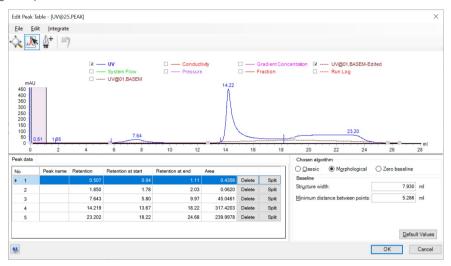
Note:

The illustration below shows the **Edit Peak Table** dialog:

Table is selected.

#### 7 Evaluate and compile the results

- 7.1 Peak integration
- 7.1.4 Edit the integration parameters



# Toolbar icons in the Edit Peak Table dialog

The table below describes the toolbar icons in the Edit Peak Table dialog.

Icon	Function
<b>\$</b>	Activates the <b>Zoom</b> mode. When it is activated, you can use the mouse pointer to drag out an area to be zoomed in. This is described in <i>The zoom function</i> below.
	Activates the <b>Edit Peaks</b> mode. In this mode, you can select a peak and edit the start and end points. This is described in <i>Edit the peaks manually</i> below.
<b>+</b>	Activates the <b>Set Curve Points</b> mode. In this mode, you can select baseline curve points for editing and add new points to re-draw the baseline. This is described in <i>Adjust the baseline</i> below.
7	Returns the result to the state it was in before the last change ( <i>Undo</i> ).  Note:
	This function can also be selected in the <b>Edit</b> menu.

#### The zoom function

Follow the instructions below to use the zoom function in the Edit Peak Table dialog.

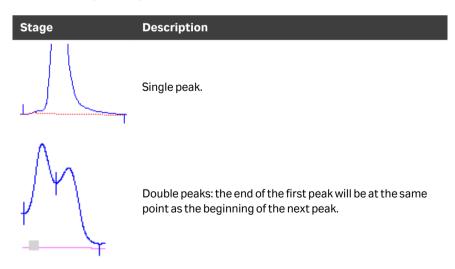
Step	Action	
1	Click the <b>Zoom</b> icon.	
	Result:	
	The mouse pointer is changed into .	
2	Position the mouse pointer over the left topmost position of the area you want to zoom in.	
	<ul> <li>Press and hold the left mouse button.</li> </ul>	
3	Drag the cursor over the area you want to zoom in.	
4	Release the mouse button.	
	Result:	
	The area is enlarged.	

# Note:

A **Zoomed mode** note will be displayed in the top right-hand corner of the chromatogram when the zoom function has been used. Right-click and select **Reset Zoom** from the shortcut menu to return to the full display.

## Peak start and end points

The beginning of each peak is marked with a dropline above the curve, and the end of each peak is marked with a dropline below the curve. The table below shows examples of start and end point droplines:



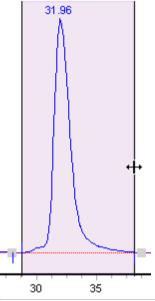
- 7 Evaluate and compile the results
- 7.1 Peak integration
- 7.1.4 Edit the integration parameters

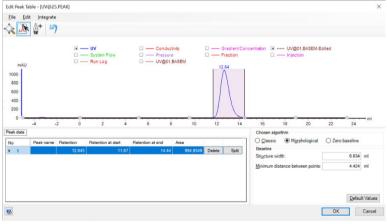
## Edit the peaks manually

The start and end points of a peak can be adjusted graphically by moving the droplines in the *Edit Peak Table* dialog.

Step	Action
1	Click the <b>Edit Peaks</b> icon.
	Result:
	The mouse pointer is changed into $\bigcirc$ .
2	Click the peak you want to edit.
	Result:
	The peak is highlighted and marker lines show the start and end points.
3	Position the mouse pointer over the marker line that you want to change.
	Result:
	The pointer is changed into a double-arrow symbol.

#### Step Action





4 Drag the marker to a new position with the mouse pointer.

#### Result:

The start or end point droplines of the peak are changed and the corresponding values are updated in the peak table.

**Note:** A dropline can never be moved beyond another dropline or beyond a point where the peak meets the baseline.

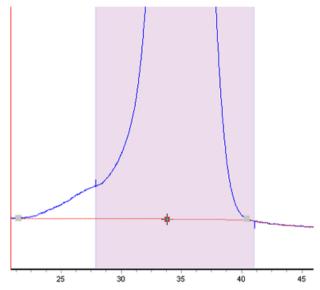
- 7.1 Peak integration
- 7.1.4 Edit the integration parameters

## Adjust the baseline

The baseline can be adjusted graphically in the *Edit Peak Table* dialog.

# Step Action 1 Click the Set Curve Points icon. Result: The mouse pointer is changed into +.

- 2 Perform one or more of the operations below as desired:
  - Click to insert a new data point.



- Double-click on a data point or right-click the point and select **Delete Point** from the shortcut menu to delete the point.
- Click a data point and drag the point to a new position to move the baseline.

#### Result:

The baseline is edited and the peak table values are recalculated accordingly.

**Note:** Accept negative peaks must be selected before the peak integration if you want to be able to drag a data point to move the baseline above the curve.

#### Calculate a new baseline

The baseline can be recalculated in the **Edit Peak Table** dialog. To alter the baseline settings:

		7.1.4 Edit the integration parameters
Step	Action	

- 1 Select an algorithm (*Morphological* is default).
- Adjust the **Baseline** parameters as desired.
  - Structure width
  - Minimum distance between points

or

- Click the **Default Values** button for the default values.
- Click **OK**.

Result:

The baseline is recalculated.

Note:

Select **Zero Baseline** and click the **Apply Zerobaseline** button to replace the calculated baseline with a zero baseline.

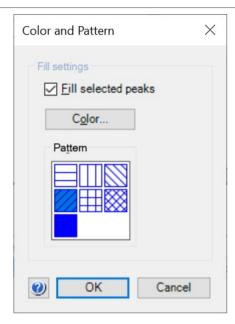
## Add color to a peak

The table below describes how to add a fill color and a pattern to an individual peak in the *Edit Peak Table* dialog:

Step	Action
1	Click the <b>Edit Peaks</b> icon.
	Result:
	The mouse pointer is changed into $\checkmark$ .
2	Click to select the peak.
3	Right-click and select <i>Fill Peaks</i> from the shortcut menu
	or
	<ul> <li>Select Edit → Fill Peak.</li> </ul>
	Result: The <b>Color and Pattern</b> dialog opens.

- 7 Evaluate and compile the results
- 7.1 Peak integration
- 7.1.4 Edit the integration parameters

#### Step Action



4 Select the *Fill selected peaks* check box.

Result:

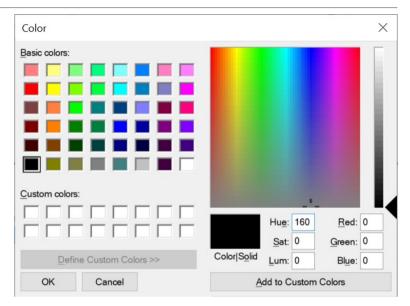
The **Color** button and **Pattern** field are activated.

5 Click the **Color** button.

Result:

The **Color** dialog opens.

#### Step Action



- Select one of the Basic colors
  - or
  - Click the **Define Custom Colors** button to open the settings field and define new colors (illustrated above).
  - After you have selected a color, click **OK** to close the dialog.
- 7 **a.** Select a fill pattern in the **Pattern** field.
  - b. Click OK.

Result:

The peak is filled according to the selections.

#### Note:

The color and pattern selections will override the general **Fill settings** that can be selected for all peaks on the **Peak Table** tab in the **Customize** dialog for the chromatogram.

## Split a peak

It is possible to split a peak into two new peaks by inserting a dropline. Follow the instructions below to split a peak in the *Edit Peak Table* dialog:

#### Step Action

1 Click the **Edit peaks** icon.

#### 7.1 Peak integration

#### 7.1.4 Edit the integration parameters

Step	tep Action	
	Result:	
	The mouse pointer is changed into $\bigcirc$ .	
2	Click the peak in the curve or in the peak table to select the peak.	
3	Right-click and select <i>Split Peak</i> from the shortcut menu	
	or	
	<ul> <li>Select Edit →Split Peaks.</li> </ul>	
	Result: A new dropline is inserted at the middle point between the two existing droplines and the peak is split. The peak numbering and the peak table are updated accordingly.	
Note:	The area under each new peak will not be the same if the symmetry of the original peak was not perfect.	

## Join peaks

It is possible to join the areas of adjacent peaks if they are separated by a dropline. Follow the instructions below to join adjacent peaks in the *Edit Peak Table* dialog:

Step	Action
1	Click the <b>Edit peaks</b> icon.
	Result:
	The mouse pointer is changed into $\checkmark$ .
2	Click the peak in the curve or in the peak table to select the peak.
3	<ul> <li>Right-click and select <b>Join Left</b> or <b>Join Right</b> from the shortcut menu or</li> </ul>
	<ul> <li>Select Edit → Join Left or Edit → Join Right.</li> </ul>
	Result: The original intervening dropline is removed, all peaks are renumbered, and the peak table is updated.

## Add peak names

Follow the instructions below to add names in the *Edit Peak Table* dialog, to identify the peaks:

7.1.4 Edit the integration parameters

#### Step Action

1 Click the **Edit peaks** icon.

Result:

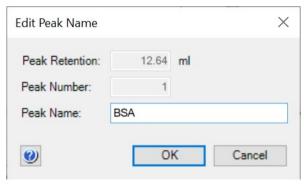
The mouse pointer is changed into

2 Click the peak in the curve or in the peak table to select the peak.



- Right-click and select **Peak Name** from the shortcut menu
  - Choose Edit →Peak name or
  - Double-click the peak.

Result: The **Edit Peak Name** dialog opens. The number and retention of the selected peak is displayed.



4 Type a name in the **Peak name** textbox.

#### 7.1 Peak integration

#### 7.1.4 Edit the integration parameters

Step	Action
5	Click <b>OK</b> .
	Result:
	The new name is added in the corresponding <b>Peak name</b> cell in the peak table. It is also shown below the peak in the chromatogram. If you have selected to show peak names in the chromatogram, it will also be displayed over the peak.

#### Note:

You can also add a peak name by selecting the **Peak name** cell in the peak table, click again in the cell and then type the peak name. A pencil symbol is displayed by the peak number to indicate that you are able to enter the text.

## **Delete peaks**

Follow the instructions below to delete a peak in the *Edit Peak Table* dialog:

Step	Action
1	Click the <b>Edit Peaks</b> icon.
	Result:
	The mouse pointer is changed into $\bigcirc$ .
2	Click the peak in the curve or in the peak table to select the peak.
3	Right-click and select <i>Delete Peaks</i> from the shortcut menu or
	Select <i>Edit</i> → <i>Delete Peaks</i>
	or
	Press the <b>Delete</b> key.
	Result: The peak is deleted, the remaining peaks are renumbered, and the peak table is updated.

## The Integrate menu

If needed, you can use the selections on the *Integrate* menu to perform a peak integration in the *Edit Peak Table* dialog box. This is useful for example if you want to re-integrate the curve using different settings or integrate only part of a curve with different settings.

## 7.1.5 Integrate part of a curve

#### Introduction

There are several possibilities to improve the results if the peak integration is unsatisfactory. This section describes how to select only part of a curve for integration.

This partial integration can be performed both in the *Integrate* dialog in preparation for the peak integration, or in the *Edit Peak Table* dialog to adjust an unsatisfactory peak integration. Both alternatives are described here.

## Select part of a curve for integration

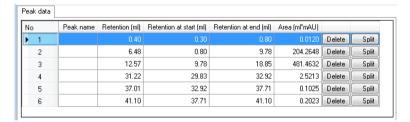
Follow the instructions below to select only part of a curve for peak integration in the *Integrate* dialog box:

#### Step Action

- Choose Integrate → Peak Integrate
  - or
  - · Click the



icon.



#### 2 Click OK.

#### Result:

The baseline will be calculated from the whole curve, but the calculation of the peak areas is only performed on the selected section.

## 7.1.6 Exclude peaks

#### Introduction

The results of the peak integration can sometimes be improved if some peaks are excluded. Also, more accurate results may be obtained if smaller peaks are skimmed off larger peaks with shoulders, as described in this section.

#### **Exclude peaks**

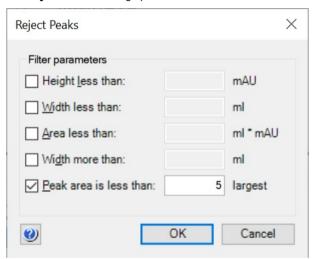
Follow the instructions below to define peaks to be excluded in the **Peak Integrate** dialog during a peak integration.

#### Step Action

Click **Settings** in the tool bar.

Result:

The Reject Peaks dialog opens.



- Select the appropriate checkboxes and type values for peak height, width, and area.
  - **b.** Define how many of the largest peaks you want to include.
- 3 Click **OK**.
- 4 Proceed with the peak integration.

# Note: You can also exclude peaks from the peak integration in the Edit Peak Table dialog. Select Integrate →Settings to open the Reject Peaks dialog.

# 7.2 Baseline operations

#### About this section

To achieve the best possible peak integration result, the baseline may have to be edited or optimized. The edit baseline operations can be accessed through the edit peak dialog.

## In this section

Section		See page
7.2.1	Edit the baseline manually	172
7.2.2	Optimize the baseline with a morphological algorithm	175
7.2.3	Optimize the baseline with a classic algorithm	179

## 7.2.1 Edit the baseline manually

#### Introduction

The first choice when you want to optimize the peak integration is to change the baseline parameters. This section describes how to optimize the baseline manually.

#### How to use the zoom function

Follow the instructions below to use the zoom function in the *Edit Baseline* dialog.

Step	Action
1	Click the <b>Zoom</b> icon.
	<b>\$</b>
	Result:
	The cursor is changed into a magnifying glass.
2	Press and hold the left mouse button.
	<ul> <li>Drag the cursor over the area you want to zoom in.</li> </ul>
	Release the mouse button.
	Result:
	The area is enlarged. Right-click and select <b>Reset zoom</b> to restore the full view.

## How to edit and insert data points

Follow the instructions below to edit and insert baseline data points:

Step	Action
1	Select Integrate → Edit Baseline.
	Result:
	If there is more than one baseline available, the <b>Select Baseline to Edit</b> dialog opens. If not, proceed to step 2.
	Select the baseline you want to edit from the list.
	• Click <b>OK</b> .
	Result:
	The <b>Edit Baseline</b> dialog opens

## Step Action

2 Click the **Set Curve Points** icon.



#### Result:

The mouse pointer is changed into a pen symbol and a cross.

- 3 Add, delete and/or move data points according to the table below.
- 4 Click **OK**.

Result:

The Save Edited Baseline dialog opens.

- Confirm the location and type a new name if necessary.
  - Click **OK**.

Result:

The new baseline is saved.

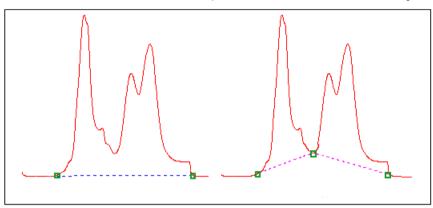
If you want to	Then
Add a data point	Click the left mouse button to place a new baseline point in the chromatogram.
	Result: A new point is created, marked by a red square. The baseline curve is redrawn as a smooth spline function based on the old and the new points. The baseline is guided by the points, but does not necessarily pass through them.
	Tip:
	If you want to create straight baselines, read the instruction later in this section.
Delete a data point	Click the data point to select it and click the <b>Delete</b> button.
	Result: The data point is deleted and the curve is redrawn.
	Note:
	Click the <b>Delete All</b> button to delete all the data points at once.
Move a data point	Select the data point and drag it to a new position.
	Result: The baseline curve is redrawn.

- 7 Evaluate and compile the results
- 7.2 Baseline operations
- 7.2.1 Edit the baseline manually

**Tip:** Click the **Undo** button to cancel the last edit if the result is not satisfactory.

#### **Edited baseline**

The illustration below is a simulated example of a baseline before and after editing:



## How to draw a straight line

Follow the instructions below to force a straight baseline between two points.

Step	Action
1	Select the first of the two points in the point list.
2	Click the <b>Draw straight to next point</b> button.
	Result:
	The baseline is drawn through the points as a straight line.

#### Optimize the baseline with a morphological algorithm 7.2.2

#### Introduction

The first choice when you want to optimize the peak integration is to change the baseline parameters. This section describes how to optimize the baseline with a morphological algorithm.

#### Morphological algorithm description

A morphological algorithm can be described as a line that follows the chromatogram parallel to the X-axis. Data points for the baseline are created whenever the line touches the curve, and the points are joined at the end to create a baseline.

A morphological algorithm gives the best result in curves with drifting baseline and peak clusters. The morphological baseline follows the curve faithfully, and a curve with a baseline at a more even level can be created by subtracting the morphological baseline.

A morphological algorithm does not work well if there are negative peaks or if quantitative data from negative peaks are important in the run.

Note: **Morphological** algorithm is the default baseline setting.

## How to set a morphological baseline

Follow the instructions to choose a **Morphological** algorithm and define baseline settings.

Step	Action
1	Clickthe
	button or select $\textit{Integrate} \rightarrow \textit{Edit Peak Table}$ to perform peak integration.
	Select $Integrate \rightarrow Edit Peak Table$ to perform peak integration.
2	• Choose <i>Calculate baseline</i> in the <i>Baseline</i> drop-down menu.
	Click the <b>Baseline settings</b> button.
	Result:
	The <b>Baseline Settings</b> dialog opens.
3	• Select the <i>Morphological</i> algorithm.
	• Change the <b>Baseline</b> parameters if necessary.
	See more information about the parameters below this table.
	• Click <b>OK</b> .

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## Morphological algorithm parameters

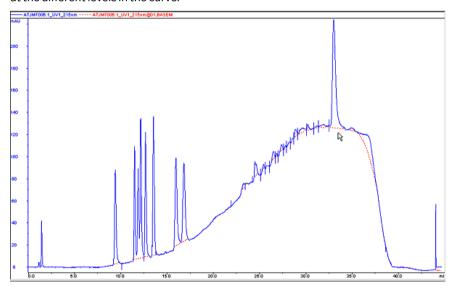
The parameters for the *Morphological* algorithm are:

- · Structure width
- · Minimum distance between points

#### Structure width

**Structure width** determines the length of the straight line that follows the chromatogram. The default value is set at the widest peak in the chromatogram multiplied by 1.5.

The illustration below is an example of how a morphological baseline follows the peaks at the different levels in the curve:



## The correct structure width settings

## **Settings too low**

**Structure width** settings that are too low can result in a baseline that reaches too high up in the peaks of the curve. Sometimes a wider peak is not recognized because it contains a cluster of smaller peaks. The **Structure width** is then set to a value according to the largest width of the identified narrower peaks, and must be increased.

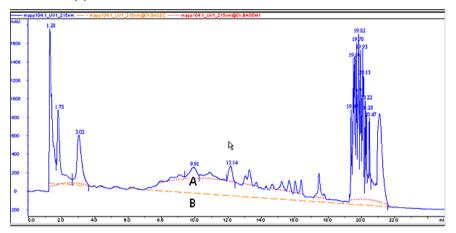
## Settings too high

**Structure width** settings that are too high mean that narrower peaks, especially in fluctuating curves, are not properly followed. This happens when an artifact in a curve is identified as the widest peak by the morphological algorithm, and then is used to set the default **Structure width** value.

7.2 Baseline operations

7.2.2 Optimize the baseline with a morphological algorithm

The illustration below is an example of baselines using the default morphological algorithm settings (A) and a morphological algorithm with an increased **Structure width** value (B).



#### Minimum distance between points

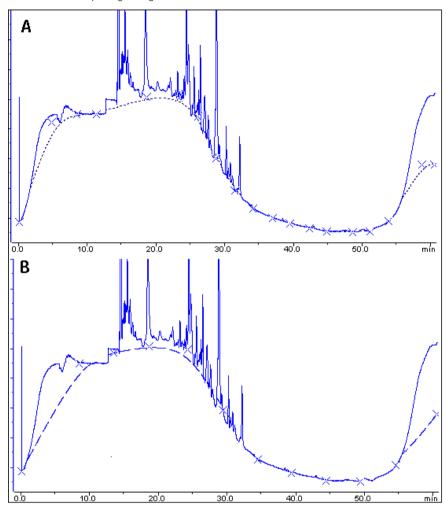
The *Minimum distance between points* is a measure of the distance between the data points used to generate a baseline. The largest number of data points is produced at the slopes of the curves. If you increase the *Minimum distance between points* value, fewer points will be collected on the slopes.

The illustration below is an example of a baseline (A) that is created with the *Minimum distance between points* parameter set at a low value. The number of data points is reduced when the *Minimum distance between points* parameter is set to a higher value (B).

## 7 Evaluate and compile the results

#### 7.2 Baseline operations

## 7.2.2 Optimize the baseline with a morphological algorithm



## 7.2.3 Optimize the baseline with a classic algorithm

# Optimize the baseline with a classic algorithm

#### Introduction

7.2.3

The first choice when you want to optimize the peak integration is to change the baseline parameters. This section describes how to optimize the baseline with a classic algorithm.

#### Classic algorithm description

A classic algorithm searches for all parts of the source curve that are longer than a defined minimum baseline segment and fall within limiting parameters. Together, the parameter values define the limits for a rectangular box. A part of the source curve must fit entirely inside this rectangular box to be identified as a baseline segment.

A classic algorithm is particularly useful when you need to integrate curves with negative peaks and when quantitative data from negative peaks are important.

#### Classic algorithm parameters

The parameters for the *Classic* algorithm are:

· Shortest baseline segment

the dialog.

- Noise window
- Max baseline level
- Slope limit

See more information about the parameters below.

#### How to set a classic baseline

Follow the instructions below to set a *Classic* algorithm and define a baseline.

Step	Action
1	Click the <b>Baseline settings</b> button in the <b>Peak Integrate</b> dialog.
	Result:
	The <b>Baseline Settings</b> dialog opens.
2	Select the <i>Classic</i> algorithm.
	Change the <i>Baseline</i> parameters.
	See more information about the parameters below this table.
	• Click <b>OK</b> .
Tip:	The same settings can be edited in the <b>Calculate Baseline</b> dialog when a

new baseline is created. Choose **Integrate** →**Calculate Baseline** to open

- 7.2 Baseline operations
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#### **Test your parameter changes**

The best way to optimize the baseline is to change the baseline parameters step by step and then check the resulting baseline after each change. When the desired effect is accomplished it is best to go back and try a parameter value in between the two last settings to avoid an unnecessarily low or high value.

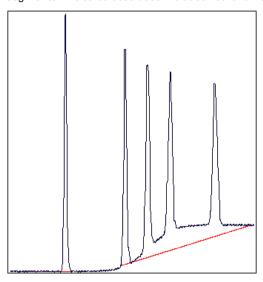
How much the values should be changed depends on the cause of the peak integration problem. The table below is a general guideline.

Baseline parameter	Recommended initial change
Shortest baseline segment	20%-50%
Noise window	10%-30%
Max baseline level	Usually not necessary to adjust
Slope limit	25%-50%

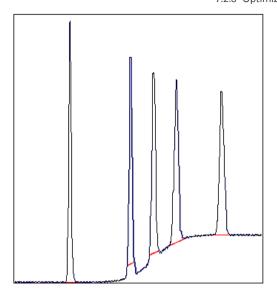
**Tip:** If necessary, click the **Default Values** button to restore the default values.

#### Shortest baseline segment

If an excessively high **Shortest baseline segment** value is set, short curve segments between peaks in the middle of the chromatogram are not identified as baseline segments. The calculated baseline does not follow the source curve, see below:



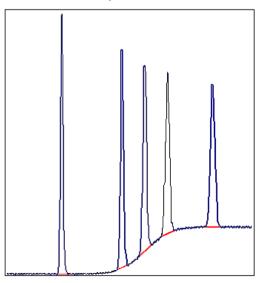
The **Shortest baseline segment** value is decreased by 50% in this example:



#### **Slope limit**

A changed **Slope limit** will often improve the baseline calculation. The **Slope limit** sets the maximum slope of the curve to define when a peak is recognized. An excessively high **Slope limit** will cause the up-slopes of the peaks to be recognized as baseline segments.

The example above was improved by the shorter baseline segments but the high slope of the short segments in the region between the second and the fourth peak still makes the baseline unacceptable. In the example below the **Slope limit** is increased by a factor of 2.5, which produces a correct baseline:



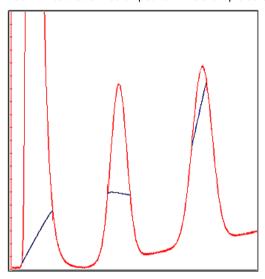
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#### Slope limit too high

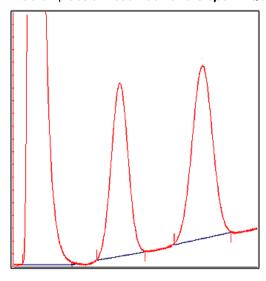
An excessively high **Slope limit** value can cause peak limits too high up on the peaks. This can be the case when the chromatogram includes a very large flowthrough or solvent peak. The large peak affects the calculation of the default parameters and leads to excessively high values for the **Slope limit**.

**Note:** An excessive value for the **Noise window** can have the same effect and be caused by the same situation, often also in combination with a high **Slope limit**.

Peak limits are defined on peaks in the example below due to the high **Slope limit**:

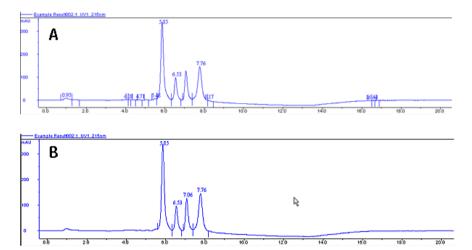


The example below has a much lower **Slope limit**, and a lower **Noise window**:



#### **Noise window**

Occasionally too many peaks occur after the peak integration, usually because noise on the baseline is erroneously detected as peaks. The solution to this is to increase the **Noise window** parameter. However, this can result in peak limits too high up on the peak slopes. The illustration below is an example of noise detected as peaks (A) and the result of a second peak integration with an increased **Noise window** (B).



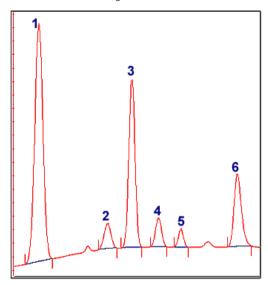
**Tip:** You can also use the **Reject peaks** function in the **Peak Integrate** dialog to reduce the number of peaks based on the total number of accepted peaks or the minimum peak height.

#### Missing peaks

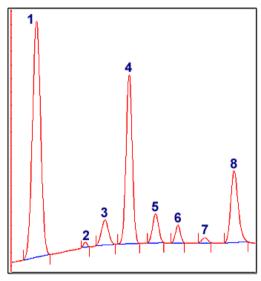
Sometimes obvious peaks are not detected in the peak integration. The probable cause is that the **Noise window** is set too high. See the illustration below:

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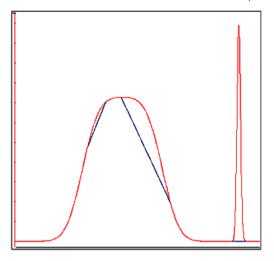
All peaks are detected if the **Noise window** is decreased-see example below:



**Note:** Missing peaks can also be caused by improper settings for **Reject peaks** in the **Peak Integrate** dialog, or **Filter peaks** in the **Chromatogram Layout** dialog.

## When to change the Max baseline level

In rare cases the top of a broad, flat peak can be incorporated as a baseline segment. This is one of the very few situations where it is useful to change the **Max baseline level**. The illustration below is an example:



#### How to set the Max baseline level

Follow the instructions below to set the *Max baseline level*.

Step	Action
1	Right-click in the chromatogram and select <i>Marker</i> .
	Result:
	A vertical line is set in the chromatogram. A text box in the top left corner of the chromatogram displays the X-axis and Y-axis values of the curve at the point where the vertical <b>Marker</b> line crosses the curve.
2	Move the <i>Marker</i> with your mouse.
	• Measure the height of the peak you want to exclude from the baseline.
3	Choose Integrate → Calculate baseline.
4	• Select <i>Classic</i> as <i>Algorithm</i> .
	• Type a new value for <b>Max baseline level</b> . Set the level slightly lower than the value that you measured in step 2.
	• Click <b>OK</b> .

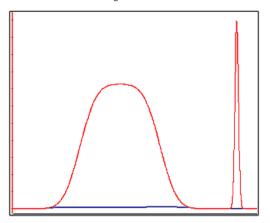
#### Example of a correct baseline

The illustration below is an example of a correct baseline after the *Max baseline level* has been changed:

#### 7 Evaluate and compile the results

#### 7.2 Baseline operations

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#### 7.3 Fraction and peak operations

#### About this section

This section describes additional operations that you can perform on your results:

- Create an average fraction absorbance curve, the *Fraction Histogram*.
- Match protein activity to a curve using the **Activity Histogram**.
- Use the absorbance ratio to verify peak purity or identify peaks.

#### In this section

Section		See page
7.3.1	The Fraction Histogram	188
7.3.2	Match protein activity to a curve	191

#### 7.3.1 The Fraction Histogram

#### Introduction

The **Fraction Histogram** dialog in the Evaluation module can be used to create a curve for the average fraction absorbance. It can also be used to create a table showing the amount of protein and the concentration in each fraction.

#### **Create a Fraction Histogram curve**

Follow the instructions below to create a *Fraction Histogram* curve.

Step	Action
1	Select <b>Operations</b> → <b>Fraction Histogram</b> .
	Result:
	The <i>Fraction Histogram</i> dialog opens.
2	Select the desired UV curve in the left $\textbf{Source chromatogram and curve}$ field.
	Tip:
	The fractions curve should already be selected on the middle field.
3	Click <b>OK</b> .
	Result:
	The average fraction absorbance values are displayed as a new curve in the chromatogram.
Tip:	By default, the <b>Fraction Histogram</b> curve will be assigned to the first available curve position, which is shown in the <b>Target chromatogram and curve</b> list. A default curve name with the ending <b>HIST</b> is suggested in the <b>Curve name</b> field. You can choose another curve position and type another curve name in the <b>Curve name</b> field if desired.

## Protein concentrations in the fractions

The protein concentration in the fractions are calculated using the following formula: Concentration [mg/ml] = A/(d\*1000\*Ext. coef.)

A = Average fraction absorbance = Area / Volume [mAU]

**d** = UV cell path length [cm]

Ext. coef. = Protein coefficient at used wavelength. [I g<sup>-1</sup> cm<sup>-1</sup>]

#### Protein amounts in the fractions

The total amount of protein found in the fraction is calculated using the following formula:

Amount [mg] = Concentration [mg/ml] \* fraction volume [ml]

# Calculate the protein concentration and amount

Follow the instructions below to calculate the protein concentration and amount in the fractions using the *Fraction* table in the *Fraction Histogram* dialog:

Step	Action
1	Select <b>Operations</b> → <b>Fraction Histogram</b> .
	Result:
	The <i>Fraction Histogram</i> dialog opens.
	Note:
	If the path length is not shown, it can be entered manually.
2	Type the extinction coefficient for each fraction in the corresponding <b>Ext. coef</b> table cell.
	Result:
	The fraction protein concentration and amount are calculated and displayed in the corresponding <b>Conc.</b> and <b>Amount</b> table cells.
	Note:
	The values are calculated using a zero baseline (i.e., no baseline subtraction is applied).

#### **Export the Fraction table**

The complete *Fraction* table can be exported as an Excel file:

Step	Action
1	Click the <b>Export Table</b> button in the <b>Fraction Histogram</b> dialog.
	Result: The <b>Export Fraction Table to Excel</b> file dialog opens.
2	Browse to the folder where you want to save the file.
3	Type a name in the <i>File name</i> field.
4	Click the <b>Save</b> button.  Result:
	The table is saved in Excel format.

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- 7.3.1 The Fraction Histogram

#### **Print the Fraction table**

The complete  $\it Fraction$  table can be printed directly from the  $\it Fraction\, Histogram$  dialog:

Step	Action
1	Click the <b>Print Table</b> button.
	Result:
	The <b>Print</b> dialog opens.
2	If necessary, select a printer and the number of copies to print. You may also click the <b>Properties</b> button in this dialog, to change the general printer settings.
3	Click <b>OK</b> .
	Result:
	The table is printed on the selected printer.

#### 7.3.2 Match protein activity to a curve

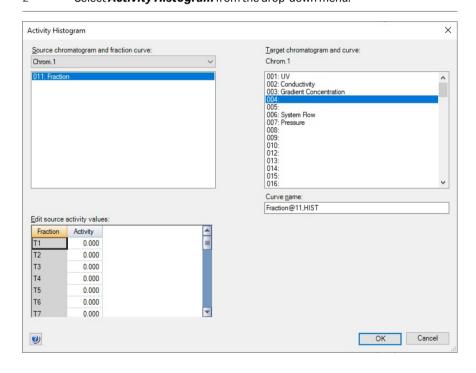
#### Introduction

You can compare data from the results of protein activity assays, such as ELISA, with the data contained in the UV curve. The activity curve and the UV curve can be compared in a combined presentation.

#### The Activity Histogram dialog

To open the Activity Histogram dialog:

# Step Action Click Operations. Select Activity Histogram from the drop-down menu.



# Enter protein activity values for comparison

Follow the instructions to enter the values from a protein activity assay in a comparison histogram:

Step	Action
1	Choose <b>Operations</b> → <b>Activity Histogram</b> .
	Result:
	The <b>Activity Histogram</b> dialog opens.
2	By default, the fraction curve for the current chromatogram is selected in the <b>Source chromatogram and fraction curve</b> list.
	If necessary, change the source and target chromatograms.
	All the component fractions of the fraction curve are listed in the <b>Fraction</b> column of the <b>Edit source activity values</b> field.
3	• Type an activity value for each fraction in the <i>Activity</i> column.
	• Click <b>OK</b> .
	Result:
	A histogram curve showing the activity values is added to the chromatogram.

Tip: By default, the Activity Histogram curve will be assigned to the first available curve position, which is shown in the Target chromatogram and curve list. A default curve name with the ending HIST is suggested in the Curve name field. You can choose another curve position and type another curve name in the Curve name field if you so wish.

#### 7.4 Compare different runs

#### About this section

This section describes how to:

- Make comparisons between curves or chromatograms from different runs.
- Present curves or chromatograms from different runs.
- Compare curve parameters among curves from different runs.
- View several chromatograms at the same time.
- Overlay curves from different runs in one chromatogram.
- Stack curves from different runs in one chromatogram.
- Create mirror images

#### In this section

Sectio	n	See page
7.4.1	Open and compare chromatograms	194
7.4.2	Open and compare curves	196
7.4.3	Shift curves	201
7.4.4	Create a mirror image	202

#### 7.4.1 Open and compare chromatograms

#### Introduction

This section describes how to:

- Import chromatograms from other result files to an open result.
- Compare chromatograms in a result.

# Import chromatograms with the command File →Add to compare

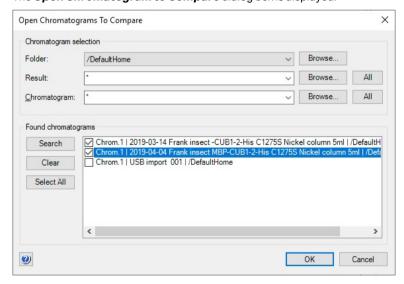
Follow the instructions below to import chromatograms with the *File* → *Add to compare* command. The search is performed at specific locations or with specific search criteria. This method is useful if, for example, you want to import chromatograms from all results of a specific folder.

#### Step Action

 Choose File →Add to compare →Chromatograms in the Evaluation module.

Result:

The **Open Chromatogram to Compare** dialog box is displayed.



2 a. Click the *Browse...* button beside the *Folder* drop-down list This opens a browse dialog where you can select the result folder where you want to search for chromatograms.

#### Step Action

#### Note:

The search will not include the contents of subfolders. You must select the subfolders individually for the search.

b. Repeat this step to define specific results or chromatograms if desired.

#### Tip:

Click the drop-down list arrow buttons to view search paths that you have used previously. You can click selections in these lists to search the same locations again.

- 3 a. Click the Search button in the Found chromatograms field and a list of chromatograms will be displayed based on the designated search criteria.
  - **b.** A new search can be performed with new search criteria without erasing the initial chromatograms from the list.
  - c. Select the chromatograms that you want to import. If you click the Select All button, all the displayed chromatograms will be imported.
  - d. If you want to clear the list of displayed chromatograms, click the Clear button.
- 4 a. Click the OK button.

#### Result:

All the selected chromatograms are opened in the **Evaluation** module.

#### Note:

If the names of the imported chromatograms are already used they will be sequentially numbered for identification purposes. Each chromatogram will be opened in its own tab. However, depending on the number of chromatograms and the length of the names, not all tabs may be visible at once. Click the arrow symbol on the top right corner of the chromatogram pane to show a list of all open chromatograms. You may select the chromatogram you want to view from this list.

## Display all chromatograms on the same scale

Step	Action
1	Click the <b>Customize</b> icon or select <b>Tools</b> → <b>Customize</b> to open the <b>Customize</b> dialog for any chromatogram.
2	Select the <b>X-Axis</b> tab and make the changes to the <b>Base</b> and the <b>Axis scale</b> .
3	Select the <i>Apply to all chromatograms</i> option and click <i>OK</i> .

#### 7.4.2 Open and compare curves

#### Introduction

This section describes how to import or copy curves from different runs into one chromatogram for comparison.

#### File:Add to compare

Follow the instructions below to import curves to a chromatogram with the command  $File \rightarrow Add$  to compare:

#### Step Action

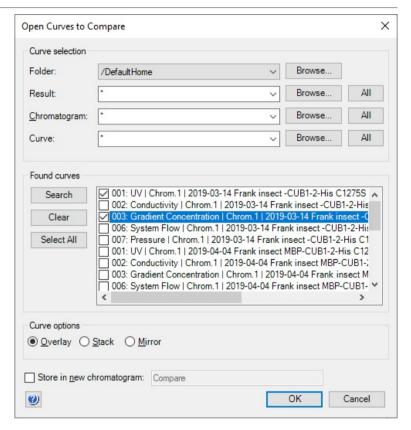
- 1 In the **Evaluation** module,
  - Choose File →Add to compare → Curves
    or
  - Click the **Open curves to compare** toolbar button.



Result:

The Open Curves to Compare dialog opens.

#### Step Action



- Select the desired search criteria in the Folder, Result, Chromatogram and Curve drop-down list of the Curve selection section.
  - Click Search and a list of curves will be displayed based on the selected search criteria.

#### Tip:

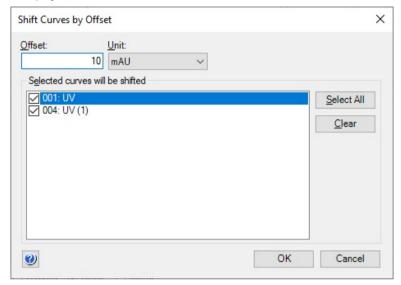
A new search can be performed with new search criteria without erasing curves located in the previous search.

- Select the check boxes for the curves that you want to import. Click the Select All button if you want to import all the curves.
- If you select the **Store in new chromatogram** option, the curves will be imported into a new chromatogram. This is recommended to keep the source chromatogram free of too many additional curves.
- 3 a. Select how to display the imported curves in the Curve options field. The options are described in "Curve options" below.
  - b. Click OK.

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- 7.4 Compare different runs
- 7.4.2 Open and compare curves

#### Step Action

If you selected the **Stack** option in step 3, the **Shift Curves by Offset** dialog is displayed:



- You can set the Offset value to increase or decrease the offset distance between the curves.
- b. Click OK.

#### Result:

Depending on your previous choices, the imported curves are now displayed in the source chromatogram or in a newly created chromatogram.

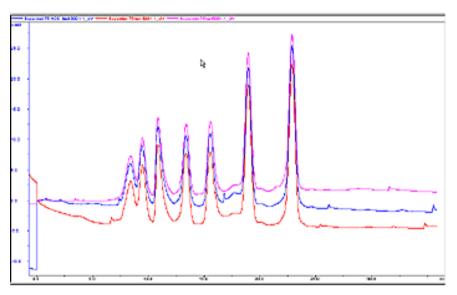
#### Note:

If curves with several different units have been selected, the curves with each different unit will be grouped together with a separate offset from the other groups.

#### **Curve options**

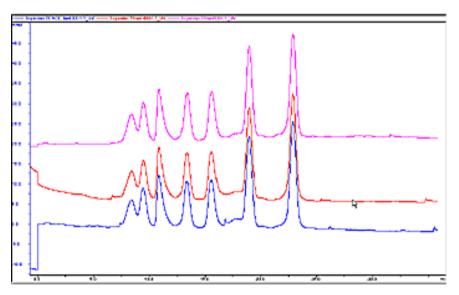
The illustrations below show the different curve presentation options. The curves can be overlaid, stacked or mirrored.

#### Overlay



The curves are presented overlaid on one another.

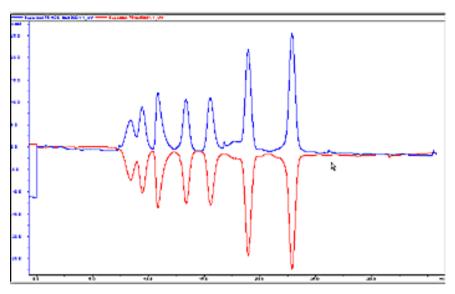
#### Stack



The curves are presented with a given offset Y-axis value so that the curves are stacked and distinct from one another.

- 7 Evaluate and compile the results
- 7.4 Compare different runs
- 7.4.2 Open and compare curves

#### Mirror



For comparison of two imported curves. One curve is inverted in the Y-axis and thus appears to mirror the other curve.

#### **Change comparison settings**

Follow the instructions below to change other comparison settings:

Step	Action
1	Click the <b>Customize</b> icon to open the <b>Customize</b> dialog.
2	Select or de-select the check boxes on the $\it {\it Curve}$ tab to compare a different set of curves.
3	<ul> <li>Select the <i>Y-Axis</i> tab to change the curve scales either</li> <li>individually</li> </ul>
	or - all with the same scale (click the <i>All With This Unit</i> button). • Click <i>OK</i> to update the curves.

#### 7.4.3 Shift curves

#### Introduction

You can use the **Shift** function to stack and move curves from different runs to better visualize the differences. Each curve is repositioned along the x- or y-axis by a precise value and the instruction is logged in the evaluation log.

**Note:** The **Shift** function requires that the curves are present in one chromatogram.

#### Move a curve with the Shift function

Follow the instructions below to use the **Shift** function:

Step	Action	
1	Make sure that a chromatogram with the relevant curves is open in the Evaluation module.	
	<ul> <li>Choose Operations → Shift.</li> </ul>	
	Result:	
	The <b>Shift</b> dialog is displayed.	
2	Select the curve to be shifted in the <b>Source chromatogram and curve</b> list.	
	• Select a curve position in the <b>Target chromatogram and curve</b> list.	
	<ul> <li>Type a new Curve name or accept the default. The default curve name ending is SHFT.</li> </ul>	
3	Select the axis/axes along which the shift is to be made:	
	<ul> <li>along the X-axis (Shift retention)</li> <li>along the Y-axis (Shift amplitude).</li> <li>Type the shift value(s).</li> </ul>	
	• Click <b>OK</b> .	
4	Repeat steps 1 to 3 to shift other curves in the chromatogram.	

1

#### 7.4.4 Create a mirror image

#### Instruction

A very useful way to compare the features of two curves is to produce a mirror image of one curve. To create a mirror image of a chromatogram:

#### Step Action

- Open a chromatogram to insert the relevant curves into.
  - Choose File → Open to Compare → Curves.

#### Result:

The **Open Curves to Compare** dialog is displayed.

- Select the desired search criteria in the Folder, Result, Chromatogram and Curve drop-down list of the Curve selection section.
  - Click Search and a list of found curves will be displayed based on the selected search criteria.

#### Tip:

A new search can be performed with new search criteria without erasing curves located in the previous search.

- Select the checkboxes for the curves that you want to import.
  - Select Mirror in the Curve options field.
  - Click OK.

#### Result:

The curves are displayed with one of the curves as a mirror image in the active chromatogram window.

#### 4 Shift the mirror image curve downwards

For an improved presentation:

a. Choose Operations → Shift.

Result:

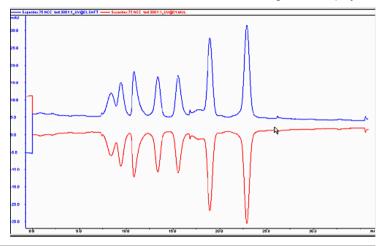
The **Shift** dialog box is displayed.

- **b.** Select the curve to be shifted in the **Source chromatogram and curve** list.
- c. Select the same curve number in the *Target chromatogram and curve* list box as in step 2.
- d. Select the Shift amplitude checkbox since the shift is to be made along the Y-axis.
- e. Type a shift value.
- f. Click OK.

#### Step Action

#### Result:

The illustration below shows a curve and a mirror image curve displayed.



#### 7.5 Rename folders, results, chromatograms, curves and peak tables

#### Rename folders and files

Folders and files may be renamed in the **Result Navigator** either by

- Right-clicking the object and choosing **Rename** from the shortcut menu or
- Choosing *Edit* → *Rename* → *Navigator Object*

This will highlight the name of the selected object and you can type a new name instead.

Note: You cannot rename assigned home folders.

#### Rename chromatograms, curves, and peak tables

Follow the instructions below to rename chromatograms, curves, or peak tables in the Evaluation module.

Step	Action	
1	Choose <i>Edit</i> → <i>Rename</i> and the relevant option <i>Chromatogram</i> , <i>Curve</i> or <i>Peak Table</i> .	
	Result:	
	The <b>Rename</b> dialog opens.	
2	Select the appropriate object.	
	• Type a new name in the <b>Name</b> field.	
	• Click <b>OK</b> .	
Note:	The original chromatogram or original raw data curves cannot be renamed.	

They will not be available in the **Rename** dialog.

#### 7.6 Save results

#### Introduction

After you have finished the evaluation process, you can save all the changes you have made to the chromatograms, including newly created curves and chromatograms that you have imported and created.

#### **Delete unwanted curves**

All the curves that you created during your manipulations will be saved in the chromatogram. If some of these curves are no longer needed, select **Edit**  $\rightarrow$ **Delete**  $\rightarrow$ **Curves** in the **Evaluation** module to remove the curves.

**Note:** The original curves that were created during the run can never be deleted.

#### Save the results

You can either save your edited results in the original file or in a new result file. The table below describes how to save the results in the **Evaluation** module.

If you want to save the edited results	then
In the original result file	<ul> <li>Select <i>File</i> → <i>Save</i>, or</li> <li>Click the <i>Save</i> toolbar icon.</li> </ul>
In a new result file	Select <i>File</i> → <i>Save as</i> .

Note:

The previous version of the result file will be overwritten if you save the changes. This cannot be reversed. However, the raw data curves remain unchanged.

#### **Exit the Evaluation module**

The table below describes how to exit the **Evaluation** module:

If you want to	then
Close only the <b>Evaluation</b> module	Click the red cross icon in the top right corner of the module window.
Exit UNICORN start 1.3	Choose <i>File</i> → <i>Exit UNICORN start</i> .

Note:

If there are unsaved changes, a dialog box opens with an option to save the changes before exit.

### 7.7 Import and export results

#### About this section

Curves and data can be imported and exported in different formats. This chapter describes how to import and export results.

#### In this section

Section		See page
7.7.1	Import results	207
7.7.2	Export results	209

#### 7.7.1 Import results

#### **Import curves**

Individual curves saved in ASCII format (as \*.asc or \*.txt files) may be imported into the active chromatogram. Follow the instructions below to import curves:

Step	Action
1	Choose <i>File → Import → Curves</i> .
	Result:
	The <i>Import</i> dialog box opens.
2	Locate the file that contains the curve and click the file.
	• Click the <b>Open</b> button.
	Result:
	The $\it Import Curves$ dialog opens, showing the available curves in the file.
3	Select the curve(s) to import.
	• Click the <b>OK</b> button.
	Result:
	The curves are opened in the active chromatogram.
Notos	A currie can only be imported in the base unit that it was experted in Far

#### Note:

A curve can only be imported in the base unit that it was exported in. For volume base, the curve is shown in chromatogram with ml and CV as x-axis. For time base, the curve is shown in in chromatogram with min as x-axis.

#### **Import results**

Results that have been exported from a UNICORN start 1.3 database may be imported into the UNICORN start 1.3 database again. Follow the instructions below to import results:

Step	Action
1	• Choose File →Import →Entire Result
	Result:
	The <i>Import</i> dialog opens.
2	• Locate and click the zip-file that contains the result. To select several files, press <b>Shift</b> while you click the files.
	Click the <i>Open</i> button.
	Result:
	The <i>Import Result</i> dialog opens.

#### 7.7.1 Import results

Step	Action
3	Select the destination folder for the result.
	Click the <i>Import</i> button.
	Result:
	The result files are individually unzipped and imported into the selected folder.
Note:	A result file cannot be imported into a folder if the folder already contains a result with the same name. In that case you must rename the file.

#### 7.7.2 Export results

#### Introduction

This section describes how to export entire results and result items, including

- Curves
- Documentation
- Peak tables

The section also describes how to copy items using the Windows clipboard.

#### **Export options**

Select  $File \rightarrow Export$  in the Evaluation module to export data from an open result file. The following export options are available:

If you choose	Then the options are
to UNICORN start	• Curves or • Entire Result
Externally	<ul> <li>Curves</li> <li>Peak Table</li> <li>Documentation or</li> <li>Copy to Meta file</li> </ul>

These options are explained further below.

#### **Export results**

One or the several of the complete results may be exported for use in another UNICORN start database. To export entire results:

Step	Action
1	• Select a result in the <b>Result Navigator</b> . To select several results, press Shift while you click the results.
	<ul> <li>Select File →Export →To UNICORN start →Entire Result.</li> </ul>
	Result:
	The <b>Export to Another UNICORN start Database</b> dialog opens if one result was chosen. The <b>Browse For Folder</b> dialog opens if several results were chosen.

2

#### Step Action

- Result files are exported as compressed .zip files only.
  - Select an export location. Make a new folder if desired.
  - Type a name in the File name field in the Export to Another UNICORN start Database dialog if one result was chosen.

#### Note:

As the results are exported individually, it is not applicable to enter a destination file name if several results are selected for export.

 Click Save in the Export to Another UNICORN start Database dialog or OK in the Browse For Folder dialog.

#### Result:

The result is saved as a zip file. If several results were exported, each result is saved as an individual zip file.

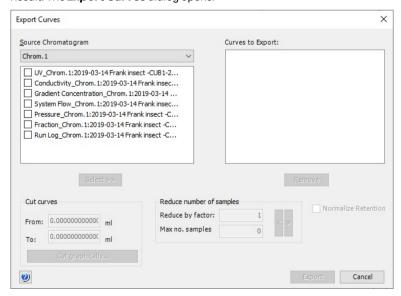
#### **Export curves**

Follow the instructions below to export curves in the *Evaluation* module.

#### Step Action

- Choose *File* → *Export* → *To UNICORN* → *Curves*.
  - or
  - Choose File → Export → Externally → Curves

Result: The Export Curves dialog opens.



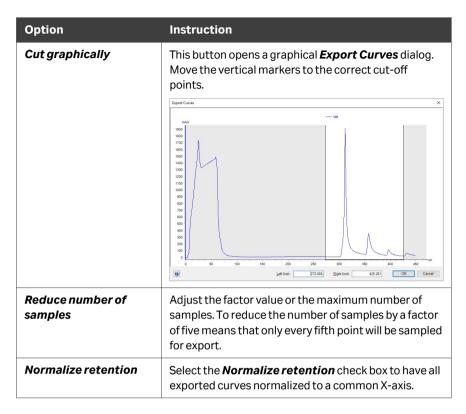
Step Action	
2	a. Select the curve(s) you want to export.
	<b>b.</b> Enter parameters to limit the curve(s) if necessary (described in "Limit the exported curves" below).
	c. Click the <b>Select &gt;&gt;</b> button.
	<b>d.</b> Repeat this step to select more curves.
3	Click the <b>Export</b> button.  Result:
	The <b>Export Curves to File</b> dialog opens.
4	Select the export file format from the <b>Save as type</b> drop-down list.
	ASCII files (*.asc)
	or
	Comma Separated Values (*.csv)
5	a. Select a destination folder.
	<b>b.</b> Type a file name and click <b>Save</b> .
Note:	Curves are exported as series of numerical coordinates that refers to the

#### Limit the exported curves

You can optimize the exported curves to only the parts that you want to focus on, in the Export Curves dialog. The table below describes how to use these editing options.

time/volume and signal respectively.

Option	Instruction
Cut curves	Enter retention values in the text boxes to limit the curve to only a portion of the original curve.



#### **Export peak tables**

Follow the instructions below to export peak tables.

# Step Action Choose File → Export → Externally → Peak Table. Result: The Export Peak Table dialog opens. Select the source chromatogram and the peak table you want to export. • Click the Export button. Result: The Export Peak Table To File dialog opens.

- 3 Select the export file format from the **Save as type** drop-down list:
  - ASCII files (\*.asc)
  - Excel files (\*.xls)
  - XML files (\*.xml)

Step 4	Action	
	Select a destination folder.	
	Type a file name.	
	• Click <b>Save</b> .	

Note:

Peak tables are exported as text strings in ASCII or XML format and numerical values in the Excel format.

# Export the chromatogram to a Windows meta file

You can export an image of the open chromatogram as a Windows enhanced meta file. This image format can be used in other applications, for example *Microsoft Word* or *PowerPoint*®. To export a chromatogram to a Windows meta file:

Action
• Choose File → Export → Externally → Copy to Meta file
or
Right-click in the chromatogram and choose <i>Copy to Meta file</i>
Result: The Export Chromatogram to Windows Enhanced Meta File dialog opens. The only available file type is Enhanced Meta File (*.emf)
Select a destination folder.
Type a file name.
• Click <b>Save</b> .

#### Copy to the clipboard

You can use the Windows clipboard to copy the contents of the active window and paste it into other programs. To copy to the clipboard:

Step	Action
1	• Choose <i>Edit</i> → <i>Copy</i>
	or
	Right-click and choose Copy to Clipboard
	or
	• Click the <b>Copy</b> icon

3

Click the **OK** button.

# Result: If the active window only contains a chromatogram, it will immediately be copied to the clipboard as a Windows meta file. If the active window also contains a peak table, the Copy to Clipboard dialog opens. Select one of the options: Copy curve window as enhanced meta file Copy selected peak table data as tabbed text

# 8 USB export and import operations with ÄKTA start

#### **About this chapter**

This chapter describes how to export a method from ÄKTA start and how to import results generated with ÄKTA start (without UNICORN start 1.3).

#### In this chapter

Section		See page
8.1	Export a method to a USB memory stick	216
8.2	Import results from a USB memory stick	217

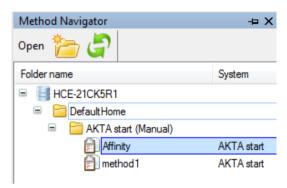
#### 8.1 Export a method to a USB memory stick

#### Instructions

Follow the instructions below to export a method to a USB memory stick:

#### Step Action

1 In the **Method Navigator** pane, select the method to be exported. It is only possible to export one method at a time to an external file.



2 Choose File → Export → Export Method To USB.

Result:

The File Explorer dialog opens.

- 3 Locate the USB flash drive and enter a file name.
- 4 Click the Save button.

#### Note:

Please refer to ÄKTA start firmware and UNICORN start Compatibility matrix document to export methods from UNICORN start 1.3 to older versions of ÄKTA start firmware with a USB device.

# 8.2 Import results from a USB memory stick

### Instructions

Results can also be imported from a ÄKTA start instrument into the UNICORN start 1.3 database using a USB stick. To import results from a USB stick:

Step	Action
1	Select File →Import →Import Result from USB.
	Result:
	The <i>Import</i> dialog opens
2	Locate the USB drive.
	<ul> <li>Locate and click the zip-file that contains the result. To select several files, press Shift while you click the files.</li> </ul>
	• Click the <b>Open</b> button.
	Result:
	The <i>Import Result</i> dialog opens.
3	Select the destination folder for the result.
	• Click the <i>Import</i> button.
	Result:
	The result files are individually unzipped and imported into the selected folder.
Note:	Please refer to ÄKTA start firmware and UNICORN start Compatibility matrix document to import results from older versions of ÄKTA start firmware to UNICORN start 1.3 with a USB device.

# 9 Administration

## In this chapter

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## 9.1 System administration

## **Purpose**

UNICORN start 1.3, system, run and result related logs provide the system administrator with a full record of UNICORN start 1.3 usage and system activity.

## **Different types of logs**

There are two types of logs:

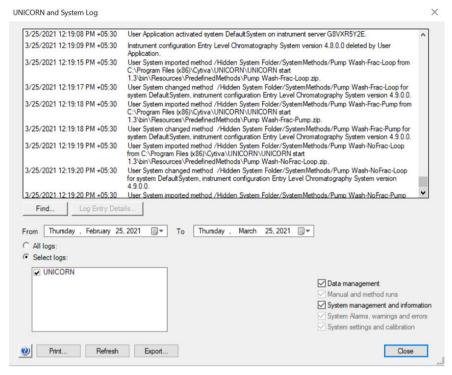
- UNICORN start 1.3 log which records all system independent events,
- System log which records events related to the ÄKTA start.

UNICORN start and system logs are available in the UNICORN start 1.3 *Administration* module, by clicking the *UNICORN and System Log* icon.



## **UNICORN** and System Log dialog

The UNICORN and System Log dialog is illustrated next.



**Tip:** Click the **Refresh** button to update the displayed events to the latest records.

## View selected log entries

By default, the **UNICORN start and System Log** dialog shows all the log entries for UNICORN start 1.3, ÄKTA start up to a month before the current date. Specific log entries can be selected so that only the items of interest are displayed. It is also possible to combine selections to narrow down the displayed items, for example, to the log entries for a specific user at a specific system on a specific date.

The table below describes how select specific log entries for display:

To view log entries	Then
From a specific time period	<ul> <li>Click the down arrow by the <i>From</i> field and select the starting date for the time period in the calendar and</li> <li>Click the down arrow by the <i>To</i> field and select the ending date.</li> </ul>

To view log entries	Then
For specific systems	<ul> <li>Click the Select logs option button and</li> <li>Select UNICORN start from the list below.</li> </ul>
From the UNICORN start log	Click the Select logs option button and Select UNICORN start from the list below.
For specific events	Select one or several of the items below:
	<ul> <li>Data management         (folder, method, and result activities, for example, create, copy, changes, etc.)     </li> <li>Manual and method runs         (run start and end, created results, etc.)     </li> <li>System management and information         (connections, session start and end, create new, activate or deactivate systems, etc.)     </li> <li>System alarms, warnings and errors         (alarm and error descriptions, acknowledgement and action records, etc.)     </li> <li>System settings and calibration</li> </ul>
	(changed settings and calibration actions)

## Long log entries

Long entries in the logs are sometimes only shown in part, ending with ellipsis ("...."). To show the full log entry in a separate window:

Step	Action	
1	Select the entry and double-click <b>Log Entry Details</b> .	
	Result:	
	The full log entry is shown in a separate window.	
Tip:	Log Entry Details is only enabled when a log entry is selected.	
Tip:	Complete entries are always included in printed logs.	

## Find specific log entries

You can search for specific log entry text by clicking the *Find* button and entering text to search for in the *Find what* field of the *Find* dialog:



## **Export logs**

You can export the selected log entries to an XML file by clicking the *Export* button. Using the XML file format, the log entries can be imported into XML-based reporting systems.

## **Archive logs**

To reduce the size of the logs, older records can be archived. The archiving procedure is part of the **Database Management** routines, and it is described in **Archive data**, on page 231. The archived logs can be retrieved by using the **Database Management** retrieval procedure.

**Note:** It is possible to retrieve logs to the same database that they were originally

archived from. That is, the retrieval procedure cannot be used to import logs

from one database to another.

**Note:** It is recommended that all archived logs be saved on a different hard drive

than where the active database is stored.

## **Print the logs**

Follow the instructions below to print selected log entries:

Step	Action
1	Select the log entries to print.
2	Click the <b>Print</b> button.
	Result:
	The <b>Print</b> dialog opens.
3	a. Select a printer from the drop-down list.
	<ul> <li>b. If desired, click the <b>Properties</b> button and select printer settings and</li> </ul>
	c. Click the <b>OK</b> button.
	Result:
	The selected log entries are printed.

3/25/2021 12:19:19 PM +05:30:

User System imported method /Hidden System Folder/SystemMethods/Pump Wash-NoFrac-Loop from C:\Program Files (x86)\Cytiva\UNICORN\UNICORN start 1.3 \bin\Resources\PredefinedMethods\Pump Wash-NoFrac-Loop.zip. 3/25/2021 12:19:20 PM +05:30:

User System changed method /Hidden System Folder/SystemMethods/Pump Wash-NoFrac-Loop for system DefaultSystem, instrument configuration Entry Level Chromatography System version 4.9.0.0.  $3/25/2021\ 12:19:20\ PM\ +05:30$ :

User System imported method /Hidden System Folder/SystemMethods/Pump Wash-NoFrac-Pump from C:\Program Files (x86)\Cytiva\UNICORN\UNICORN start 1.3 \bin\Resources\PredefinedMethods\Pump Wash-NoFrac-Pump.zip.

3/25/2021 12:19:20 PM +05:30: User System changed method /Hidden System Folder/SystemMethods/Pump Wash-NoFrac-Pump for system DefaultSystem, instrument configuration Entry Level Chromatography System version 4.9.0.0.

## 9.2 Database administration

### About this section

This section describes the standard database solution used for data storage and the maintenance procedures required to operate the database.

This section also contains general instructions on how to upgrade the database server software to editions with greater data handling capacity.

### In this section

Section		See page
9.2.1	Database overview	225
9.2.2	Database maintenance	226

#### 9.2.1 Database overview

#### About the UNICORN database

Data storage in UNICORN start 1.3 is handled by a database. The database is a relational database. SQL (Structured Query Language) is the querying language used for the retrieval, insertion, updating, deleting and general management of the data.

The UNICORN start 1.3 database server installation for a stand-alone workstation includes a distributable version of Microsoft SQL Server Express. It is described in *SQL Server Express usage with UNICORN start 1.3, on page 225* below.

The installation and basic set up of the database is an integrated part of the installation sequence for the options including the database server functions. It will not require any user settings other than the initial selection of the option and selection of destination folders for backup and archives.

Normally, the database maintenance functions offered by UNICORN start 1.3 are sufficient for the UNICORN start 1.3 administrator. Using management tools such as SQL Server Management Studio should not be necessary and it is not recommended since improper changes may corrupt the database.

# SQL Server Express usage with UNICORN start 1.3

The list below includes some properties of Microsoft SQL Server Express distributed with UNICORN start 1.3:

- Primarily suited for small-scale applications.
- Up to 4 GB of server RAM can be utilized.
- Utilizes one single physical CPU.
- Operates databases up to 10 GB, excluding log files.

### 9.2.2 Database maintenance

Once the database is installed and a system is running and producing data, it is essential that routines and schedules for backup and archiving of data are established. This section describes how to perform these routines.

### **Database backup**

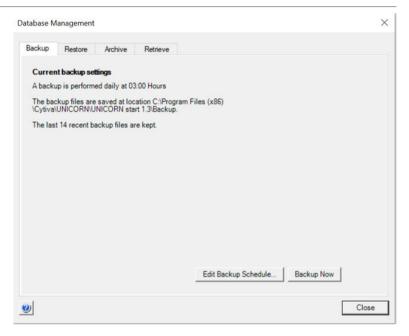
During the database server installation, a target folder for the regular, scheduled backup was set up. By default, backups were also scheduled at the installation. The default backup settings are:

- · Daily backup
- At 03:00 local time
- In the selected target folder
- With the last 14 recent backup files kept.
   (when additional files are saved, the oldest backup is deleted)

# Verify or edit the database backup schedule

Follow the instructions below to schedule the database backup:

Step	Action
1	Click the <b>Database Management</b> icon in the <b>Administration</b> module.
	Result:
	The <b>Database Management</b> dialog opens.



#### Tip:

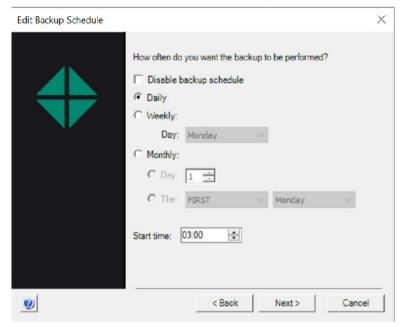
The **Backup** tab shows the current schedule. If the settings are suitable, click the **Close** button at this point.

2 Click the Edit Backup Schedule button.

#### Result:

The Edit Backup Schedule wizard opens.

3 Click the **Next** button to open the next wizard dialog:



4 Select the frequency and the start time for the scheduled backups.

#### Note:

It is possible to disable the scheduled backups at this point.

- 5 Click the **Next** button to proceed.
- 6 Choose the number of backup files that are to be kept.

#### Note:

It is possible to keep all files. However, depending on the number of results that are created, this may cause disk space problems.

7 Click the **Next** button to proceed.

The final wizard page shows a summary of the selected settings.

8 Click the **Finish** button to accept and apply the schedule.

#### Note:

The backup files will be named UNICORN SCHEDULED BACKUP <Date> <Time>.BAK by default.

Note:

It is recommended that another physical drive than where the active database is stored is selected for the scheduled backups at the installation of UNICORN start 1.3. If this is not possible, it is recommended that the backup files are copied to another physical drive or storage media at regular intervals.

Note:

Make sure that the database server computer is turned on at the time when the scheduled backup will occur. If it is not turned on and operational, the backup will fail, it will not be postponed.

### **Manual backup**

Extra backups can be made manually between the scheduled backups. Follow the instructions below to make extra backups:

## Step Action

1 Click the **Database Management** icon in the **Administration** module.

Result:

The **Database Management** dialog opens.

Click the **Backup Now** button in the **Backup** tab.

Result:

A Backup Now confirmation dialog opens.

- 3 Click the **OK** button to start the backup.
  - This process may take several minutes. A progress dialog is displayed while the backup is performed.

Result:

A Backup confirmation dialog opens when the backup is completed. The backup file will be named  ${\tt UNICORN}$ 

start\_MANUAL\_BACKUP\_<Date>\_<Time>.BAK by default.

- 4 Verify that the backup has been performed in the default folder and then either
  - Click the **Close** button

or

• Click Go To Backup File to open the backup folder and access the files.

#### Tip:

It is recommended to copy or move the backup files to an external storage device to minimize the risk for loss of data. The backup also includes the database content and log reports of UNICORN start 1.3.

## Restore backup data

It is possible to restore data from a previous backup, for example, if data has been deleted by mistake. The restored data replaces all the current data in the database with the data generated up to the point of time when the backup was made. All changes performed after the time of the backup will be lost.

This procedure can be used to move data from one database instance to another but all data in the database instance where the backup is restored will be overwritten.

**Note:** Please take the backup folder of the older version of UNICORNStart and place under the given path, as below:

C:\Program Files (x86)\Cytiva\UNICORN\UNICORN
start 1.3\Backup

Follow the instructions below to restore data from a previous backup:

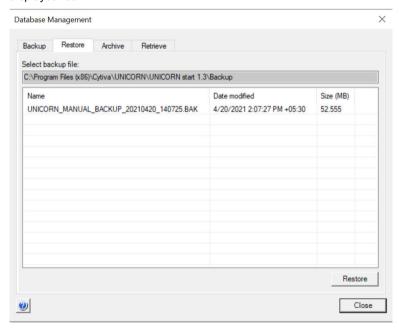
#### Step Action

1 Click the **Database Management** icon in the **Administration** module.

Result:

The **Database Management** dialog opens.

2 Click the **Restore** tab. All the available backup files are shown in the displayed list.



3 Select the backup file to restore and click the **Restore** button.

#### Result:

A warning dialog opens. This dialog suggests that a backup of the database be made before restoring the previous backup. This ensures that all the essential data is saved before it is replaced with the data from the backup.

#### 4 Fither

- Click the **Yes** button to backup before restoring
- Click the No button to proceed without making a new backup.

#### Result:

A second warning dialog opens. This dialog shows the backup file that is restored and explains the implications for the current data. All other UNICORN start 1.3 modules are closed and no other actions may be performed during the restoration.

5 Click the **OK** button to proceed.

#### Result:

A progress dialog is displayed while the restoration is performed. The process is complete when the dialog closes.

#### Note:

Run the **Update Stored Procedure.exe** from the desktop.

#### **Archive data**

When the database size reaches a level where performance is affected and the remaining space is not enough for the immediate storage needs, selected parts of the result data can be archived. This can either be the entire result contents from before a specified date, or selected results. It is also possible to archive logs. It is normally a good idea to archive data that is not immediately needed at regular intervals.

Archiving will reduce the size of the database. It is not the same as exporting result data, which creates copies of the results and does not reduce the size of the database at all.

Follow the instructions below to archive complete result data or logs:

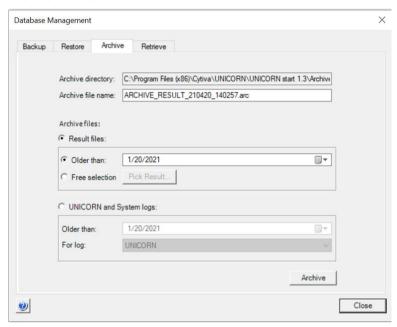
## Step Action

1 Click the **Database Management** icon in the **Administration** module.

#### Result:

The **Database Management** dialog opens.

2 Click the Archive tab.



The archive directory and a suggested archive file name are shown in the dialog. The directory was selected at the installation and cannot be edited here. However, the file name can be changed. The default name suggested by UNICORN start 1.3 includes the date of archiving, and reflects if the selected content is results or logs.

- 3 Choose the data to be archived:
  - If Result files is selected
    - then choose a latest date from the drop-down list calendar or
    - select Free selection and click the Pick Result button.
       (this option is described in Archive selected results, below)
  - **a.** Then choose a latest date from the drop-down list calendar and
  - b. Select the log to archive UNICORN or a system log from the For log list.

If UNICORN and System log is selected

4 Click the **Archive** button.

#### Result:

A warning dialog opens. This dialog suggests that a backup of the database be made before archiving the selected data.

#### Note:

It is recommended to always perform a backup before archiving.

#### 5 Either

- Click the **Yes** button to backup before restoring
- Click the **No** button to proceed without making a new backup.

#### Result:

A progress dialog opens while the data is archived. Depending on the selections above, this may take several minutes. When archiving is completed, a confirmation dialog opens.

6 Click the **Close** button to close the confirmation dialog.

#### Note:

It is recommended to save the archived data on another storage media than where the active database is stored.

#### **Archive selected results**

Follow the instructions below to select individual results for archiving:

#### Step Action

- 1 In the **Database Management** dialog, select the **Archive** tab.
  - Select Result files
  - Select Free selection

and

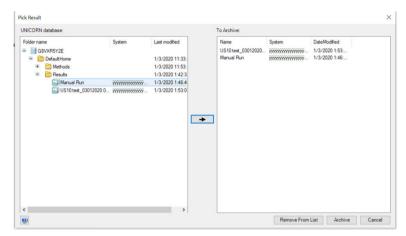
• Click the Pick Result button

#### Result:

The Pick Result dialog opens.

- 2 In the **Pick Result** dialog:
  - Locate and select the result to archive and

• Click the arrow button to add it to the **To Archive** list.



- 3 Repeat step 2 until all the results have been added to the archive list.
  - It is possible to remove results from the list by selecting them and clicking the *Remove From List* button.
- 4 Click the **Archive** button.

#### Result:

A warning dialog opens. This dialog suggests that a backup of the database be made before archiving the selected data.

- 5 Either
  - Click the **Yes** button to backup before restoring or
  - Click the No button to proceed without making a new backup.

#### Result:

A progress dialog opens while the data is archived. Depending on the selections above, this may take several minutes. When the archiving is completed, a confirmation dialog opens.

6 Click the **Close** button to close the confirmation dialog.

#### Retrieve archived data

Results and logs that have been archived can be retrieved from the archive and included in the active database again. Individual results are selected from the archive directory, not the entire archive. However, when logs are retrieved, the entire log is restored.

9.2.2 Database maintenance

**Note:** Only archived results and logs can be retrieved back into the same database

that they were originally archived from. This procedure cannot be used to

migrate data from one database to another.

Tip: Archived results can be shown in the **Evaluation** module **Result** 

**Navigator** by selecting the **Show All** option. This can help in locating the archived results to be retrieved. Archived results are represented by a special icon:



Follow the instructions below to

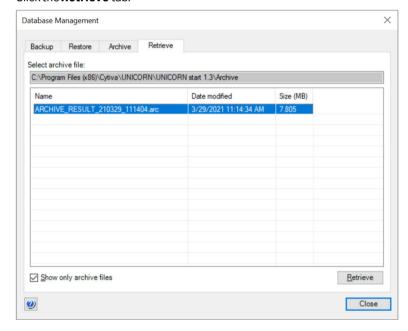
#### Step Action

1 Click the **Database Management** icon in the **Administration** module.

Result:

The Database Management dialog opens.

2 Click the **Retrieve** tab.



#### Note:

Archived results or logs can be retrieved. All archived files have the extension . arc. The **Show only archived files** option displays only these files. By deselecting this option additional files that have the extension . log are also displayed. These log files record the archiving operations and are not connected to the actual UNICORN start 1.3 results or logs.

- 3 Select the archive file to be retrieved and click the **Retrieve** button.
  - a. The Pick Result dialog opens.

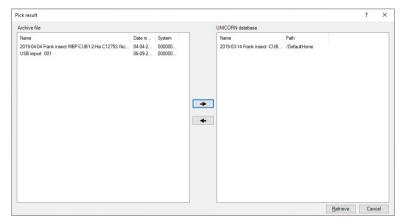
(Proceed with the steps below.)

If a log has been selected to be retrieved:

 The log is restored to the active database and a confirmation dialog is shown.

(This completes the retrieving process.)

If archived results have been selected to be retrieved:



4 Select an archived result in the **Archive file** list and click the right arrow button.

#### Result:

The result is moved to the **UNICORN database** list. The destination folder search path is shown in the list. It is possible to remove selected results from the **UNICORN database** list by clicking the left arrow button.

5 Click the **Retrieve** button in the **Pick Result** dialog.

#### Result:

The results are restored to the active database and a confirmation dialog is displayed.

**Note:** The retrieved results are restored to the folder where they were originally archived from. All archived material is, however, still saved in the archive.

# 10 Troubleshooting

## **About this chapter**

This chapter describes different operational scenarios which may arise in UNICORN start 1.3 and their solutions or consequences.

## In this chapter

Section		See page
10.1	Installing and uninstalling	238
10.2	System connections	240
10.3	System access errors	242
10.4	Database functions	243
10.5	Evaluation errors	244

# 10.1 Installing and uninstalling

Problem description	Solution
SQL Server Express or .net framework installation fails.	Make sure that any other installation or an operating system upgrade is not in progress. Retry the installation after the existing installation is complete. Alternatively, try restarting the computer and install again.
	If the above fails in addition check the following locations for additional clues on SQL Server failure:
	C:\Program Files\Microsoft SQL Server\100\Setup Bootstrap\Log \Summary.txtorfor64-bit operating system
	<pre>C:\Program Files (x86)\Microsoft SQL Server\100\Setup Bootstrap\Log \Summary.txt</pre>
	SQL Server installation can fail due to an incompatible Operating System, a pending reboot, incompatible pre-existing SQL Server version among other reasons.
Uninstalling UNICORN	Please follow the steps below:
start	Uninstall the UNICORNStart Application.
	2. Uninstall Microsoft SQL Server 2008/2014/2019.
	Uninstall the remaining SQL Server from Control Panel.
	Delete the UNICORNStart folder from the installation path.
	5. Delete Microsoft SQL Server from Program Files and Program Files (x86).
	Note:
	Make sure SQL Server is not installed on the PC before deleting these two folders.
	It is recommended to take a backup before uninstalling the UNICORNStart Application.

Problem description	Solution
Unable to configure a license.	Open the .liclicense file and note the address mentioned as the value for HOSTID.
	<ol> <li>Compare this value to the address generated by the configure e-license tool found at <b>Start</b> →<b>Programs</b> →<b>Cytiva</b> menu.</li> </ol>
	3. If none of the addresses match the HOSTID value, you may have an incorrect file or the file was generated for another machine. Contact your Cytiva representative to generate a new license file.
	<b>Note:</b> The UNICORN start license is a Nodelock license and only works on a physical PC.
Prerequisite software fails to install.	Make sure that there is enough disk space on the C:\ drive.
An error message stating that your access rights are insufficient appears when you try to uninstall UNICORN start 1.3 from the <i>Add or Remove Programs</i> dialog in Windows 7 or later.	This is caused by UAC in Windows 7 or later. Turning off UAC may solve this problem. If not, use the software removal procedure from the UNICORN start 1.3 installation DVD.

## 10.2 System connections

#### **Problem description** Solution The connections are not available • Unplug and then plug in the USB cable, even though: wait for a minute and try to reconnect from UNICORN start 1.3. • The connection between the If the above solution fails: PC and the ÄKTA start instrument appears to be Switch off the ÄKTA start instrument. correct. 2. Exit UNICORN start 1.3. • The power is turned on. 3. Restart the computer. • The ÄKTA start instrument touch screen status is depicted Restart the ÄKTA start instrument. as the image below: 5. Log on to UNICORN start 1.3 and try to connect again. ÄKTA start If the connection still does not work, run the UNICORN start Diagnostics *Manager* from the folder *All Programs* →Cytiva →UNICORN start 1.3, for description see section Running the Diagnostics manager. You are connected to a UNICORN Check that the instrument is turned on. start 1.3 system but have no Check that all cable connections are contact with the ÄKTA start intact. instrument. The system hangs at initialization. Do the following steps: The UNICORN start 1.3 system connection is lost when the PC 1. Go to the search bar and type Edit power \Laptop goes to sleep or the plan. laptop lid is closed. 2. Choose Never in the Turn off the display fields for both On battery and Plugged in. 3. Click on Change advanced power settings. 4. Power buttons and lid → Lid close action $\rightarrow$ Plugged in $\rightarrow$ Do nothing.

## Running the Diagnostics Manager

Start the UNICORN start 1.3 *Diagnostics Manager* from the folder *All Programs*→ *Cytiva* → *UNICORN start 1.3*. The following statuses will be displayed:

Status	Description
Port	If the port status is not displayed in green, it may indicate that some other application is blocking the ports reserved for UNICORN start 1.3 communication.
	The process name which is blocking the port will be displayed.
Service	If the service status is not displayed in green, it may indicate that the service used for UNICORN start 1.3 communication and data base operations is not running.
	Try to restart the service from the application.
USB	Examples of status messages:
	USB device is working properly.
	USB device cannot work properly until the computer is restarted.
	USB device is disabled.
	Note:
	There are often multiple USB devices connected to the computer and a failure indication here might not indicate a failure in connecting to ÄKTA start.
	Try the following:
	Unplug and then plug in the USB cable.
	Restart the computer.

# 10.3 System access errors

Problem description	Solution
UNICORN start 1.3 modules are not available for selection in the <b>Log On</b> dialog.	Check that the modules in question are not already open.
The help viewer cannot be opened using help buttons or the <b>F1</b> key.	<ol> <li>Open the <i>MadCap HelpViewer</i> from the Windows desktop icon.</li> <li>MadCap HelpView</li> <li>Try the <i>Help</i> button or <b>F1</b> key again.</li> </ol>
The <i>Microsoft Office Document Image Writer</i> causes UNICORN start 1.3 to terminate.	This writer application will not work. Set another printer driver as the default printer.
	<ol> <li>Open the Windows Control Panel (in Category View).</li> </ol>
	2. Choose <i>Hardware and Sound</i> .
	3. Choose <b>Devices and Printers</b> .
	4. Right-click on an alternate printer and choose <b>Set</b> as <b>default printer</b> from the menu.

## 10.4 Database functions

Problem description	Solution
The database is not available at log on	Wait a couple of minutes to allow the SQL Server services to start and try to log on again.
	(This may be necessary when logging on after a computer re-start)
	Verify in the Windows Services dialog that the SQL Server and SQL Server Browser services are running properly.
The scheduled database backups are not performed	Re-set the scheduled backup in the Database backup wizard.
Unable to connect to the database server.	Verify in the <b>Windows Services</b> dialog that the <b>SQL Server</b> service <b>MSSQL \$UNICORNSTART</b> and the <b>SQL Server Browser</b> service are running.
	If the services are <i>not</i> running, try to start them.
	If the services are running, try to restart the same.

## 10.5 Evaluation errors

# Maximum number of curves exceeded

Problem description	Solution
The maximum number of curves (100) are exceeded when importing curves	The injection curve is also included for each curve that is imported. Delete the unnecessary curves before importing more curves.

## **Distorted peaks**

Problem description	Solution
Peaks appear distorted	A high UV averaging time value will distort and delay the peaks.

## **Retention time differences**

Problem description	Solution
The retention time in the peak table differs from that in the chromatogram.	Verify that the integration was performed on the currently displayed chromatogram.
	Note:
	Only applicable if there are more than one chromatogram in the result.
The retention value in the peak label differs from that in the x-axis.	Verify that the x-axis base has not changed after the peak integration was performed, but is the same as in the peak table. This is applicable for ml and CV bases.

Problem description	Solution
The retention value is not displayed in the peak label.	Verify that the x-axis base has not changed after the peak integration was performed, but is the same as in the peak table. This is applicable for volume and time bases.
	Verify that the chromatogram has not been changed, for example by selecting or deselecting <i>Adjust</i> retention zero to injection number in the Customize dialog, X-Axis tab.

## **General evaluation errors**

Problem description	Solution
Computer restarts abruptly.	If it is a 64 bit computer\laptop there are known incompatibilities between Intel's graphic driver (igdkmd64.sys) and Windows 7. Please ask your administrator to update to the latest graphics driver.

**Note:** In case you are unable to resolve your problem, please contact your local

Cytiva representative.

**Note:** For UNICORN start product updates, please visit: www.cytiva.com/

unicornstart.

# Appendix A

# **Evaluation functions and instructions**

### Introduction

This appendix contains background information about the algorithms and calculation theories applied in UNICORN start.

It also describes the peak table column components and the instructions that are available for automated evaluation procedures.

## In this chapter

Section	on	See page
A.1	Smoothing algorithms	247
A.2	Baseline calculation theory	250
A.3	Peak table column components	256

## A.1 Smoothing algorithms

#### Introduction

This section describes how the smoothing functions are calculated. Choose **Operations** → **Smooth** in the Evaluation module to view and edit the options.

## **Moving average**

The table below describes the process when the  ${\it Moving average}$  smoothing algorithm is used.

Stage	Description
1	For each data point in the source curve, the processed curve is calculated as the average of the data points within a window centered on the source data point.
	• The width of the window is determined by the <i>Filter Parameter</i> value, expressed as number of data points.
2	When the source point is less than half the window size from the beginning of the end of the curve, the average is calculated symmetrically round the source point over as many data points as possible.
	• If you increase the window width, the smoothing effect is also increased.

#### Note:

The filter algorithm only accepts odd integer parameter values between 1 and 151. If an even number has been given, it is incremented by adding the number one (1).

## **Autoregressive**

The table below describes the process when the **Autoregressive** smoothing algorithm is used:

Stage	Description
1	The first data point in the source curve is copied to the processed curve.
2	For each subsequent data point, the previous processed point is multiplied with the filter parameter value and added to the current source data point.

### Stage Description

3 The result is then divided by the parameter value plus 1 according to the following formulae:

$$t_1 = S_1$$

$$t_n = \frac{(p \star t_{n-1} + S_n)}{(p+1)}$$

#### Where:

t<sub>n</sub> = current processed point

 $t_{n-1}$  = previous processed point

 $S_n$  = current source point

p = smoothing parameter value

#### Note:

If you increase the parameter value, the smoothing effect is also increased.

Note:

The filter algorithm only accepts integer parameter values between 1 and 25.

#### Median

The table below describes the process when the *Median* smoothing algorithm is used.

#### Stage Description

- 1 For each data point in the source curve, the processed curve is calculated as the median of the data points within a window centered on the source data point.
  - The width of the window is determined by the parameter value, expressed as number of data points.
- When the source point is less than half the window size from the beginning of the end of the curve, the median is calculated symmetrically round the source point over as many data points as possible.
  - If you increase the window width, the smoothing effect is also increased.
  - To completely remove a noise spike, the window width should in effect be slightly more than twice the width of the spike.

Note:

The filter algorithm only accepts odd integer parameter values between 1 and 151. If an even number has been given, it is incremented by adding the number one (1).

## Savitzky-Golay

The table below describes the process when the **Savitzky-Golay** smoothing algorithm is used.

Stage	Description	
1	The algorithm is based on performing a least squares linear regression f of a polynomial of degree k over at least k+1 data points around each point in the curve to smooth the data.	
	The derivative is the derivative of the fitted polynomial at each point.	
	The calculation uses a convolution formalism to calculate 1st through 9th derivatives.	
2	The calculation is performed with the data in low X to high X order.	
	If the input trace goes from low to high, it is reversed for the calculation and is re-reversed afterwards.	

**Tip:** See Gorry, Peter A, General Least-Squares Smoothing and Differentiation by the Convolution (Savitsky-Golay) Method (Analytical Chemistry 1990, Volume 62, 570-573) for information about the Savitzky-Golay algorithm.

## A.2 Baseline calculation theory

#### Introduction

There are two methods to calculate the baseline, based on a morphological and classic algorithm.

#### **Overall process**

The table below describes the overall process of a baseline calculation.

Stage	Description
1	The baseline segments are defined.
2	The baseline points are selected.
3	The baseline is drawn.

### **Baseline segment definition**

Baseline parameters are used to find the baseline segments. The default values for the parameters are determined from the source curve. The baseline segments are found by different parameters that are based on the type of algorithm that is selected.

Tip:

The parameters can be displayed in the Evaluation module if you choose **Integrate** →**Calculate baseline** function. You can also click the **Baseline settings** button in the **Integrate** →**Peak integrate** dialog.

## Morphological algorithm

The *Morphological* algorithm searches for all parts of the source curve where:

- The curve parts come into contact at both ends of a horizontal line of the length defined in the **Structure width** parameter. The default value of this parameter is based on the widest detected peak in the curve. The horizontal line is moved along the curve up the peak until it reaches the contact points. The curve parts below the horizontal line and the line will now form a "curve" with a plateau. The center point in the plateau formed by the horizontal line will be the data point for the baseline.
- The data points fulfil the Minimum distance between data points. This
  parameter reduces the total number of data points that are created from a curve.

## Classic algorithm

The *Classic* algorithm searches for all parts of the source curve where:

The curve parts are longer than the **Shortest baseline segment**. This parameter
determines the minimum length for a part of the source curve to be considered a
possible baseline segment.

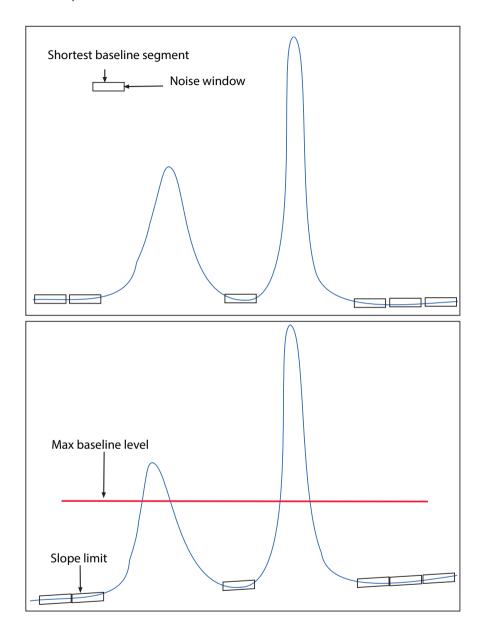
- The curve has no point outside the *Noise window*. The noise window is defined as a
  rectangular corridor parallel to the slope of the curve and centered on the first and
  last points within the currently inspected segment.
- The slope is less than the Slope limit. This limits the maximum slope of the baseline to differentiate baseline segments from peaks.
- The curve parts are lower than the *Max baseline level*. This parameter determines the highest acceptable signal level for the baseline.

The baseline parameters can be illustrated as a rectangular box that the source curve has to fit into in order to be identified as a baseline segment, where:

- The length of the box corresponds to the **Shortest baseline segment**.
- The height of the box corresponds to the maximum level of noise on the baseline segments. This is referred to as the *Noise window*.
- The box is allowed to be tilted with a maximum slope corresponding to the Slope limit.
- The box is not allowed to move up above the *Max baseline level*.

# Baseline parameters - illustration (Classic algorithm)

The illustrations below show the baseline parameters graphically.



# Baseline segment identification (Classic algorithm)

The table below describes the baseline segment identification process:

Stage	Description
-------	-------------

The box is virtually moved along the source curve in steps of one third of the **Shortest baseline segment** length to look for baseline segments.

Stage	Description
2	A baseline segment is found whenever the currently examined part of the source curve fits completely within the box.
3	The found baseline segments are joined by connecting adjacent segments, provided that the slope of the joining lines does not exceed the <b>Slope limit</b> .

#### **Baseline points (Classic algorithm)**

When the baseline segments have been defined and joined, they are replaced by baseline points at the start and end of each segment. The line between these is also filled with points.

**Tip:** The baseline points are shown as gray squares in the **Integrate** →**Edit baseline** function of the Evaluation module.

#### Baseline drawing (Classic algorithm)

The baseline points are used to create the baseline curve using a spline interpolation. The spline function ensures that the baseline curve is guided by the baseline points. However, the curve does not necessarily pass through the baseline points. The baseline will be a smoothly curved function passing close to or through the points.

To reduce the effect of noise at the peak integration, the created baseline is forced equal to the source curve in every position where the difference between the baseline and the source curve is small enough. Choose *Integrate*  $\rightarrow$  *Calculate Baseline*. If the *Accept negative peaks* option is off, the baseline will be forced down to the level of the source curve whenever the created baseline goes above the source curve.

# Measure a Classic algorithm baseline segment

You can try to measure the **Shortest baseline segment** length directly on your chromatogram. To measure the shortest baseline segment:

Step	Action
1	Locate the shortest segment of the curve that you consider a part of the baseline.
2	Right-click in the chromatogram and choose <b>Vertical marker</b> .
3	Use the <b>Vertical marker</b> to measure the length of the segment by reading the start and end point values.
4	Choose Integrate → Calculate Baseline and insert this value as the Shortest baseline segment value.

# Measure noise level (Classic algorithm)

Curve coordinates can also be used to measure noise levels on the source curve. To measure noise levels:

Step	Action	
1	Use the $\it Zoom$ function to focus on a part of the curve that is representative for the baseline noise.	
2	Select an appropriate Y-axis scale.	
3	Measure the Y-axis coordinates.	
4	Calculate the noise range as the difference between the max. and min. values.	
	<ul> <li>Add an extra 20%.</li> </ul>	
	<ul> <li>Choose Integrate → Calculate Baseline and insert this value as the Noise window value.</li> </ul>	

# Measure the slope limit (Classic algorithm)

Follow the instructions to measure the slope at any part of the curve.

Step	Action
1	Select <b>Operations</b> → <b>Differentiate</b> in the Evaluation module.  Result:  The <b>Differentiate</b> dialog opens.
2	<ul> <li>Select the desired source curve.</li> <li>Select the <i>First order</i> calculation option.</li> <li>Click <i>OK</i>.</li> <li>Result: The differentiated curve will appear in the active chromatogram.</li> </ul>
3	Select an appropriate Y-axis scale, right-click and select <b>Vertical marker</b> to measure the Y-axis values for the differentiated curve with the curve coordinates function.  Result:  The Y-axis value is interpreted as the UV curve slope at the selected retention point.
4	• Determine the highest slope value of the baseline (non-peak) part of the curve.

#### Step Action

- Add 10%.
- Select Integrate → Calculate Baseline and use this value as the Slope limit.

**Tip:** If the differentiated curve is very noisy, it can be filtered with a light **Moving** average filter in the **Operations** →**Smooth** function.

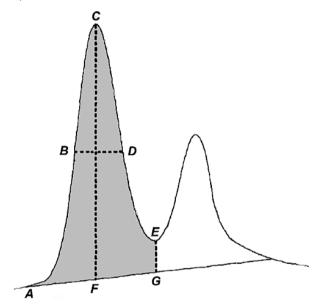
## A.3 Peak table column components

#### Introduction

This section contains a list of peak parameters with explanations and calculation formulae when applicable.

### Peak parameters - illustration

The diagram below illustrates the peak parameters. See the parameter list below for explanations.



### **Peak parameter descriptions**

The list below contains descriptions of the peak parameters.

Parameter	Description
Area	Calculated as the area between the curve and baseline, between the peak start and peak end, time or volume base (gray area in the diagram above).
Asymmetry	Peak asymmetry (indicator of column packing). See definition below this table.
Baseline height	Baseline amplitude at peak start, peak maximum and peak end (A, F and G in the diagram above).

Parameter	Description
Retention factor	The retention factor will only be calculated when the chromatogram is in volume base. The total liquid volume, $V_t$ , must be entered in the <b>Peak Integrate</b> dialog for this parameter to be calculated. See definition below this table.
Fraction tube id	Fraction number at peak start, peak maximum and peak end.
Height	Maximum amplitude above the baseline (C-F in the diagram above).
Kav	Gel phase distribution constant in Size Exclusion Chromatography (SEC). Kav will only be calculated when a SEC column was used and when the chromatogram is in volume base. The void volume, $V_0$ , must be entered in the <b>Peak Integrate</b> dialog for this parameter to be calculated. See definition below this table.
Plate height (HETP)  Note: HETP calculations are not available in UNICORN start. HETP calculations are available in UNICORN 5 and later versions.	Height equivalent to theoretical plate and plates/meter. The column height must be entered in the <b>Peak Integrate</b> dialog for this parameter to be calculated. See definition below this table.
Peak endpoint heights	Amplitude above the baseline at left (A in the diagram above) and right peak limits (E-G in the diagram above).
Peak endpoint retention	Retention value at peak start and peak end, time or volume base (A and G in the diagram above).
Peak name	Name of the peak.
Percent of total area	Peak area as a percent of the total area under the curve above the baseline. Time or volume base.
	Note:
	This value can differ in time and volume base if the flow rate is not constant throughout the method.
Percent of total	Peak area as a percent of the sum of all integrated peaks.
peak area	Note:
	This value can differ in time and volume base if the flow rate is not constant
	throughout the method.

Parameter	Description
Retention	Retention at the peak maximum, time or volume base (C in the diagram above).
Sigma	Standard deviation for a Gaussian-shaped peak. See definition below this table.
Type of peak limits	Identifies the criteria for peak start and peak end as either the baseline intersection or dropline to the baseline or skim line.
Width	Difference in retention between the peak end and peak start, time or volume base (G-A in the diagram above).
Width at half height	Calculated by taking the maximum height of the peak above the baseline, then determining the peak width at half this value above the baseline. Time or volume base. (B-D in the diagram above, where BD bisects CF)

Note:

In the **Options** dialog (which is available in all UNICORN modules from the **Tools** menu) you can select if negative retentions should be displayed or not. The default selection is that negative retention is displayed.

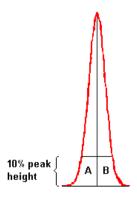
### **Asymmetry formula**

The formula below is used to calculate the Asymmetry.

Asymmetry = B / A

Where:

- A is a partial peak width, measured at a percentage of the peak height (asymmetry ratio), for the leading part of the peak.
- B is a partial peak width, measured at a percentage of the peak height (asymmetry ratio), for the tailing part of the peak.



#### **Change the Asymmetry Ratio**

The **Asymmetry Ratio** is selected in the **Options** dialog. The table below describes how to select a value:

cti	on
C	ti

Choose the **Tools** → **Options** menu item.

Result:

The **Options** dialog opens.

- Type a ratio value in the Asymmetry Ratio at text box.
  - Click OK.

Result:

The ratio value is changed and the dialog closes.

Note:

You must repeat the peak integrations after the change to update the values based on the new asymmetry ratio. The default ratio is 10%.

#### **Retention factor formula**

The formula below is used to calculate the **Retention factor**.

$$\mathbf{k}^{\scriptscriptstyle 1} = \frac{V_R - V_t}{V_t}$$

Where:

- V<sub>R</sub> = retention volume.
- V<sub>t</sub> = total liquid volume.

#### K<sub>av</sub> formula

The formula below is used to calculate **Kav**.

$$k_{av} = \frac{V_R - V_0}{V_C - V_0}$$

Where:

- V<sub>R</sub> = retention volume.
- $V_0$  = void volume.
- V<sub>C</sub> = column volume.

#### **HETP formula**

The formula below is used to calculate the **HETP** value.

HETP = L/N

 $N = 5.54 \times (V_R/W_h)^2$  assuming a Gaussian peak.

Where:

- N = no. of theoretical plates.
- L = bed height in cm.
- V<sub>R</sub> = peak retention volume or time.
- $w_h$  = peak width at half height expressed in the same units as  $V_R$ .

#### **Peak resolution algorithms**

The peak resolution is calculated with one of the following three algorithms:

- 1.  $(V_{R2} V_{R1}) / ((W_{b2} + W_{b1}) / 2)$
- 2.  $(V_{R2} V_{R1}) / ((Sigma_2 + Sigma_1) \times 2)$
- 3.  $((V_{R2} V_{R1}) / (2 \times (W_{h2} + W_{h1}))) / 2.354$

#### Where:

- V<sub>R1</sub>, W<sub>b1</sub>, Sigma<sub>1</sub>, and W<sub>h1</sub> are the retention, width, Sigma, and width at half height of the previous peak.
- V<sub>R2</sub>, W<sub>b2</sub>, Sigma<sub>2</sub>, and W<sub>h2</sub> are the retention, width, Sigma, and width at half height of the current peak.

**Note:** The **Resolution Algorithm** variable in the **Options** dialog determines which of the three algorithms is used. The default setting is algorithm 3.

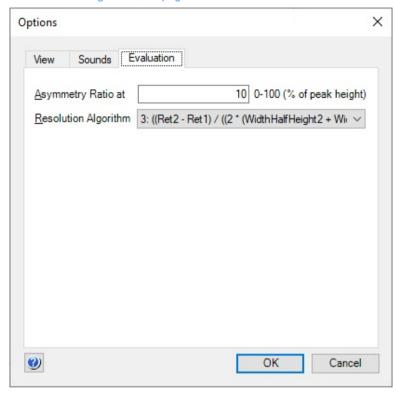
# Change the peak resolution algorithm

Follow the instructions to change the peak resolution algorithm in the **Options** dialog.

Step	Action
1	In the <b>Evaluation</b> module, on the <b>Tools</b> menu, click <b>Options</b> .
	Result:
	The <b>Options</b> dialog opens.
2	Click the <b>Evaluation</b> tab.

#### Step Action

In the **Resolution Algorithm** list, click the desired algorithm as described in Peak resolution algorithms, on page 260.



4 Click **OK**.

Result: The dialog closes and the peak resolution algorithm is changed.

Note:

You must repeat the peak integrations after the change to update the values based on the new algorithm.

### Sigma formula

The formula below is used to calculate **Sigma**.

Sigma = 
$$\sqrt{\frac{\sum_{i=1}^{n} \left(y_i \left(x_i - x_{ymax}\right)^2\right)}{A_{peak}}}$$

Where:

• *n* is the number of data points.

#### A. Evaluation functions and instructions

#### A.3 Peak table column components

- x is the volume or time value.
- $x_{ymax}$  is the volume or time value at the maximum amplitude value.
- $\bullet \quad \mathsf{A}_{\mathsf{peak}} \, \mathsf{is} \, \mathsf{the} \, \mathsf{area} \, \mathsf{of} \, \mathsf{the} \, \mathsf{peak}.$

**Note:** The peak width for a Gaussian peak is (4 x Sigma).

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